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(54) Title: NUCLEIC ACIDS AND PROTEINS RELATED TO ALZHEIMER'S DISEASE, AND USES THEREFOR

(57) Abstract

The present invention describes the identification, isolation, sequencing and characterization of several human genes which interact with the presenilins, mutations in which may lead to Familial Alzheimer's Disease. These presenilin-interacting protein genes may be involved in the pathways which, when affected by mutant presentilins, lead to the development of Alzheimer's Disease. In addition, mutations in the presentilin-interacting protein genes, even in the absence of defects in the presentilins, may be causative of Alzheimer's Disease. Nucleic acids and proteins comprising or derived from the presentlin-interacting proteins are useful in screening and diagnosing Alzheimer's Disease, in identifying and developing therapeutics for treatment of Alzheimer's Disease, and in producing cell lines and transgenic animals useful as models of Alzheimer's Disease.

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NUCLEIC ACIDS AND PROTEINS RELATED TO ALZHEIMER'S DISEASE, AND USES THEREFOR

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Field of the Invention

The present invention relates generally to the field of neurological and physiological dysfunctions associated with Alzheimer's Disease. More particularly, the invention is concerned with the identification, isolation and cloning of genes which are associated with Alzheimer's Disease, as well as their corresponding transcripts and protein products. The present invention also relates to methods for detecting and diagnosing carriers of normal and mutant alleles of these genes, to methods for detecting and diagnosing Alzheimer's Disease, to methods of identifying other genes and proteins related to, or interacting with, the genes and proteins of the invention, to methods of screening for potential therapeutics for Alzheimer's Disease, to methods of treatment for Alzheimer's Disease, and to cell lines and animal models useful in screening for and evaluating potentially useful therapies for Alzheimer's Disease.

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Background of the Invention

Alzheimer's Disease (AD) is a degenerative disorder of the human central nervous system characterized by progressive memory impairment and cognitive and intellectual decline during mid to late adult life (Katzman, 1986). The disease is accompanied by a constellation of neuro-pathologic features principal amongst which are the presence of extracellular amyloid or senile plaques, and neurofibrillary tangles in neurons. The etiology of this disease is complex, although in some families it appears to be inherited as an autosomal dominant trait. Linkage studies have identified three genes associated with the development of AD: β-amyloid precursor protein (APP) (Chartier-Harlin et al., 1991; Goate et al., 1991; Murrell et al., 1991; Karlinsky et al., 1992; Mullan et al., 1992), presenilin-1 (PS-1) (Sherrington, 1995), and presenilin-2 (PS-2) (Rogaev, 1995, and Levy-Lahad, 1995).

The presentiins are multi-spanning membrane proteins which were described in substantial detail in PCT Publication WO96/34099, the entire disclosure of which is incorporated herein by reference. Although the functions of the presentiins are unknown, a number of autosomal dominant presentiin mutations have been identified

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which are strongly associated with the development of early-onset, aggressive, Familial Alzheimer's Disease (FAD).

The present disclosure describes the identification, isolation, sequencing and characterization of several human genes which interact with the presentions, mutations in which may lead to FAD. These present in-interacting protein genes may be involved in the pathways which, when affected by mutant present ins, lead to the development of Alzheimer's Disease. In addition, mutations in the present interacting protein genes, even in the absence of defects in the present ins, may be causative of Alzheimer's Disease.

10 <u>Summary of the Invention</u>

The present invention is based, in part, upon the identification, isolation, sequencing and characterization of several human genes, referred to herein as "presenilin-interacting protein genes" or "PS-interacting protein genes." The products of these genes are believed to interact in vivo with the human presenilin-1 proteins and, therefore, are implicated in the biochemical pathways which are affected in Alzheimer's Disease. Each of these genes, therefore, presents a new therapeutic target for the treatment of Alzheimer's Disease. In addition, PS-interacting protein nucleic acids, PS-interacting proteins and peptides, antibodies to the PS-interacting proteins, cells transformed with PS-interacting protein nucleic acids, and transgenic animals altered with PS-interacting protein nucleic acids, all possess various utilities, as described herein, for the diagnosis, therapy and continued investigation of Alzheimer's Disease and related disorders.

Thus, it is one object of the invention to provide isolated nucleic acids encoding at least a PS-interacting domain of a PS-interacting protein. These PS-interacting proteins include mammalian S5a subunits of the 26S proteasome, the GT24 protein, the p0071 protein, the Rab11 protein, the retinoid X receptor-β, the cytoplasmic chaperonin, and several sequences identified herein as clones Y2H35, Y2H171, and Y2H41. Preferred nucleotide and amino acid sequences are provided herein. It is another object of the invention to provide probes and primers for these

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PS-interacting protein genes, and to provide nucleic acids which encode small antigenic determinants of these genes. Therefore, preferred embodiments include sequences of at least 10, 15 or 20 consecutive nucleotides selected from the disclosed sequences.

Using the nucleic acid sequences and antibodies disclosed and enabled herein, methods for identifying allelic variants or heterospecific homologues of a human PS-interacting protein and gene are provided. The methods may be practiced using nucleic acid hybridization or amplification techniques, immunochemical techniques, or any other technique known in the art. The allelic variants may include other normal human alleles as well as mutant alleles of the PS-interacting protein genes which may be causative of Alzheimer's Disease. The heterospecific homologues may be from other mammalian species, such as mice, rats, dogs, cats or non-human primates, or may be from invertebrate species, such as <u>Drosophila</u> or <u>C. elegans</u>. Thus, it is another object of the invention to provide nucleic acids which encode allelic or heterospecific variants of the disclosed sequences, as well as the allelic or heterospecific proteins encoded by them.

The it another object of the invention to provide vectors, and particularly expression vectors, which include any of the above-described nucleic acids. It is a further object of the invention to provide vectors in which PS-interacting protein nucleic acid sequences are operably joined to exogenous regulatory regions to produce altered patterns of expression, or to exogenous coding regions to produce fusion proteins. Conversely, it is another object to provide nucleic acids in which PS-interacting protein regulatory regions are operably joined to exogenous coding regions, including standard marker genes, to produce constructs in which the regulation of PS-interacting protein genes may be studied and used in assays for therapeutics.

It is another object of the invention to provide host cells and transgenic animals which have been transformed with any of the above-described nucleic acids

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of the invention. The host cells may be prokaryotic or eukaryotic cells and, in particular, may be gametes, zygotes, fetal cells, or stem cells useful in producing transgenic animal models.

In particularly preferred embodiments, the present invention provides a non-human animal model for Alzheimer's Disease, in which the genome of the animal, or an ancestor thereof, has been modified by at least one recombinant construct which has introduced one of the following modifications: (1) insertion of nucleotide sequences encoding at least a functional domain of a heterospecific normal PS-interacting protein, (2) insertion of nucleotide sequences encoding at least a functional domain of a heterospecific mutant PS-interacting protein, (3) insertion of nucleotide sequences encoding at least a functional domain of a conspecific homologue of a heterospecific mutant PS-interacting protein, and (4) inactivation of an endogenous PS-interacting protein gene. Preferred transgenic animal models are rats, mice, hamsters, guinea pigs, rabbits, dogs, cats, goats, sheep, pigs, and non-human primates, but invertebrates are also contemplated for certain utilities.

It is another object of the invention to provide methods for producing at least a functional domain of a PS-interacting protein using the nucleic acids of the invention. In addition, the present invention also provides substantially pure preparations of such proteins, including short peptide sequences for used as immunogens. Thus, the invention provides peptides comprising at least 10 or 15 consecutive amino acid residues from the disclosed and otherwise enabled sequences. The invention further provides substantially pure preparations of peptides which comprise at least a PS-interacting domain of a PS-interacting protein, as well as substantially pure preparations of the entire proteins.

Using the substantially pure peptides and proteins enabled herein, the invention also provides methods for producing antibodies which selectively bind to a PS-interacting protein, as well as cell lines which produce these antibodies.

Another object of the present invention is to provide methods of identifying compounds which may have utility in the treatment of Alzheimer's Disease and related disorders. These methods include methods for identifying compounds which can modulate the expression of a PS-interacting protein gene, methods for identifying compounds which can selectively bind to a PS-interacting protein, and methods of identifying compounds which can modulate activity of a PS-interacting protein. These methods may be conducted in vitro or in vivo, and may employ the transformed cell lines and transgenic animal models of the invention. The methods also may be part of a clinical trial in which compounds identified by the methods of the invention are further tested in human subjects.

It is another object of the invention to provide methods of diagnosing or screening for inherited forms of Alzheimer's Disease by determining if a subject bears a mutant PS-interacting protein gene. Mutant PS-interacting genes may be detected by assays including direct nucleotide sequencing, probe specific hybridization, restriction enzyme digest and mapping, PCR mapping, ligase-mediated PCR detection, RNase protection, electrophoretic mobility shift detection, or chemical mismatch cleavage. Alternatively, mutant forms of a PS-interacting protein may be detected by assays including immunoassays, protease assays, or electrophoretic mobility assays.

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It is also an object of the invention to provide pharmaceutical preparations which may be used in the treatment of Alzheimer's Disease and related disorders which result from aberration in biochemical pathways involving the PS-interacting proteins disclosed and enabled herein. Thus, the present invention also provides pharmaceutical preparations comprising a substantially pure PS-interacting protein, an expression vector operably encoding a PS -interacting protein, an expression vector operably encoding a PS-interacting protein antisense sequence, an antibody which selectively binds to a mutant PS-interacting protein, or an antigenic determinant of a mutant PS-interacting protein. These pharmaceutical preparations may be used to

treat a patient bearing a mutant PS-interacting protein gene which is causative of Alzheimer's Disease or related disorders.

These an other objects of the present invention are described more fully in the following specification and appended claims.

Detailed Description of the Invention

I. Definitions

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In order to facilitate review of the various embodiments of the invention, and an understanding of the various elements and constituents used in making and using the invention, the following definitions are provided for particular terms used in the description and appended claims:

Presenilin. As used without further modification herein, the terms "presenilin" or "presenilins" mean the presenilin-1 (PS1) and/or the presenilin-2 (PS2) genes/proteins. In particular, the unmodified terms "presenilin" or "presenilins" refer to the mammalian PS1 and/or PS2 genes/proteins and, preferably, the human PS1 and/or PS2 genes/proteins as described and disclosed in PCT Publication WO96/34099.

Normal. As used herein with respect to genes, the term "normal" refers to a gene which encodes a normal protein. As used herein with respect to proteins, the term "normal" means a protein which performs its usual or normal physiological role and which is not associated with, or causative of, a pathogenic condition or state. Therefore, as used herein, the term "normal" is essentially synonymous with the usual meaning of the phrase "wild type." For any given gene, or corresponding protein, a multiplicity of normal allelic variants may exist, none of which is associated with the development of a pathogenic condition or state. Such normal allelic variants include, but are not limited to, variants in which one or more nucleotide substitutions do not result in a change in the encoded amino acid sequence.

Mutant. As used herein with respect to genes, the term "mutant" refers to a gene which encodes a mutant protein. As used herein with respect to proteins, the term "mutant" means a protein which does not perform its usual or normal

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physiological role and which is associated with, or causative of, a pathogenic condition or state. Therefore, as used herein, the term "mutant" is essentially synonymous with the terms "dysfunctional," "pathogenic," "disease-causing," and "deleterious." With respect to the presenilin and presenilin-interacting protein genes and proteins of the present invention, the term "mutant" refers to genes/proteins bearing one or more nucleotide/amino acid substitutions, insertions and/or deletions which typically lead to the development of the symptoms of Alzheimer's Disease and/or other relevant inheritable phenotypes (e.g. cerebral hemorrhage, mental retardation, schizophrenia, psychosis, and depression) when expressed in humans. This definition is understood to include the various mutations that naturally exist, including but not limited to those disclosed herein, as well as synthetic or recombinant mutations produced by human intervention. The term "mutant," as applied to these genes, is not intended to embrace sequence variants which, due to the degeneracy of the genetic code, encode proteins identical to the normal sequences disclosed or otherwise enabled herein; nor is it intended to embrace sequence variants which, although they encode different proteins, encode proteins which are functionally equivalent to normal proteins.

Substantially pure. As used herein with respect to proteins (including antibodies) or other preparations, the term "substantially pure" means that the preparation is essentially free of other substances to an extent practical and appropriate for its intended use. In particular, a protein preparation is substantially pure if it is sufficiently free from other biological constituents so as to be useful in, for example, generating antibodies, sequencing, or producing pharmaceutical preparations. By techniques well known in the art, substantially pure proteins or peptides may be produced in light of the nucleic acid and amino acid sequences disclosed herein. In particular, in light of the nucleic acid and amino acid sequences disclosed herein, one of ordinary skill in the art may, by application or serial application of well-known methods including HPLC or immuno-affinity chromatography or electrophoretic separation, obtain proteins or peptides of any generally feasible purity. Preferably, but not necessarily, "substantially pure"

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preparations include at least 60% by weight (dry weight) the compound of interest. More preferably the preparation is at least 75% or 90%, and most preferably at least 99%, by weight the compound of interest. Purity can be measured by any appropriate method, e.g., column chromatography, gel electrophoresis, or HPLC analysis. With respect to proteins, including antibodies, if a preparation includes two or more different compounds of interest (e.g., two or more different antibodies, immunogens. functional domains, or other polypeptides of the invention), a "substantially pure" preparation is preferably one in which the total weight (dry weight) of all the compounds of interest is at least 60% of the total dry weight. Similarly, for such 10 preparations containing two or more compounds of interest, it is preferred that the total weight of the compounds of interest be at least 75%, more preferably at least 90%, and most preferably at least 99%, of the total dry weight of the preparation. Finally, in the event that the protein of interest is mixed with one or more other proteins (e.g., serum albumin) or compounds (e.g., diluents, excipients, salts, polysaccharides, sugars, lipids) for purposes of administration, stability, storage, and the like, such other proteins or compounds may be ignored in calculation of the purity of the preparation.

Isolated nucleic acid. As used herein, an "isolated nucleic acid" is a ribonucleic acid, deoxyribonucleic acid, or nucleic acid analog comprising a polynucleotide sequence that is isolated or separate from sequences that are immediately contiguous (one on the 5' end and one on the 3' end) in the naturally occurring genome of the organism from which it is derived. The term therefore includes, for example, a recombinant nucleic acid which is incorporated into a vector, into an autonomously replicating plasmid or virus, or into the genomic DNA of a prokaryote or eukaryote; or which exists as a separate molecule (e.g., a cDNA or a genomic DNA fragment produced by PCR or restriction endonuclease treatment) independent of other sequences. It also includes a recombinant DNA which is part of a hybrid gene encoding additional polypeptide sequences and/or including exogenous regulatory elements.

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Substantially identical sequence. As used herein, a "substantially identical" amino acid sequence is an amino acid sequence which differs only by conservative amino acid substitutions, for example, substitution of one amino acid for another of the same class (e.g., valine for glycine, arginine for lysine, etc.) or by one or more non-conservative substitutions, deletions, or insertions located at positions of the amino acid sequence which do not destroy the function of the protein (assayed, e.g., as described herein). Preferably, such a sequence is at least 85%, more preferably 90%, and most preferably 95% identical at the amino acid level to the sequence of the protein or peptide to which it is being compared. For nucleic acids, the length of comparison sequences will generally be at least 50 nucleotides, preferably at least 60 nucleotides, more preferably at least 75 nucleotides, and most preferably 110 nucleotides. A "substantially identical" nucleic acid sequence codes for a substantially identical amino acid sequence as defined above.

Transformed cell. As used herein, a "transformed cell" is a cell into which (or into an ancestor of which) has been introduced, by means of recombinant DNA techniques, a nucleic acid molecule of interest. The nucleic acid of interest will typically encode a peptide or protein. The transformed cell may express the sequence of interest or may be used only to propagate the sequence. The term "transformed" may be used herein to embrace any method of introducing exogenous nucleic acids including, but not limited to, transformation, transfection, electroporation, microinjection, viral-mediated transfection, and the like.

Operably joined. As used herein, a coding sequence and a regulatory region are said to be "operably joined" when they are covalently linked in such a way as to place the expression or transcription of the coding sequence under the influence or control of the regulatory region. If it is desired that the coding sequences be translated into a functional protein, two DNA sequences are said to be operably joined if induction of promoter function results in the transcription of the coding sequence and if the nature of the linkage between the two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the regulatory region to direct the transcription of the coding sequences, or (3) interfere with the ability of the

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corresponding RNA transcript to be translated into a protein. Thus, a regulatory region would be operably joined to a coding sequence if the regulatory region were capable of effecting transcription of that DNA sequence such that the resulting transcript might be translated into the desired protein or polypeptide.

Stringent hybridization conditions. Stringent hybridization conditions is a term of art understood by those of ordinary skill in the art. For any given nucleic acid sequence, stringent hybridization conditions are those conditions of temperature, chaotrophic acids, buffer, and ionic strength which will permit hybridization of that nucleic acid sequence to its complementary sequence and not to substantially different sequences. The exact conditions which constitute "stringent" conditions, depend upon the nature of the nucleic acid sequence, the length of the sequence, and the frequency of occurrence of subsets of that sequence within other non-identical sequences. By varying hybridization conditions from a level of stringency at which non-specific hybridization occurs to a level at which only specific hybridization is observed, one of ordinary skill in the art can, without undue experimentation, determine conditions which will allow a given sequence to hybridize only with complementary sequences. Suitable ranges of such stringency conditions are described in Krause and Aaronson (1991). Hybridization conditions, depending upon the length and commonality of a sequence, may include temperatures of 20°C-65°C and ionic strengths from 5x to 0.1x SSC. Highly stringent hybridization conditions may include temperatures as low as 40-42°C (when denaturants such as formamide are included) or up to 60-65°C in ionic strengths as low as 0.1x SSC. These ranges, however, are only illustrative and, depending upon the nature of the target sequence, and possible future technological developments, may be more stringent than necessary. Less than stringent conditions are employed to isolate nucleic acid sequences which are substantially similar, allelic or homologous to any given sequence.

<u>Selectively binds.</u> As used herein with respect to antibodies, an antibody is said to "selectively bind" to a target if the antibody recognizes and binds the target of interest but does not substantially recognize and bind other molecules in a sample, e.g., a biological sample, which includes the target of interest. That is, the antibody

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must bind to its target with sufficient specificity so as to distinguish the target from essentially all of molecules which would reasonably be present in a biological sample including the target.

II. The Presentlins and Presentlin-Interacting Proteins

The present invention is based, in part, upon the discovery of a family of mammalian genes which, when mutated, are associated with the development of Alzheimer's Disease. The discovery of these genes, designated presenilin-1 (PS1) and presenilin-2 (PS2), as well as the characterization of these genes, their protein products, mutants, invertebrate homologues, and possible functional roles, are described in PCT Publication WO96/34099. The present invention is further based, in part, upon the discovery of a group of proteins which interact with the presenilins under physiological conditions and which, therefore, are believed to be involved in the biochemical pathways which are altered in Alzheimer's Disease. These proteins are referred to herein as presenilin-interacting (PS-interacting) proteins. Because mutations in the presenilins are known to be causative of Alzheimer's Disease, each of the PS-interacting genes and proteins disclosed and described herein presents a novel target for therapeutic intervention in Alzheimer's Disease. That is, modulation of the interactions of these proteins with the presenilins, or modulation of the interactions of at least the PS-interacting domains of these PS-interacting proteins with at least the interacting domains of the presenilins, provides a means of modulating the activity and/or availability of the presenilins, or of modulating the activity and/or availability of the PS-interacting proteins. Furthermore, as aberrations in the interactions of mutant presenilins with one or more of these PS-interacting proteins is causative of Alzheimer's Disease, mutations in one or more of these PS-interacting proteins are also likely to be causative of Alzheimer's Disease. Therefore, each of the PSinteracting genes and proteins disclosed and described herein presents a novel target for diagnosis of forms of familial and/or sporadic Alzheimer's Disease with an etiology independent of mutations in the presenilins. Finally, as described more fully below, the PS-interacting genes and proteins described and disclosed herein provide for new assays for compounds which affect the interactions of the presenilins and PS-

interacting proteins, assays for other members of the biochemical pathways involved in the etiology of Alzheimer's Disease, and new cell lines and transgenic animal models for use in such assays.

5 1. Presentlin Processing

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Employing the antibodies and protein-binding assays described and/or enabled in PCT Publication WO96/34099, the processing and protein-protein interactions of both normal and mutant presentlins were investigated. It was found that mutations in the presentlins appear to lead to changes in both their intracellular processing (e.g., endoproteolytic cleavage, ubiquitination, and clearance) and their intracellular interactions with other proteins expressed in human brain. As described below, knowledge of presentlin processing and interactions, and particularly changes in mutant presentlin processing and interactions, provides for new diagnostic and therapeutic targets for Alzheimer's Disease and related disorders.

Western blot analysis suggests that the normal presenilins undergo proteolytic cleavage to yield characteristic N- and C-terminal fragments. As noted above, the normal presentiin proteins have an expected molecular mass of 47-51 kDa depending, in part, upon mRNA splice variations, electrophoretic conditions, etc. Analysis of Western blots suggests, however, that the normal presenilin proteins undergo proteolytic cleavage to yield an approximately 35 kDa N-terminal fragment and an approximately 18 kDa C-terminal fragment. In particular, Western blots bearing lysates from wild-type native human fibroblasts, human neocortical brain tissue from control subjects, and neocortical brain tissue from non-transgenic and PS1 transgenic mice using antibodies ("14.2") recognizing PS1-specific residues 1-25 at the N-terminus reveal the presence of a strong immunoreactive band of approximately 35 kDa and, after longer exposures, a weaker band of approximately 45 kDa which presumably represents the full-length PS1 protein. Antibodies ("520") directed at residues 304-318 at the apex of the TM6-7 loop of PS1, and antibodies ("4627") directed at residues 457-467 in the C-terminus of PS1, both recognize the same strong band of approximately 18 kDa. Antibodies 520 also recognize a weak band of 45 kDa

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coincident with the PS1 band detected by 14.2. Sequencing of the major C-terminal fragment from PS1-transfected human embryonic kidney cells (HEK 293) showed that the principal endoproteolytic cleavage occurs near M298 in the proximal portion of the TM6 \rightarrow 7 loop, possibly by enzymes other than the proteasome. These observations suggest that an endoproteolytic cleavage event occurs near the junction of exons 9 and 10 of PS1. Full length PS1 in these cells is quickly turned over ($t_{1/2}$ < 60 min.) by the proteasome.

To determine whether mutations in the presentiin proteins result in alterations of their proteolytic cleavage, Western blots containing lysates of fibroblast and neocortical brain homogenates from normal subjects and subjects carrying PS1 mutations were investigated with the PS1 specific antibody Ab 14.2. In fibroblasts, there were no obvious differences in the relative intensities of the protein bands when lysates from heterozygous carriers of the PS1 mutations were compared with normal homozygotes. In contrast, there appeared to be a difference between PS1 mutation carriers and normals in homogenates of temporal neocortex from AD affected heterozygous carriers of either the PS1 A246E or C410Y mutations (which are located in TM6 and TM7 respectively). In heterozygotes, a strongly immunoreactive band of approximately 45 kDa was detected which initially appeared to correspond to the fulllength PS1 protein. Further analysis, however, revealed that this band represents an alternatively processed presenilin product. A similar band corresponding to this mutant processed PS1 was observed in neocortical homogenates from some sporadic late-onset AD patients. These data suggest that (1) some pathogenic PS1 mutations associated with early-onset AD alter the way in which the presenilins are processed through endoproteolytic and proteasome pathways and (2) the presenilin proteins, and changes in the processing of the presenilins in the brain, are also implicated in lateonset and sporadic AD.

2. Presenilin-Interacting Proteins

In order to identify proteins which may bind to or otherwise interact with the presentilins in vivo, a yeast two-hybrid system was used as described below

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(Example 1). In particular, because mutations in the TM6→7 loop domains are known to be causative of AD, a yeast two-hybrid system was used to identify cellular proteins which may interact with normal and mutant presentilin TM6→7 loop domains. Yeast two-hybrid studies were also done with cDNAs corresponding to the C-terminal 18 kDa endoproteolytic cleavage fragment, and with cDNAs corresponding to the TM1→2 intraluminal loop domain, which is also the site of the FAD associated Y115H missense mutation. In brief, cDNA sequences encoding the TM6→7 loop (i.e., residues 266 to 409 of PS1) were ligated in-frame to the GAL4 DNA-binding domain in the pAS2-1 yeast expression plasmid vector (Clontech). This plasmid was then co-transformed into S. cerevisiae strain Y190 together with a 10 library of human brain cDNAs ligated into the pACT2 yeast expression vector bearing the GAL4 activation domain (Clontech). After appropriate selection and re-screening, a number of clones were recovered and sequenced bearing human brain cDNAs encoding peptides which interacted with the normal presentilin TM6→7 domain. To 15 determine whether these presenilin interactions would be modified by AD related mutations within the TM6→7 loop, the yeast two-hybrid system was again used with TM6→7 loop peptides containing the L286V, the L392V, and the exon 10 splicing mutants. When these mutant constructs were used as "bait" to re-screen the brain cDNA:GAL4 activation domain library, some but not all of the brain cDNA 20 sequences which interacted with the normal presenilin were recovered. In addition, several new clones were identified which interacted with the mutant but not the normal presenilins. The clones corresponding to the PS-interacting proteins with the highest presenilin affinity are described in Example 1 and below.

PS-interacting proteins, particularly those which interact selectively with either the normal or mutant presenilins, provide new targets for the identification of useful pharmaceuticals, new targets for diagnostic tools in the identification of individuals at risk, new sequences for the production of transformed cell lines and transgenic animal models, and new bases for therapeutic intervention in Alzheimer's Disease. In particular, the onset of AD may be associated with aberrant interactions between mutant presentin proteins and normal forms of PS-interacting proteins such

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as those identified using the methods described herein. These changes may increase or decrease interactions present with normal PS1 or may cause interaction with a novel mutation-specific PS-interacting protein. In addition, however, aberrant interactions may result from normal presentions binding to mutant forms of the PS-interacting proteins and, therefore, mutations in the PS-interacting proteins may also be causative of AD.

A. The S5a Subunit of the 26S Proteasome

Two overlapping clones have been identified as representing a portion of the human protein alternatively known as Antisecretory Factor ("ASF") or the Multiubiquitin chain-binding S5a subunit of the 26S proteasome ("S5a"). These clones, which together include residues 70-377 of S5a, were shown to interact with the normal presenilin TM6 \rightarrow 7 loop domain but only weakly with two TM6 \rightarrow 7 loop domain mutants tested (L286V, L392V). The PS1:S5a interaction was confirmed by co-immunoprecipitation studies, and immunocytochemical studies showed S5a and PS1 are expressed in contiguous intracellular compartments in brain cells typically affected by AD.

The interaction between PS1 and the proteasome could be relevant to the pathogenesis of Alzheimer's Disease (AD) through several possible mechanisms. First, most mammalian cells seem to maintain very low levels of the PS1 holoprotein. A notable exception to this are cells expressing the PS1 Δ290-319 splicing mutation, which results in a mutant PS1 holoprotein which is not endoproteolytically cleaved and which is, therefore, readily detectable. In the case of the Δ290-319 splicing mutation at least, the presence of the mutant PS1 holoprotein, or the absence or reduction in the 35 kDa N-terminal and 18 kDa C-terminal fragments, appears sufficient to cause AD. It is possible, therefore, that even very subtle changes in the turnover of the mutant PS1 holoprotein might have significant pathophysiological effects. Thus, mutations in either the presenilins or S5a which perturb the PS1:S5a interaction in the mammalian CNS may cause the presenilin holoprotein to be aberrantly processed and cause AD. Therefore, modulation of presenilin proteolytic pathways might be applied therapeutically to enhance removal of mutant holoprotein.

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To assess a potential in vivo relationship between PS1 and the S5a subunit of the 26S proteasome, the effects of proteasome inhibitors on PS1 metabolism were investigated. Short term organotypic cultures of neonatal rat hippocampus and carcinoma of colon (CaCo2) cells (which express high levels of both PS1 and PS2) were administered either the specific, reversible proteasome inhibitor N-acetylleucinyl-leucinyl-norleucinyl-H (LLnL) (Rock et al., 1994), or the specific irreversible proteasome inhibitor lactacystin (Fenteany et al., 1995). Both agents caused an increase in the steady state levels of PS1 holoprotein. Both agents also prolonged the half-life of the PS1 holoprotein in pulse chase experiments in hippocampal slices from ~15 minutes to ~35 minutes. As noted above, the PS1 holoprotein appears to be rapidly turned over in normal cells. However, even after four hours of metabolic labelling, neither of the proteasome inhibitors affected the level of the 35 kDa Nterminal PS1 fragment, or resulted in the appearance of novel species. These studies imply that the majority of the PS1 holoprotein is catabolized directly via a rapid. proteasome dependent pathway in a manner similar to several other integral membrane proteins (e.g. Sec61 and CFTR). On the other hand, because the ~35 kDa and ~ 18 kDa terminal fragments are still produced in the presence of proteasome inhibitors, this endoproteolytic cleavage of PSI is probably not mediated by the proteasome pathway. Therefore, it appears that at least two proteolytic pathways act upon the PS1 holoprotein.

An alternate possibility is that mutant PS1:S5a interactions may modify the function or the cellular regulation of S5a. To address this possibility, S5a levels were examined by Western blotting of lysates from postmortem temporal neocortex from non-AD neurologic controls (n = 8), sporadic AD (n = 8) and PS1-linked FAD (n = 4). In the majority of non-AD brains, polyclonal anti-S5a antibodies specifically detected an S5a species with Mr of ~ 50 kDa, which could be abolished by preabsorption of the antibody with recombinant His₆-S5a or with extracts of myc-S5a transfected cells. In a subset of these control cases an additional S5a reactive band was observed at ~ 34 kDa. In contrast, in tissue from all subjects with sporadic late onset AD, the predominant S5a reactive species was observed at ~ 40 kDa which was

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not seen in control tissue. The origin, and the functional significance of this altered electrophoretic mobility is unclear but indicates that S5a processing is altered in AD brains, irrespective of whether the AD is presentillin-linked or sporadic.

Thus, the presenilin-proteasome interaction appears significant in several respects. First, the facts that the normal presenilin TM6→7 loop domain interacts 5 with the S5a protein, that the mutant presenilin TM6→7 loop domains fail to interact (or interact very weakly) with the S5a protein, that presenilins bearing mutations in the TM6→7 loop domain appear to be differently cleaved and multiubiquitinated, that proteasomes are known to be involved in the cleavage and clearance of a variety of proteins (particularly multiubiquitinated proteins), that inhibition of proteasome activity inhibits cleavage of the presenilin holoproteins, and that S5a processing is altered in AD brains, all suggest (1) that the S5a subunit and the 26S proteasome are involved in the normal processing of the presenilins and that mutations which disrupt this normal interaction may be responsible for the abnormal processing observed in TM6→7 loop domain mutants; or (2) that the presentilin-proteasome interaction may modulate the activity of PS1, S5a, or both, with or without involving proteasomemediated presenilin processing; or (3) that modulation of the normal quality control function of proteasome-mediated degradation of misfolded or mutant membrane proteins trafficking through the ER and Golgi (such as APP, Notch, or Prion proteins), and of misfolded, mutant, or ubiquitinated cytoplasmic proteins (including structural proteins such as tau, and short lived, proteasome processed signaling molecules such as NFkB). Thus, defective proteasome function might selectively cause these proteins (especially β APP, tau, Prion) to be aberrantly metabolized. The latter would lead to the accumulation of neurotoxic, amyloidogenic protease-resistant derivatives such as Aβ and PrPsc, the accumulation of neurofibrillary tangles, and defective intracellular signaling functions. In support of these hypotheses, it should be noted that failure to clear hyperubiquitinated phosphorylated tau and other microtubule associated proteins is a prominent feature of Alzheimer's Disease (Kosik and Greenberg, 1994), suggesting a possible link between TM6→7 loop domain mutants, presentlinproteasome interactions, tau-proteasome interactions, and the neurofibrillary tangles

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of tau protein in AD brains. Finally, proteasomes are known to be capable of degrading APP and of binding the A β peptides which are associated with Alzheimer's Disease, suggesting a possible link between TM6 \rightarrow 7 loop domain mutants, presentilin-proteasome interactions, APP-proteasome interactions, and the amyloid plaques characteristic of AD brains. Furthermore, administration of proteasome inhibitors such as LLnL and Lactacystin cause severe disturbances in β APP metabolism with increases in intracellular immature N-glycosylated β APP, and the secretion of much larger amounts of A β 42 isoforms into the media (Klafki, et al., 1996).

Therefore, presentilin processing and the presentilin-proteasome interaction are clear targets for the diagnosis as well as therapeutic intervention in AD. Thus, as described below, assays may now be provided for drugs which affect the proteasome-mediated cleavage of the presentilins, which affect the alternative endoproteolytic cleavage and ubiquitination of the mutant presentilins, or which otherwise affect the processing and trafficking of the presentilins or the S5a subunit of the proteasome. In addition, as mutations in the 26S proteasome which disrupt the normal processing of the presentilins are likely to be causative of Alzheimer's Disease, additional diagnostic assays are provided for detecting mutations in the S5a or other subunits of the proteasome. Finally, additional transformed cell lines and transgenic models may now be provided which have been altered by the introduction of a normal or mutant sequence encoding at least a functional domain of the proteasome. The appearance of abnormal electrophoretic forms of S5a (and/or other proteasome subunits) in biologic tissues and fluids can be used as a clinical test for diagnosis and monitoring of disease activity in subjects with sporadic forms of AD.

B. GT24: A Protein with "Armadillo" Repeats

Another PS-interacting protein, designated GT24, was identified from several over-lapping clones obtained using a PS1₂₆₆₋₄₀₉ domain as bait in the yeast two-hybrid system and a human adult brain cDNA library. Six longer GT24 clones of ~3.8 kb in size were subsequently obtained by screening of conventional cDNA libraries. The open reading frame within the longest GT24 clone obtained to date

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(Accession number U81004) suggests that GT24 is a protein of at least 1040 amino acids with a unique N-terminus, and considerable homology to several armadillo (arm) repeat proteins at its C-terminus. Thus, for example, residues 440-862 of GT24 (numbering from Accession number U81004) have 32-56% identity (p=1.2e⁻¹³³) to residues 440-854 of murine p120 protein (Accession number Z17804), and residues 367-815 of GT24 have 26-42% identity (p=0.0017) to residues 245-465 of the D. melanogaster armadillo segment polarity protein (Accession number P18824). The GT24 gene maps to chromosome 5p15 near the anonymous microsatellite marker D5S748 and the Cri-du-Chat syndrome locus.

Hybridization of unique 5' sequences of GT24 to Northern blots reveals that the GT24 gene is expressed as a range of transcripts varying in size between ~3.9 and 5.0 kb in several regions of human brain, and in several non-neurologic tissues such as heart. In addition, in situ hybridization studies using a 289 bp single copy fragment from the 5' end of GT24 in four month old murine brain reveal GT24 transcription closely parallels that of PS1, with robust expression in dentate and hippocampal neurons, in scattered neocortical neurons, and in cerebellar Purkinje cells. In day E13 murine embryos, GT24 is widely expressed at low levels, but is expressed at somewhat higher levels in somites and in the neural tube. A physiological in vivo interaction between GT24 and PS1 is supported by coimmunoprecipitation studies in HEK293 cells transiently transfected with a wild type human PS1 cDNA, a c-myc-tagged cDNA encoding residues 484-1040 of GT24 (including the C-terminal arm repeats), or both cDNAs. Cell lysates were immunoprecipitated with anti-PS1 antibodies and then investigated for the presence of the myc-GT24 protein by immuno-blotting. In PS1/myc-GT24 double transfected cells, the immunoprecipitates contained a robust anti- \underline{myc} reactive band of Mr \sim 60 kDa, which co-migrated with a myc-GT24 control. In cells transfected with myc-GT24 only, a very weak band was detected after long exposures, presumably reflecting interaction of the myc-GT24 with low levels of endogenous PS1. No mycreactive bands were detected in cells transfected with PS1 alone, or in any of the transfected cells immunoprecipitated with pre-immune serum. Taken together, these

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observations strongly suggest that the observed PS1:GT24 interaction is physiologically relevant.

To explore whether mutations in the TM6-TM7 loop of PS1 might influence the PS1:GT24 interaction, we employed quantitative liquid β-galactosidase assays to directly compare the yeast-two-hybrid interaction of the C-terminal residues 499-1040 of GT24 with wildtype and mutant PS1₂₆₆₋₄₀₉. These studies revealed that the interaction of GT24₄₉₉₋₁₀₄₀ with a L286V mutant PS1 domain was not significantly different from the interaction with the corresponding wild type PS1 domain. In contrast, there was a significant reduction in the GT24₄₉₉₋₁₀₄₀ interaction with the L392V mutant PS1 construct. The absence of an effect of the L286V mutation, and the presence of an effect with the L392V mutation, may suggest that some mutations may effect PS1:GT24 binding, while others may modulate the PS1 response to GT24 binding.

The PS1:GT24 interaction could support several functions. The <u>arm</u> repeat motif of GT24 has been detected in several proteins with diverse functions including β-catenin and its invertebrate homologue <u>armadillo</u>, plakoglobin, p120, the adenomatous polyposis coli (APC) gene, suppressor of RNA polymerase 1 in yeast (SRP1), and smGDS. For example, β-catenin, p120 and plakoglobin play an essential role in intercellular adhesion. β-catenin/<u>armadillo</u> is involved in transduction of <u>wingless/Wnt</u> signals during cell fate specification, and β-catenin and p120 may play a role in other receptor mediated signal transduction events including responses to trophic factors such as PDGF, EGF, CSF-1 and NGF.

If the PS1:GT24 interaction is part of intercellular signaling pathways for trophic factors, or is involved in cell-cell adherence, disruption of the interaction may be involved in the neurodegenerative processes in PS-linked FAD brains, and in the increased sensitivity of PS1 or PS2 transfected cells to apoptosis (Wolozin et al., 1996). It is of note that at least one arm protein, smGDS, stimulates GDP/GTP exchange on intracellular G-proteins (Kikuchi et al. 1992; Borguski et al., 1993), and that mutant forms of both βAPP and PS2 are thought to activate programmed cell

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death pathways through mechanisms involving heterotrimeric GTP/GDP proteins (Wolozin et al, 1996; Okamoto, et al., 1995; Yamatsuji, et al, 1996).

The interaction between PS1 and GT24 may also be involved in some of the developmental phenotypes associated with homozygous PS1 knockouts in mice such as failed somitogenesis of the caudal embryo, short tail, and fatal cerebral hemorrhage at around day E13.5 (Wong et al., 1996). The resemblance of these skeletal phenotypes to those associated with null mutations in PAX1 and Notch, and the apparent suppressor effect of mutations in sel12 on Notch/lin12 mediated signaling in C. elegans suggest that the PS proteins function in the Notch signaling pathway. In addition, mice homozygous for a knockout of the Wnt-3a gene (Takada et al., 1994), and murine homozygotes for a spontaneous mutation, "vestigial tail" or vt, in the Wnt-3a gene (Greco et al., 1996), have skeletal phenotypes of defective caudal somite and tail bud formation. The Wnt-3a knockouts are embryonic lethal by day 12.5. These phenotypes are similar to those of homozygous knockouts of the murine PS1 gene (Wong et al., 1996). The observation that GT24 binds to PS1, is expressed in embryonic somites, and contains the armadillo repeat motif of other proteins used in the downstream signaling in the Wingless/Wnt pathway suggests that PS1 is a downstream element in the GT24-Wingless/Wnt pathway. This can be exploited to create a bioassay for drugs affecting the GT24-PS1 interaction directly, or affecting upstream or downstream components of that interaction pathway, and can therefore be used to monitor the effects of presenilin mutations. For example, cells transfected with normal or mutant presenilins may be exposed to soluble Wnt-3a protein (or other Wnt proteins such as Wnt-1) and assayed for changes which are specific to the Wingless/Wnt signaling pathway, or for any of the other changes described herein for cell assays (e.g., intracellular ion levels, $A\beta$ processing, apoptosis, etc.).

Thus, the GT24 protein also presents new targets for diagnosis as well as therapeutic intervention in AD. For example, as mutations in the GT24 protein may also be causative of Alzheimer's Disease, additional diagnostic assays are provided for detecting mutations in these sequences. Similarly, additional transformed cell lines

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and transgenic models may now be provided which have been altered by introduction of a normal or mutant nucleic acid encoding at least a functional domain of the GT24 protein, and particularly the functional domains (e.g., residues 70-377) which interact with the presenilins. Such transformed cells and transgenics will have utility in assays for compounds which modulate the presenilin-GT24 interactions.

C. p0071: A Protein with "Armadillo" Repeats

Another independent clone isolated in the initial screening with the wild type PS1₂₆₆₋₄₀₉ "bait" also encodes a peptide with C-terminal arm repeats (clone Y2H25, Accession number U81005). A longer cDNA sequence corresponding to the Y2H25 clone has been deposited with GenBank as human protein p0071 (Accession number X81889) and is reproduced herein as SEQ ID NO: 5. Clone Y2H25 corresponds essentially to nucleotide positions 1682-1994 of SEO ID NO: 5. Comparison of the predicted sequence of the Y2H25/p0071 ORF with that of GT24 confirms that they are related proteins with 47% overall amino acid sequence identity. and with 70% identity between residues 346-862 of GT24 and residues 509-1022 of p0071. This suggests that PS1 interacts with a novel class of arm repeat containing proteins. The broad ~4.5 kb hybridization signal obtained on Northern blots with the unique 5' end of GT24 could reflect either alternative splicing/polyadenylation of GT24 or, less likely, the existence of additional members of this family with higher degrees of N-terminal homology to GT24 than p0071. Cells transformed with these sequences, or transgenic animals including these sequences, will have additional utility as animal models of AD and for use in screening for compounds which modulate the action of normal and mutant presentlins.

D. Rab 11

One clone (Y2H9), disclosed herein as SEQ ID NO: 5, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a known gene, Rab11, available through Accession numbers X56740 and X53143.

Rab11 is believed to be involved in protein/vesicle trafficking in the ER/Golgi. Note the possible relationship to processing of membrane proteins such as βAPP and Notch

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with resultant overproduction of toxic AB peptides (especially neurotoxic AB₁₋₄₂₍₄₃₎ isoforms) (Scheuner, et al, 1995).

E. Retinoid X Receptor-β

One clone (Y2H23b), disclosed herein as SEQ ID NO: 6, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a known gene, known variously as the retinoid X receptor-β, nuclear receptor coregulator, or MHC Class I regulatory element, and is available through Accession numbers M84820, X63522 and M81766. This gene is believed to be involved in intercellular signaling, suggesting a possible relationship to the intercellular signaling function mediated by C. elegans sel12 and Notch/lin-12 (transcription activator).

F. Cytoplasmic Chaperonin

One clone (Y2H27), disclosed herein as SEQ ID NO: 8, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a known gene, a cytoplasmic chaperonin containing TCP-1, available through Accession numbers U17104 and X74801.

G. Clone Y2H35

One clone (Y2H35), disclosed herein as SEQ ID NO: 7, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a sequence that codes for a protein of unknown function, available through Accession number R12984, but which displays evolutionary conservation in yeast sequences.

H. Clone Y2H171

One clone (Y2H171), disclosed herein as SEQ ID NO: 9, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a known expressed repeat sequence available through Accession number D55326.

I. <u>Clone Y2H41</u>

One clone (Y2H41) was identified which reacts strongly with the $TM6 \rightarrow 7$ loop domains of both PS1 and PS2 as well as the mutant loop domains of PS1. The sequence, disclosed as SEQ ID NO: 10, shows strong homology to an EST of unknown function (Accession number T64843).

III. Preferred Embodiments

Based, in part, upon the discoveries disclosed and described herein, the following preferred embodiments of the present invention are provided.

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1. Isolated Nucleic Acids

In one series of embodiments, the present invention provides isolated nucleic acids corresponding to, or relating to, the nucleic acid sequences disclosed herein, which encode at least the PS-interacting domain of a PS-interacting protein. As described more fully below, the disclosed and enabled sequences include normal sequences from humans and other mammalian species, mutant sequences from humans and other mammalian species, homologous sequences from non-mammalian species such as Drosophila and C. elegans, subsets of these sequences useful as probes and PCR primers, subsets of these sequences encoding fragments of the PS-interacting proteins or corresponding to particular structural domains or polymorphic regions, complementary or antisense sequences corresponding to fragments of the PS-interacting protein genes, sequences in which the PS-interacting protein coding regions have been operably joined to exogenous regulatory regions, and sequences encoding fusion proteins in which portions of the PS-interacting proteins are fused to other proteins useful as markers of expression, as "tags" for purification, or in screens and assays for other proteins which interact with the PS-interacting proteins.

Thus, in a first series of embodiments, isolated nucleic acid sequences are provided which encode at least a PS-interacting domain of a normal or mutant version of a PS-interacting protein. Examples of such nucleic acid sequences are disclosed herein as SEQ ID NOs: 1, 3, and 5-10. In addition, given the sequences of the PS-interacting domains of the PS-interacting proteins disclosed herein, one of ordinary skill in the art is clearly enabled to obtain the entire genomic or cDNA sequence encoding the entire PS-interacting proteins. Thus, for example, based upon the initial clone of the GT24 protein obtained using the yeast two-hybrid system (Example 1), the larger GT24 clone disclosed as SEQ ID NO: 3 was obtained by standard methods

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known in the art. Complete cDNA or genomic clones of each of the genes encoding the disclosed sequences may be similarly obtained by one of ordinary skill in the art. Therefore, the present invention provides complete genomic sequences as well as cDNA sequences corresponding to the PS-interacting protein genes of the invention.

Alternatively, the nucleic acids of the invention may comprise recombinant genes or "minigenes" in which all or some introns of the PS-interacting protein genes have been removed, or in which various combinations of introns and exons and local cisacting regulatory elements have been engineered in propagation or expression constructs or vectors. For purposes of reducing the size of a recombinant PS-interacting protein gene, a cDNA gene may be employed, or various combinations of introns and untranslated exons may be removed from a DNA construct. These and many variations on these embodiments are now enabled by the identification and description of the PS-interacting proteins provided herein.

In addition to the disclosed PS-interacting protein and gene sequences, one of ordinary skill in the art is now enabled to identify and isolate nucleic acids representing PS-interacting genes or cDNAs which are allelic to the disclosed sequences or which are heterospecific homologues. Thus, the present invention provides isolated nucleic acids corresponding to these alleles and homologues, as well as the various above-described recombinant constructs derived from these sequences, by means which are well known in the art. Briefly, one of ordinary skill in the art may now screen preparations of genomic or cDNA, including samples prepared from individual organisms (e.g., human AD patients or their family members) as well as bacterial, viral, yeast or other libraries of genomic or cDNA, using probes or PCR primers to identify allelic or homologous sequences. Because it is desirable to identify mutations in the PS-interacting proteins which may contribute to the development of AD or other disorders, because it is desirable to identify polymorphisms in the PS-interacting proteins which are not pathogenic, and because it is also desirable to create a variety of animal models which may be used to study AD and screen for potential therapeutics, it is particularly contemplated that additional PS-interacting protein sequences will be isolated from other preparations or libraries

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of human nucleic acids and from preparations or libraries from animals including rats, mice, hamsters, guinea pigs, rabbits, dogs, cats, goats, sheep, pigs, and non-human primates. Furthermore, PS-interacting protein homologues from yeast or invertebrate species, including <u>C. elegans</u> and other nematodes, as well as <u>Drosophila</u> and other insects, may have particular utility for drug screening.

Standard hybridization screening or PCR techniques may be employed (as used, for example, in the identification of the mPS1 gene disclosed in PCT Publication WO96/34099) to identify and/or isolate such allelic and homologous sequences using relatively short PS-interacting protein gene sequences. The sequences may include 8 or fewer nucleotides depending upon the nature of the target sequences, the method employed, and the specificity required. Future technological developments may allow the advantageous use of even shorter sequences. With current technology, sequences of 9-50 nucleotides, and preferably about 18-24 are preferred. These sequences may be chosen from those disclosed herein, or may be derived from other allelic or heterospecific homologues enabled herein. When probing mRNA or screening cDNA libraries, probes and primers from coding sequences (rather than introns) are preferably employed, and sequences which are omitted in alternative splice variants typically are avoided unless it is specifically desired to identify those variants. Allelic variants of the PS-interacting protein genes may be expected to hybridize to the disclosed sequences under stringent hybridization conditions, as defined herein, whereas lower stringency may be employed to identify heterospecific homologues.

In another series of embodiments, the present invention provides for isolated nucleic acids which include subsets of the PS-interacting protein sequences or their complements. As noted above, such sequences will have utility as probes and PCR primers in the identification and isolation of allelic and homologous variants of the PS-interacting protein genes. Subsequences corresponding to polymorphic regions of the PS-interacting proteins, will also have particular utility in screening and/or genotyping individuals for diagnostic purposes, as described below. In addition, and also as described below, such subsets will have utility for encoding (1)

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fragments of the PS-interacting proteins for inclusion in fusion proteins, (2) fragments which comprise functional domains of the PS-interacting proteins for use in binding studies, (3) fragments of the PS-interacting proteins which may be used as immunogens to raise antibodies against the PS-interacting proteins, and (4) fragments of the PS-interacting proteins which may act as competitive inhibitors or as mimetics of the PS-interacting proteins to inhibit or mimic their physiological functions. Finally, such subsets may encode or represent complementary or antisense sequences which can hybridize to the PS-interacting protein genes or PS-interacting protein mRNA transcripts under physiological conditions to inhibit the transcription or translation of those sequences. Therefore, depending upon the intended use, the present invention provides nucleic acid subsequences of the PS-interacting protein genes which may have lengths varying from 8-10 nucleotides (e.g., for use as PCR primers) to nearly the full size of the PS-interacting protein genomic or cDNAs. Thus, the present invention provides isolated nucleic acids comprising sequences corresponding to at least 8-10, preferably 15, and more preferably at least 20 consecutive nucleotides of the PS-interacting protein genes, as disclosed or otherwise enabled herein, or to their complements. As noted above, however, shorter sequences may be useful with different technologies.

In another series of embodiments, the present invention provides nucleic acids in which the coding sequences for the PS-interacting proteins, with or without introns or recombinantly engineered as described above, are operably joined to endogenous or exogenous 5' and/or 3' regulatory regions. Using the present disclosure and standard genetic techniques (e.g., PCR extensions, targeting gene walking), one of ordinary skill in the art is now enabled to clone the 5' and/or 3' endogenous regulatory regions of any of the disclosed PS-interacting protein genes. Similarly, allelic variants of these endogenous regulatory regions, as well as endogenous regulatory regions from other mammalian homologues, are similarly enabled without undue experimentation. Alternatively, exogenous regulatory regions (i.e., regulatory regions from a different conspecific gene or a heterospecific regulatory region) may be operably joined to the PS-interacting protein coding sequences in order to drive

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expression. Appropriate 5' regulatory regions will include promoter elements and may also include additional elements such as operator or enhancer sequences. ribosome binding sequences, RNA capping sequences, and the like. The regulatory region may be selected from sequences that control the expression of genes of prokaryotic or eukaryotic cells, their viruses, and combinations thereof. Such regulatory regions include, but are not limited to, the lac system, the trp system, the tac system, and the trc system; major operator and promoter regions of phage λ ; the control region of the fd coat protein; early and late promoters of SV40; promoters derived from polyoma, adenovirus, retrovirus, baculovirus, and simian virus; 3phosphoglycerate kinase promoter; yeast acid phosphatase promoters; yeast alphamating factors; promoter elements of other eukaryotic genes expressed in neurons or other cell types; and combinations thereof. In particular, regulatory elements may be chosen which are inducible or repressible (e.g., the β-galactosidase promoter) to allow for controlled and/or manipulable expression of the PS-interacting protein genes in cells transformed with these nucleic acids. Alternatively, the PS-interacting protein coding regions may be operably joined with regulatory elements which provide for tissue specific expression in multicellular organisms. Such constructs are particularly useful for the production of transgenic organisms to cause expression of the PSinteracting protein genes only in appropriate tissues. The choice of appropriate regulatory regions is within the ability and discretion of one of ordinary skill in the art and the recombinant use of many such regulatory regions is now established in the art.

In another series of embodiments, the present invention provides for isolated nucleic acids encoding all or a portion of the PS-interacting proteins in the form of a fusion protein. In these embodiments, a nucleic acid regulatory region (endogenous or exogenous) is operably joined to a first coding region which is covalently joined in-frame to a second coding region. The second coding region optionally may be covalently joined to one or more additional coding regions and the last coding region is joined to a termination codon and, optionally, appropriate 3' regulatory regions (e.g., polyadenylation signals). The PS-interacting protein sequences of the fusion protein may represent the first, second, or any additional

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coding regions. The PS-interacting protein sequences may be conserved or non-conserved domains and can be placed in any coding region of the fusion. The non-PS-interacting protein sequences of the fusion may be chosen according to the needs and discretion of the practitioner and are not limited by the present invention. Useful non-PS-interacting protein sequences include, for example, short sequence "tags" such as antigenic determinants or poly-His tags which may be used to aid in the identification or purification of the resultant fusion protein. Alternatively, the non-PS-interacting protein coding region may encode a large protein or protein fragment, such as an enzyme or binding protein which also may assist in the identification and purification of the protein, or which may be useful in an assay such as those described below. Particularly contemplated fusion proteins include poly-His and GST (glutathione S-transferase) fusions which are useful in isolating and purifying the presenilins-interacting proteins, and the yeast two hybrid fusions, described below, which are useful in assays to identify other proteins which bind to or interact with the PS-interacting proteins.

In another series of embodiments, the present invention provides isolated nucleic acids in the form of recombinant DNA constructs in which a marker or reporter gene (e.g., β -galactosidase, luciferase) is operably joined to the 5' regulatory region of a PS-interacting protein gene such that expression of the marker gene is under the control of those regulatory sequences. Using the PS-interacting protein regulatory regions enabled herein, including regulatory regions from human and other mammalian species, one of ordinary skill in the art is now enabled to produce such constructs. As discussed more fully below, such isolated nucleic acids may be used to produce cells, cell lines or transgenic animals which are useful in the identification of compounds which can, directly or indirectly, differentially affect the expression of the PS-interacting proteins.

Finally, the isolated nucleic acids of the present invention include any of the above described sequences when included in vectors. Appropriate vectors include cloning vectors and expression vectors of all types, including plasmids, phagemids, cosmids, episomes, and the like, as well as integration vectors. The vectors may also

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include various marker genes (e.g., antibiotic resistance or susceptibility genes) which are useful in identifying cells successfully transformed therewith. In addition, the vectors may include regulatory sequences to which the nucleic acids of the invention are operably joined, and/or may also include coding regions such that the nucleic acids of the invention, when appropriately ligated into the vector, are expressed as fusion proteins. Such vectors may also include vectors for use in yeast "two hybrid," baculovirus, and phage-display systems. The vectors may be chosen to be useful for prokaryotic, eukaryotic or viral expression, as needed or desired for the particular application. For example, vaccinia virus vectors or simian virus vectors with the SV40 promoter (e.g., pSV2), or Herpes simplex virus or adeno-associated virus may be useful for transfection of mammalian cells including neurons in culture or in vivo, and the baculovirus vectors may be used in transfecting insect cells (e.g., butterfly cells). A great variety of different vectors are now commercially available and otherwise known in the art, and the choice of an appropriate vector is within the ability and discretion of one of ordinary skill in the art.

2. Substantially Pure Proteins

The present invention provides for substantially pure preparations of the PS-interacting proteins, fragments of the PS-interacting proteins, and fusion proteins including the PS-interacting proteins or fragments thereof. The proteins, fragments and fusions have utility, as described herein, in the generation of antibodies to normal and mutant PS-interacting proteins, in the identification of proteins (aside from the presentlins) which bind to the PS-interacting proteins, and in diagnostic and therapeutic methods. Therefore, depending upon the intended use, the present invention provides substantially pure proteins or peptides comprising amino acid sequences which are subsequences of the complete PS-interacting proteins and which may have lengths varying from 4-10 amino acids (e.g., for use as immunogens), or 10-100 amino acids (e.g., for use in binding assays), to the complete PS-interacting proteins. Thus, the present invention provides substantially pure proteins or peptides comprising sequences corresponding to at least 4-5, preferably 6-10, and more

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preferably at least 50 or 100 consecutive amino acids of the PS-interacting proteins, as disclosed or otherwise enabled herein.

The proteins or peptides of the invention may be isolated and purified by any of a variety of methods selected on the basis of the properties revealed by their protein sequences. For example, the PS-interacting proteins may be isolated from cells in which the PS-interacting protein is normally highly expressed. Alternatively the PS-interacting protein, fusion protein, or fragment thereof, may be purified from cells transformed or transfected with expression vectors (e.g., baculovirus systems such as the pPbac and pMbac vectors (Stratagene, La Jolla, CA); yeast expression systems such as the pYESHIS Xpress vectors (Invitrogen, San Diego, CA); eukaryotic expression systems such as pcDNA3 (Invitrogen, San Diego, CA) which has constant constitutive expression, or LacSwitch (Stratagene, La Jolla, CA) which is inducible; or prokaryotic expression vectors such as pKK233-3 (Clontech, Palo Alto, CA). In the event that the protein or fragment integrates into the endoplasmic reticulum or plasma membrane of the recombinant cells (e.g., eukaryotic cells), the protein may be purified from the membrane fraction. Alternatively, if the protein aggregates in inclusion bodies within the recombinant cells (e.g., prokaryotic cells), the protein may be purified from whole lysed cells or from solubilized inclusion bodies.

Purification can be achieved using standard protein purification procedures including, but not limited to, gel-filtration chromatography, ion-exchange chromatography, high-performance liquid chromatography (RP-HPLC, ion-exchange HPLC, size-exclusion HPLC, high-performance chromatofocusing chromatography, hydrophobic interaction chromatography, immunoprecipitation, or immunoaffinity purification. Gel electrophoresis (e.g., PAGE, SDS-PAGE) can also be used to isolate a protein or peptide based on its molecular weight, charge properties and hydrophobicity.

A PS-interacting protein, or a fragment thereof, may also be conveniently purified by creating a fusion protein including the desired PS-interacting protein sequence fused to another peptide such as an antigenic determinant or poly-His tag (e.g., QIAexpress vectors, QIAGEN Corp., Chatsworth, CA), or a larger protein (e.g.,

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GST using the pGEX-27 vector (Amrad, USA) or green fluorescent protein using the Green Lantern vector (GIBCO/BRL. Gaithersburg, MD). The fusion protein may be expressed and recovered from prokaryotic or eukaryotic cells and purified by any standard method based upon the fusion vector sequence. For example, the fusion protein may be purified by immunoaffinity or immunoprecipitation with an antibody to the non-PS-interacting protein portion of the fusion or, in the case of a poly-His tag, by affinity binding to a nickel column. The desired PS-interacting protein or fragment may then be further purified from the fusion protein by enzymatic cleavage of the fusion protein. Methods for preparing and using such fusion constructs for the purification of proteins are well known in the art and several kits are commercially available for this purpose. In light of the present disclosure, one is now enabled to employ such fusion constructs with the PS-interacting proteins.

3. Antibodies to the PS-interacting Proteins

The present invention also provides antibodies, and methods of making antibodies, which selectively bind to the PS-interacting proteins or fragments thereof. Of particular importance, by identifying the PS-interacting domains of the PS-interacting proteins, and methods of identifying mutant forms of the PS-interacting proteins associated with Alzheimer's Disease, the present invention provides antibodies, and methods of making antibodies, which will selectively bind to and, thereby, identify and/or distinguish normal and mutant (i.e., pathogenic) forms of the PS-interacting proteins. The antibodies of the invention have utility as laboratory reagents for, inter alia, immunoaffinity purification of the PS-interacting proteins, and immunocytochemistry or immunofluorescence techniques to establish the subcellular location of the proteins. In addition, as described below, the antibodies of the invention may be used as diagnostics tools to identify carriers of AD-related PS-interacting protein alleles, or as therapeutic tools to selectively bind and inhibit pathogenic forms of the PS-interacting proteins in vivo.

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The antibodies of the invention may be generated using the entire PS-interacting proteins of the invention, or using any PS-interacting protein epitope which is characteristic of that protein and which substantially distinguishes it from other host proteins. Any method of choosing antigenic determinants known in the art may, of course, be employed. Such epitopes may be identified by comparing sequences of, for example, 4-10 amino acid residues from a PS-interacting protein sequence to computer databases of protein sequences from the relevant host. In addition, larger fragments (e.g., 8-20 or, preferably, 9-15 residues) including one or more potential epitopes may also be employed. Antibodies to the PS-interacting domains (identified by the yeast two-hybrid assays described below) are expected to have the greatest utility both diagnostically and therapeutically. On the other hand, antibodies against highly conserved domains are expected to have the greatest utility for purification or identification of PS-interacting proteins.

PS-interacting protein immunogen preparations may be produced from crude extracts (e.g., lysates or membrane fractions of cells highly expressing the proteins), from proteins or peptides substantially purified from cells which naturally or recombinantly express them or, for short immunogens, by chemical peptide synthesis. The immunogens may also be in the form of a fusion protein in which the non-PS-interacting protein region is chosen for its adjuvant properties. As used herein, a PS-interacting protein immunogen shall be defined as a preparation including a peptide comprising at least 4-8, and preferably at least 9-15 consecutive amino acid residues of a PS-interacting proteins, as disclosed or otherwise enabled herein. Sequences of fewer residues may, of course, also have utility depending upon the intended use and future technological developments. Therefore, any PS-interacting protein derived sequences which are employed to generate antibodies to the PS-interacting proteins should be regarded as PS-interacting protein immunogens.

The antibodies of the invention may be polyclonal or monoclonal, or may be antibody fragments, including Fab fragments, F(ab')₂, and single chain antibody fragments. In addition, after identifying useful antibodies by the method of the invention, recombinant antibodies may be generated, including any of the antibody

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fragments listed above, as well as humanized antibodies based upon non-human antibodies to the PS-interacting proteins. In light of the present disclosure, as well as the characterization of other PS-interacting proteins enabled herein, one of ordinary skill in the art may produce the above-described antibodies by any of a variety of standard means well known in the art. For an overview of antibody techniques, see Antibody Engineering: A Practical Guide, Borrebaek, ed., W.H. Freeman & Company, NY (1992), or Antibody Engineering, 2nd Ed., Borrebaek, ed., Oxford University Press, Oxford (1995).

As a general matter, polyclonal antibodies may be generated by first immunizing a mouse, rabbit, goat or other suitable animal with the PS-interacting protein immunogen in a suitable carrier. To increase the immunogenicity of the preparation, the immunogen may be coupled to a carrier protein or mixed with an adjuvant (e.g., Freund's adjuvant). Booster injections, although not necessary are recommended. After an appropriate period to allow for the development of a humoral response, preferably several weeks, the animals may be bled and the sera may be purified to isolate the immunoglobulin component.

Similarly, as a general matter, monoclonal anti-PS-interacting protein antibodies may be produced by first injecting a mouse, rabbit, goat or other suitable animal with a PS-interacting protein immunogen in a suitable carrier. As above, carrier proteins or adjuvants may be utilized and booster injections (e.g., bi- or tri-weekly over 8-10 weeks) are recommended. After allowing for development of a humoral response, the animals are sacrificed and their spleens are removed and resuspended in, for example, phosphate buffered saline (PBS). The spleen cells serve as a source of lymphocytes, some of which are producing antibody of the appropriate specificity. These cells are then fused with an immortalized cell line (e.g., myeloma), and the products of the fusion are plated into a number of tissue culture wells in the presence of a selective agent such as HAT. The wells are serially screened and replated, each time selecting cells making useful antibody. Typically, several screening and replating procedures are carried out until over 90% of the wells contain single clones which are positive for antibody production. Monoclonal antibodies

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produced by such clones may be purified by standard methods such as affinity chromatography using Protein A Sepharose, by ion-exchange chromatography, or by variations and combinations of these techniques.

The antibodies of the invention may be labelled or conjugated with other compounds or materials for diagnostic and/or therapeutic uses. For example, they may be coupled to radionuclides, fluorescent compounds, or enzymes for imaging or therapy, or to liposomes for the targeting of compounds contained in the liposomes to a specific tissue location.

10 4. Transformed Cell Lines

The present invention also provides for cells or cell lines, both prokaryotic and eukaryotic, which have been transformed or transfected with the nucleic acids of the present invention so as to cause clonal propagation of those nucleic acids and/or expression of the proteins or peptides encoded thereby. Such cells or cell lines will have utility both in the propagation and production of the nucleic acids and proteins of the present invention but also, as further described herein, as model systems for diagnostic and therapeutic assays. In particular, it is expected that cells cotransformed with PS-interacting protein sequences as well as presentlin sequences will have improved utility as models of the biochemical pathways which may be affected in AD. For example, cells co-transformed with the interacting domains of PSinteracting sequences and presenilins in yeast two-hybrid fusion constructs, will have utility in screening for compounds which either enhance or inhibit interactions between these domains. Similarly, for cells transformed with a heterospecific presenilin, co-transformation with a similarly heterospecific PS-interacting protein, or co-transformation and homologous recombination to introduce a similarly heterospecific PS-interacting domain of a PS-interacting protein (e.g., "humanizing" a non-human endogenous PS-interacting protein), will result in a better model system for studying the interactions of the presenilins and the PS-interacting proteins. Cells transformed with only PS-interacting sequences will, of course, have utility of their

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own for studying the role of these proteins in the etiology of AD, and also as precursors for presentilin co-transformed cells.

As used herein, the term "transformed cell" is intended to embrace any cell, or the descendant of any cell, into which has been introduced any of the nucleic acids of the invention, whether by transformation, transfection, infection, or other means. Methods of producing appropriate vectors, transforming cells with those vectors, and identifying transformants are well known in the art and are only briefly reviewed here (see, for example, Sambrook et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York).

Prokaryotic cells useful for producing the transformed cells of the invention include members of the bacterial genera Escherichia (e.g., E. coli),

Pseudomonas (e.g., P. aeruginosa), and Bacillus (e.g., B. subtillus, B. stearothermophilus), as well as many others well known and frequently used in the art. Prokaryotic cells are particularly useful for the production of large quantities of the proteins or peptides of the invention (e.g., normal or mutant PS-interacting proteins, fragments of the PS-interacting proteins, fusion proteins of the PS-interacting proteins). Bacterial cells (e.g., E. coli) may be used with a variety of expression vector systems including, for example, plasmids with the T7 RNA polymerase/promoter system, bacteriophage λ regulatory sequences, or M13 Phage mGPI-2. Bacterial hosts may also be transformed with fusion protein vectors which create, for example, lacZ, trpE, maltose-binding protein, poly-His tags, or glutathione-S-transferase fusion proteins. All of these, as well as many other prokaryotic expression systems, are well known in the art and widely available commercially (e.g., pGEX-27 (Amrad, USA) for GST fusions).

Eukaryotic cells and cell lines useful for producing the transformed cells of the invention include mammalian cells and cell lines (e.g., PC12, COS, CHO, fibroblasts, myelomas, neuroblastomas, hybridomas, human embryonic kidney 293, oocytes, embryonic stem cells), insect cells lines (e.g., using baculovirus vectors such as pPbac or pMbac (Stratagene, La Jolla, CA)), yeast (e.g., using yeast expression

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vectors such as pYESHIS (Invitrogen, CA)), and fungi. Eukaryotic cells are particularly useful for embodiments in which it is necessary that the PS-interacting proteins, or functional fragments thereof, perform the functions and/or undergo the intracellular interactions associated with either the normal or mutant proteins. Thus, for example, transformed eukaryotic cells are preferred for use as models of PS-interacting protein function or interaction, and assays for screening candidate therapeutics preferably employ transformed eukaryotic cells.

To accomplish expression in eukaryotic cells, a wide variety of vectors have been developed and are commercially available which allow inducible (e.g., LacSwitch expression vectors, Stratagene, La Jolla, CA) or cognate (e.g., pcDNA3 vectors, Invitrogen, Chatsworth, CA) expression of PS-interacting protein nucleotide sequences under the regulation of an artificial promoter element. Such promoter elements are often derived from CMV or SV40 viral genes, although other strong promoter elements which are active in eukaryotic cells can also be employed to induce transcription of PS-interacting protein nucleotide sequences. Typically, these vectors also contain an artificial polyadenylation sequence and 3' UTR which can also be derived from exogenous viral gene sequences or from other eukaryotic genes. Furthermore, in some constructs, artificial, non-coding, spliceable introns and exons are included in the vector to enhance expression of the nucleotide sequence of interest. These expression systems are commonly available from commercial sources and are typified by vectors such as pcDNA3 and pZeoSV (Invitrogen, San Diego, CA). Innumerable commercially-available as well as custom-designed expression vectors are available from commercial sources to allow expression of any desired PSinteracting protein transcript in more or less any desired cell type, either constitutively or after exposure to a certain exogenous stimulus (e.g., withdrawal of tetracycline or exposure to IPTG).

Vectors may be introduced into the recipient or "host" cells by various methods well known in the art including, but not limited to, calcium phosphate transfection, strontium phosphate transfection, DEAE dextran transfection, electroporation, lipofection (e.g., Dosper Liposomal transfection reagent, Boehringer

Mannheim, Germany), microinjection, ballistic insertion on micro-beads, protoplast fusion or, for viral or phage vectors, by infection with the recombinant virus or phage.

5. Transgenic Animal Models

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The present invention also provides for the production of transgenic nonhuman animal models in which mutant or wild type PS-interacting protein sequences are expressed, or in which the PS-interacting protein genes have been inactivated (e.g., "knock-out" deletions), for the study of Alzheimer's Disease, for the screening of candidate pharmaceutical compounds, for the creation of explanted mammalian CNS cell cultures (e.g., neuronal, glial, organotypic or mixed cell cultures), and for the evaluation of potential therapeutic interventions. Prior to the present invention, a partial animal model for Alzheimer's Disease existed via the insertion and overexpression of a mutant form of the human amyloid precursor protein gene as a minigene under the regulation of the platelet-derived growth factor β receptor promoter element (Games et al., 1995). This mutant (βAPP_{717} Val \rightarrow Ile) causes the appearance of synaptic pathology and amyloid β peptide deposition in the brain of transgenic animals bearing this transgene in high copy number. These changes in the brain of the transgenic animal are very similar to that seen in human AD (Games et al., 1995). It is, however, as yet unclear whether these animals become demented, but there is general consensus that it is now possible to recreate at least some aspects of AD in mice. In addition, transgenic animal models in which the presentilin genes are genetically engineered are disclosed in PCT Publication WO96/34099. These transgenic animal models have been shown to have altered A\(\beta\) production and altered hippocampus-dependent memory function.

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Animal species suitable for use in the animal models of the present invention include, but are not limited to, rats, mice, hamsters, guinea pigs, rabbits, dogs, cats, goats, sheep, pigs, and non-human primates (e.g., Rhesus monkeys, chimpanzees). For initial studies, transgenic rodents (e.g., mice) may be preferred due to their relative ease of maintenance and shorter life spans. However, transgenic yeast or invertebrates (e.g., nematodes, insects) may be preferred for some studies because

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they will allow for even more rapid and inexpensive screening. For example, invertebrates bearing mutant PS-interacting protein homologues (or mammalian PS-interacting protein transgenes) which cause a rapidly occurring and easily scored phenotype (e.g., abnormal vulva or eye development after several days) can be used as screens for drugs which block the effect of the mutant gene. Such invertebrates may prove far more rapid and efficient for mass screenings than larger vertebrate animals. Once lead compounds are found through such screens, they may be tested in higher animals such a rodents. Ultimately, transgenic non-human primates may be preferred for longer term studies due to their greater similarity to humans and their higher cognitive abilities.

Using the nucleic acids disclosed and otherwise enabled herein, there are now several available approaches for the creation of a transgenic animal model for Alzheimer's Disease. Thus, the enabled animal models include: (1) Animals in which sequences encoding at least a functional domain of a normal human PS-interacting protein gene have been recombinantly introduced into the genome of the animal as an additional gene, under the regulation of either an exogenous or an endogenous promoter element, and as either a minigene or a large genomic fragment; in which sequences encoding at least a functional domain of a normal human PS-interacting protein gene have been recombinantly substituted for one or both copies of the animal's homologous PS-interacting protein gene by homologous recombination or gene targeting; and/or in which one or both copies of one of the animal's homologous PS-interacting protein genes have been recombinantly "humanized" by the partial substitution of sequences encoding the human homologue by homologous recombination or gene targeting. These animals are useful for evaluating the effects of the transgenic procedures, and the effects of the introduction or substitution of a human or humanized PS-interacting protein gene. (2) Animals in which sequences encoding at least a functional domain of a mutant (i.e., pathogenic) human PSinteracting protein gene have been recombinantly introduced into the genome of the animal as an additional gene, under the regulation of either an exogenous or an endogenous promoter element, and as either a minigene or a large genomic fragment;

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in which sequences encoding at least a functional domain of a mutant human PSinteracting protein gene have been recombinantly substituted for one or both copies of the animal's homologous PS-interacting protein gene by homologous recombination or gene targeting; and/or in which one or both copies of one of the animal's homologous PS-interacting protein genes have been recombinantly "humanized" by the partial substitution of sequences encoding a mutant human homologue by homologous recombination or gene targeting. These animals are useful as models which will display some or all of the characteristics, whether at the biochemical, physiological and/or behavioral level, of humans carrying one or more alleles which are pathogenic of Alzheimer's Disease or other diseases associated with mutations in the PS-interacting protein genes. (3) Animals in which sequences encoding at least a functional domain of a mutant version of one of that animal's PS-interacting protein genes (bearing, for example, a specific mutation corresponding to, or similar to, one of the pathogenic mutations of the human PS-interacting proteins) have been recombinantly introduced into the genome of the animal as an additional gene, under the regulation of either an exogenous or an endogenous promoter element, and as either a minigene or a large genomic fragment; and/or in which sequences encoding at least a functional domain of a mutant version of one of that animal's PS-interacting protein genes (bearing, for example, a specific mutation corresponding to, or similar to, one of the pathogenic mutations of the human PS-interacting proteins) have been recombinantly substituted for one or both copies of the animal's homologous PSinteracting protein gene by homologous recombination or gene targeting. These animals are also useful as models which will display some or all of the characteristics, whether at the biochemical, physiological and/or behavioral level, of humans carrying one or more alleles which are pathogenic of Alzheimer's Disease. (4) "Knock-out" animals in which one or both copies of one of the animal's PS-interacting protein genes have been partially or completely deleted by homologous recombination or gene targeting, or have been inactivated by the insertion or substitution by homologous recombination or gene targeting of exogenous sequences (e.g., stop codons, lox p sites). Such animals are useful models to study the effects which loss of

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PS-interacting protein gene expression may have, to evaluate whether loss of function is preferable to continued expression of mutant forms, and to examine whether other genes can be recruited to replace a mutant PS-interacting protein or to intervene with the effects of other genes (e.g., PS1, PS2, APP or ApoE) causing AD as a treatment for AD or other disorders. For example, a normal PS-interacting protein gene may be necessary for the action of mutant presentilin or APP genes to actually be expressed as AD and, therefore, transgenic PS-interacting protein animal models may be of use in elucidating such multigenic interactions.

In addition to transgenic animal models in which the expression of one or more of the PS-interacting proteins is altered, the present invention also provides for the production of transgenic animal models in which the expression of one or more of the presenilins, APP, or ApoE is altered. The nucleic acids encoding the presenilins, APP, and ApoE are known in the art, a methods for producing transgenic animals with these sequences are also known (see, e.g., PCT Publication WO96/34099; Games et al., 1995). Indeed, because non-human animals may differ from humans not only in their PS-interacting protein sequences, but also in the sequences of their presenilin, APP and/or ApoE homologues, it is particularly contemplated that transgenics may be produced which bear recombinant normal or mutant human sequences for at least one presenilin, APP and/or ApoE gene in addition to recombinant sequences for one or more PS-interacting proteins. Such co-transformed animal models would possess more elements of the human molecular biology and, therefore, are expected to be better models of human disorders. Thus, in accordance with the present invention, transgenic animal models may be produced bearing normal or mutant sequences for one or more PS-interacting proteins, or interacting domains of these proteins. These animals will have utility in that they can be crossed with animals bearing a variety of normal or mutant presenilin, APP or ApoE sequences to produce co-transformed animal models. Furthermore, as detailed below, it is expected that mutations in the PS-interacting genes, like mutations in the presenilins themselves, may be causative of Alzheimer's Disease and/or other disorders as well (e.g., other cognitive, intellectual, neurological or psychological disorders such as cerebral hemorrhage,

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schizophrenia, depression, mental retardation and epilepsy). Therefore, transgenic animal models bearing normal or mutant sequences corresponding to the PS-interacting proteins, absent transformation with any presentilin, APP or ApoE sequences, will have utility of their own in the study of such disorders.

As detailed below, preferred choices for transgenic animal models transformed with PS-interacting proteins, or domains of PS-interacting proteins, include those transformed with normal or mutant sequences corresponding to the clones identified and described in Example 1 and disclosed in SEQ ID NOs: 1-12. These clones, which interact with normal or mutant PS1 TM6 \rightarrow 7 loop domains, were identified according to the methods described in Example 1, below, and PCT Publication WO96/34099. These clones, longer nucleic acid sequences comprising these clones, and other clones identified according to this and other methods of the invention (e.g., allelic and splice variants or heterospecific homologues of these clones) may all be employed in accordance with the present invention to produce animal models which, with or without co-transformation with presenilin, APP and/or ApoE sequences, will have utility in the study of Alzheimer's Disease and/or other cognitive, intellectual, neurological or psychological disorders.

Thus, using the nucleic acids disclosed and otherwise enabled herein, one of ordinary skill in the art may now produce any of the following types of transgenic animal models with altered PS-interacting protein expression: (1) Animals in which sequences encoding at least a functional domain of a normal human PS-interacting protein gene have been recombinantly introduced into the genome of the animal as an additional gene, under the regulation of either an exogenous or an endogenous promoter element, and as either a minigene or a large genomic fragment; in which sequences encoding at least a functional domain of a normal human PS-interacting protein gene have been recombinantly substituted for one or both copies of the animal's homologous PS-interacting protein gene by homologous recombination or gene targeting; and/or in which one or both copies of one of the animal's homologous PS-interacting protein genes have been recombinantly "humanized" by the partial substitution of sequences encoding the human homologue by homologous

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recombination or gene targeting. These animals are particularly useful for providing transgenic models which express human PS-interacting proteins as well as human presenilin proteins. They are also useful in evaluating the effects of the transgenic procedures, and the effects of the introduction or substitution of a human or humanized PS-interacting protein gene. (2) Animals in which sequences encoding at least a functional domain of a mutant (i.e., pathogenic) human PS-interacting protein gene have been recombinantly introduced into the genome of the animal as an additional gene, under the regulation of either an exogenous or an endogenous promoter element, and as either a minigene or a large genomic fragment; in which sequences encoding at least a functional domain of a mutant human PS-interacting protein gene have been recombinantly substituted for one or both copies of the animal's homologous PS-interacting protein gene by homologous recombination or gene targeting; and/or in which one or both copies of one of the animal's homologous PS-interacting protein genes have been recombinantly "humanized" by the partial substitution of sequences encoding a mutant human homologue by homologous recombination or gene targeting. These animals are useful as models which will display some or all of the characteristics, whether at the biochemical, physiological and/or behavioral level, of humans carrying one or more alleles which are pathogenic of Alzheimer's Disease or other diseases associated with mutations in these PSinteracting genes. (3) Animals in which sequences encoding at least a functional domain of a mutant version of one of that animal's PS-interacting protein genes (bearing, for example, a specific mutation corresponding to, or similar to, one of the pathogenic mutations of the human PS-interacting proteins) have been recombinantly introduced into the genome of the animal as an additional gene, under the regulation of either an exogenous or an endogenous promoter element, and as either a minigene or a large genomic fragment; and/or in which sequences encoding at least a functional domain of a mutant version of one of that animal's PS-interacting protein genes (bearing, for example, a specific mutation corresponding to, or similar to, one of the pathogenic mutations of the humans PS-interacting proteins) have been recombinantly substituted for one or both copies of the animal's homologous PS-interacting protein

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gene by homologous recombination or gene targeting. These animals are also useful as models which will display some or all of the characteristics, whether at the biochemical, physiological and/or behavioral level, of humans carrying one or more alleles which are pathogenic of Alzheimer's Disease. (4) "Knock-out" animals in which one or both copies of one of the animal's PS-interacting protein genes have been partially or completely deleted by homologous recombination or gene targeting, or have been inactivated by the insertion or substitution by homologous recombination or gene targeting of exogenous sequences (e.g., stop codons, lox p sites). Such animals are useful models to study the effects which loss of PSinteracting protein gene expression may have, to evaluate whether loss of function is preferable to continued expression, and to examine whether other genes can be recruited to replace a mutant PS-interacting protein or to intervene with the effects of other genes (e.g., APP or ApoE) causing AD as a treatment for AD or other disorders. For example, a normal PS-interacting protein may be necessary for the action of mutant PS1, PS2 or APP genes to actually be expressed as AD and, therefore, transgenic PS-interacting protein animal models may be of use in elucidating such multigenic interactions.

In some preferred embodiments, transgenic animal models are produced in which just the PS-interacting domains of the PS-interacting proteins are introduced into the genome of the animal by homologous recombination. Thus, for example, preferred embodiments include transgenic animals in which the PS-interacting domains of PS-interacting proteins are "humanized" by homologous recombination with sequences from human PS-interacting proteins. These animals may then be bred with transgenics in which normal or mutant presentilin sequences have been introduced. The progeny of these animals, having both human presentilin and human PS-interacting protein sequences, will provide improved animal models for Alzheimer's Disease.

To create an animal model (e.g., a transgenic mouse), a normal or mutant PS-interacting gene (e.g., normal or mutant S5a, GT24, p0071, Rab11, etc.), or a normal or mutant version of a recombinant nucleic acid encoding at least a functional

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domain of a PS-interacting gene (e.g., the PS-interacting domains obtained in the yeast two-hybrid system), can be inserted into a germ line or stem cell using standard techniques of oocyte microinjection, or transfection or microinjection into embryonic stem cells. Animals produced by these or similar processes are referred to as transgenic. Similarly, if it is desired to inactivate or replace an endogenous presentiin or PS-interacting protein gene, homologous recombination using embryonic stem cells may be employed. Animals produced by these or similar processes are referred to as "knock-out" (inactivation) or "knock-in" (replacement) models.

For oocyte injection, one or more copies of the recombinant DNA constructs of the present invention may be inserted into the pronucleus of a just-fertilized oocyte. This oocyte is then reimplanted into a pseudo-pregnant foster mother. The liveborn animals are screened for integrants using analysis of DNA (e.g., from the tail veins of offspring mice) for the presence of the inserted recombinant transgene sequences. The transgene may be either a complete genomic sequence injected as a YAC, BAC, PAC or other chromosome DNA fragment, a cDNA with either the natural promoter or a heterologous promoter, or a minigene containing all of the coding region and other elements found to be necessary for optimum expression.

Retroviral infection of early embryos can also be done to insert the recombinant DNA constructs of the invention. In this method, the transgene (e.g., a normal or mutant S5a, GT24, p0071, Rab 11, etc., sequence) is inserted into a retroviral vector which is used to infect embryos (e.g., mouse or non-human primate embryos) directly during the early stages of development to generate chimeras, some of which will lead to germline transmission.

Homologous recombination using stem cells allows for the screening of gene transfer cells to identify the rare homologous recombination events. Once identified, these can be used to generate chimeras by injection of blastocysts, and a proportion of the resulting animals will show germline transmission from the recombinant line. This methodology is especially useful if inactivation of a gene is desired. For example, inactivation of the S5a gene in mice may be accomplished by designing a DNA fragment which contains sequences from an S5a coding region

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flanking a selectable marker. Homologous recombination leads to the insertion of the marker sequences in the middle of the coding region, causing inactivation of the S5a gene and/or deletion of internal sequences. DNA analysis of individual clones can then be used to recognize the homologous recombination events.

The techniques of generating transgenic animals, as well as the techniques for homologous recombination or gene targeting, are now widely accepted and practiced. A laboratory manual on the manipulation of the mouse embryo, for example, is available detailing standard laboratory techniques for the production of transgenic mice (Hogan et al., 1986). To create a transgene, the target sequence of interest (e.g., normal or mutant presentiin sequences, normal or mutant PS-interacting protein sequences) are typically ligated into a cloning site located downstream of some promoter element which will regulate the expression of RNA from the sequence. Downstream of the coding sequence, there is typically an artificial polyadenylation sequence. In the transgenic models that have been used to successfully create animals which mimic aspects of inherited human neurodegenerative diseases, the most successful promoter elements have been the platelet-derived growth factor receptor β gene subunit promoter and the harmster prion protein gene promoter, although other promoter elements which direct expression in central nervous system cells would also be useful. An alternate approach to creating a transgene is to use an endogenous presentilin or PS-interacting protein gene promoter and regulatory sequences to drive expression of the transgene. Finally, it is possible to create transgenes using large genomic DNA fragments such as YACs which contain the entire desired gene as well as its appropriate regulatory sequences. Such constructs have been successfully used to drive human APP expression in transgenic mice (Lamb et al., 1993).

Animal models can also be created by targeting the endogenous presentiin or PS-interacting protein gene in order to alter the endogenous sequence by homologous recombination. These targeting events can have the effect of removing endogenous sequence (knock-out) or altering the endogenous sequence to create an amino acid change associated with human disease or an otherwise abnormal sequence

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(e.g., a sequence which is more like the human sequence than the original animal sequence) (knock-in animal models). A large number of vectors are available to accomplish this and appropriate sources of genomic DNA for mouse and other animal genomes to be targeted are commercially available from companies such as GenomeSystems Inc. (St. Louis, Missouri, USA). The typical feature of these targeting vector constructs is that 2 to 4 kb of genomic DNA is ligated 5' to a selectable marker (e.g., a bacterial neomycin resistance gene under its own promoter element termed a "neomycin cassette"). A second DNA fragment from the gene of interest is then ligated downstream of the neomycin cassette but upstream of a second selectable marker (e.g., thymidine kinase). The DNA fragments are chosen such that mutant sequences can be introduced into the germ line of the targeted animal by homologous replacement of the endogenous sequences by either one of the sequences included in the vector. Alternatively, the sequences can be chosen to cause deletion of sequences that would normally reside between the left and right arms of the vector surrounding the neomycin cassette. The former is known as a knock-in, the latter is known as a knock-out. Again, innumerable model systems have been created, particularly for targeted knock-outs of genes including those relevant to neurodegenerative diseases (e.g., targeted deletions of the murine APP gene by Zheng et al., 1995; targeted deletion of the murine prion gene associated with adult onset human CNS degeneration by Bueler et al., 1996).

Finally, equivalents of transgenic animals, including animals with mutated or inactivated presentilin genes, or mutated or inactivated PS-interacting protein genes, may be produced using chemical or X-ray mutagenesis of gametes, followed by fertilization. Using the isolated nucleic acids disclosed or otherwise enabled herein, one of ordinary skill may more rapidly screen the resulting offspring by, for example, direct sequencing RFLP, PCR, or hybridization analysis to detect mutants, or Southern blotting to demonstrate loss of one allele by dosage.

6. Assays for Drugs Which Affect PS-Interacting Protein Expression

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In another series of embodiments, the present invention provides assays for identifying small molecules or other compounds which are capable of inducing or inhibiting the expression of the PS-interacting genes and proteins (e.g., S5a or GT24). The assays may be performed <u>in vitro</u> using non-transformed cells, immortalized cell lines, or recombinant cell lines, or <u>in vivo</u> using the transgenic animal models enabled herein.

In particular, the assays may detect the presence of increased or decreased expression of S5a, GT24, p0071, Rab 11, or other PS-interacting genes or proteins on the basis of increased or decreased mRNA expression (using, e.g., the nucleic acid probes disclosed and enabled herein), increased or decreased levels of PS-interacting proteins (using, e.g., the anti-PS-interacting protein antibodies disclosed and enabled herein), or increased or decreased levels of expression of a marker gene (e.g., β-galactosidase or luciferase) operably joined to a PS-interacting protein 5' regulatory region in a recombinant construct.

Thus, for example, one may culture cells known to express a particular PS-interacting protein and add to the culture medium one or more test compounds. After allowing a sufficient period of time (e.g., 0-72 hours) for the compound to induce or inhibit the expression of the PS-interacting protein, any change in levels of expression from an established baseline may be detected using any of the techniques described above and well known in the art. In particularly preferred embodiments, the cells are from an immortalized cell line such as a human neuroblastoma, glioblastoma or a hybridoma cell line. Using the nucleic acid probes and /or antibodies disclosed and enabled herein, detection of changes in the expression of a PS-interacting protein, and thus identification of the compound as an inducer or repressor of PS-interacting protein expression, requires only routine experimentation.

In particularly preferred embodiments, a recombinant assay is employed in which a reporter gene such a β-galactosidase, green fluorescent protein, alkaline phosphatase, or luciferase is operably joined to the 5' regulatory regions of a PS-interacting protein gene. Preferred vectors include the Green Lantern 1 vector (GIBCO/BRL, Gaithersburg, MD) and the Great EScAPe pSEAP vector (Clontech,

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Palo Alto). The PS-interacting protein regulatory regions may be easily isolated and cloned by one of ordinary skill in the art in light of the present disclosure of coding regions from these genes. The reporter gene and regulatory regions are joined inframe (or in each of the three possible reading frames) so that transcription and translation of the reporter gene may proceed under the control of the PS-interacting protein regulatory elements. The recombinant construct may then be introduced into any appropriate cell type, although mammalian cells are preferred, and human cells are most preferred. The transformed cells may be grown in culture and, after establishing the baseline level of expression of the reporter gene, test compounds may be added to the medium. The ease of detection of the expression of the reporter gene provides for a rapid, high through-put assay for the identification of inducers and repressors of the PS-interacting protein gene.

Compounds identified by this method will have potential utility in modifying the expression of the PS-interacting protein genes in vivo. These compounds may be further tested in the animal models disclosed and enabled herein to identify those compounds having the most potent in vivo effects. In addition, as described herein with respect to small molecules having binding activity for PS-interacting proteins, these molecules may serve as "lead compounds" for the further development of pharmaceuticals by, for example, subjecting the compounds to sequential modifications, molecular modeling, and other routine procedures employed in rational drug design.

7. Identification of Compounds with PS-Interacting Protein Binding Capacity

In light of the present disclosure, one of ordinary skill in the art is enabled to practice new screening methodologies which will be useful in the identification of proteins and other compounds which bind to, or otherwise directly interact with, the PS-interacting proteins. The proteins and compounds will include endogenous cellular components, aside from the presentilins, which interact with the PS-interacting proteins in vivo and which, therefore, provide new targets for pharmaceutical and therapeutic interventions, as well as recombinant, synthetic and otherwise exogenous

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compounds which may have PS-interacting protein binding capacity and, therefore, may be candidates for pharmaceutical agents. Thus, in one series of embodiments, cell lysates or tissue homogenates (e.g., human brain homogenates, lymphocyte lysates) may be screened for proteins or other compounds which bind to one of the normal or mutant PS-interacting proteins. Alternatively, any of a variety of exogenous compounds, both naturally occurring and/or synthetic (e.g., libraries of small molecules or peptides), may be screened for PS-interacting protein binding capacity. Small molecules are particular preferred in this context because they are more readily absorbed after oral administration, have fewer potential antigenic determinants, and/or are more likely to cross the blood brain barrier than larger molecules such as nucleic acids or proteins. The methods of the present invention are particularly useful in that they may be used to identify molecules which selectively or preferentially bind to a mutant form of a PS-interacting protein (rather than a normal form) and, therefore, may have particular utility in treating cases of AD which arise from mutations in the PS-interacting proteins.

Once identified by the methods described above, the candidate compounds may then be produced in quantities sufficient for pharmaceutical administration or testing (e.g., µg or mg or greater quantities), and formulated in a pharmaceutically acceptable carrier (see, e.g., Remington's Pharmaceutical Sciences, Gennaro, A., ed., Mack Pub., 1990). These candidate compounds may then be administered to the transformed cells of the invention, to the transgenic animal models of the invention, to cell lines derived from the animal models or from human patients, or to Alzheimer's patients. The animal models described and enabled herein are of particular utility in further testing candidate compounds which bind to normal or mutant PS-interacting proteins for their therapeutic efficacy.

In addition, once identified by the methods described above, the candidate compounds may also serve as "lead compounds" in the design and development of new pharmaceuticals. For example, as in well known in the art, sequential modification of small molecules (e.g., amino acid residue replacement with peptides; functional group replacement with peptide or non-peptide compounds) is a standard

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approach in the pharmaceutical industry for the development of new pharmaceuticals. Such development generally proceeds from a "lead compound" which is shown to have at least some of the activity (e.g., PS-interacting protein binding or blocking ability) of the desired pharmaceutical. In particular, when one or more compounds having at least some activity of interest (e.g., modulation of PS-interacting protein activity) are identified, structural comparison of the molecules can greatly inform the skilled practitioner by suggesting portions of the lead compounds which should be conserved and portions which may be varied in the design of new candidate compounds. Thus, the present invention also provides a means of identifying lead compounds which may be sequentially modified to produce new candidate compounds for use in the treatment of Alzheimer's Disease. These new compounds then may be tested both for binding to PS-interacting proteins and/or blocking PS-interacting protein activity, and for therapeutic efficacy (e.g., in the animal models described herein). This procedure may be iterated until compounds having the desired therapeutic activity and/or efficacy are identified.

In each of the present series of embodiments, an assay is conducted to detect binding between a "PS-interacting protein component" and some other moiety. Of particular utility will be sequential assays in which compounds are tested for the ability to bind to only normal or only mutant forms of the PS-interacting domains of PS-interacting proteins in the binding assays. Such compounds are expected to have the greatest therapeutic utilities, as described more fully below. The "PS-interacting protein component" in these assays may be a complete normal or mutant form of a PS-interacting protein (e.g., S5a, GT24, p0071, Rab 11, etc.) but need not be. Rather, particular functional domains of the PS-interacting proteins, particularly the PS-interacting domains as described above, may be employed either as separate molecules or as part of a fusion protein. For example, to isolate proteins or compounds that interact with these functional domains, screening may be carried out using fusion constructs and/or synthetic peptides corresponding to these regions.

Thus, for S5a, GST-fusion peptides may be made including sequences corresponding approximately to amino acids 70-377 of SEQ ID NO: 2 (included in clones Y2H29)

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and Y2H31, see Example 1), approximately to amino acids 206-377 of SEQ ID NO: 2 (which includes protein-protein interaction motifs, see Ferrell et al., 1996), or to any other S5a domain of interest. Similarly, for GT24, GST- or other fusion peptides may be produced including sequences corresponding approximately to amino acids 440-815 of SEQ ID NO: 4 (including part of the armadillo repeat segment). Obviously, various combinations of fusion proteins and PS-interacting protein functional domains are possible and these are merely examples. In addition, the functional domains may be altered so as to aid in the assay by, for example, introducing into the functional domain a reactive group or amino acid residue (e.g., cysteine) which will facilitate immobilization of the domain on a substrate (e.g., using sulfhydryl reactions). Thus, for example, the PS-interacting domain of S5a may be synthesized containing an additional C-terminal cysteine residue to facilitate immobilization of the domain. Such peptides may be used to create an affinity substrate for affinity chromatography (Sulfo-link; Pierce) to isolate binding proteins for microsequencing. Similarly, other functional domain or antigenic fragments may be created with modified residues (see, e.g., Example 4).

The proteins or other compounds identified by these methods may be purified and characterized by any of the standard methods known in the art. Proteins may, for example, be purified and separated using electrophoretic (e.g., SDS-PAGE, 2D PAGE) or chromatographic (e.g., HPLC) techniques and may then be microsequenced. For proteins with a blocked N-terminus, cleavage (e.g., by CNBr and/or trypsin) of the particular binding protein is used to release peptide fragments. Further purification/characterization by HPLC and microsequencing and/or mass spectrometry by conventional methods provides internal sequence data on such blocked proteins. For non-protein compounds, standard organic chemical analysis techniques (e.g., IR, NMR and mass spectrometry; functional group analysis; X-ray crystallography) may be employed to determine their structure and identity.

Methods for screening cellular lysates, tissue homogenates, or small molecule libraries for candidate PS-interaction protein-binding molecules are well known in the art and, in light of the present disclosure, may now be employed to

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identify compounds which bind to normal or mutant PS-interacting protein components or which modulate PS-interacting protein activity as defined by non-specific measures (e.g., changes in intracellular Ca²*, GTP/GDP ratio) or by specific measures (e.g., changes in Aβ peptide production or changes in the expression of other downstream genes which can be monitored by differential display, 2D gel electrophoresis, differential hybridization, or SAGE methods). The preferred methods involve variations on the following techniques: (1) direct extraction by affinity chromatography; (2) co-isolation of PS-interacting protein components and bound proteins or other compounds by immunoprecipitation; (3) the Biomolecular Interaction Assay (BIAcore); and (4) the yeast two-hybrid systems. These and others are discussed separately below.

A. Affinity Chromatography

In light of the present disclosure, a variety of affinity binding techniques well known in the art may be employed to isolate proteins or other compounds which bind to the PS-interacting protein disclosed or otherwise enabled herein. In general, a PS-interacting protein component may be immobilized on a substrate (e.g., a column or filter) and a solution including the test compound(s) is contacted with the PS-interacting protein, fusion or fragment under conditions which are permissive for binding. The substrate is then washed with a solution to remove unbound or weakly bound molecules. A second wash may then elute those compounds which strongly bound to the immobilized normal or mutant PS-interacting protein component.

Alternatively, the test compounds may be immobilized and a solution containing one or more PS-interacting protein components may be contacted with the column, filter or other substrate. The ability of the PS-interacting protein component to bind to the test compounds may be determined as above or a labeled form of the PS-interacting protein component (e.g., a radio-labeled or chemiluminescent functional domain) may be used to more rapidly assess binding to the substrate-immobilized compound(s).

B. Co-Immunoprecipitation

Another well characterized technique for the isolation of PS-interacting protein components and their associated proteins or other compounds is direct

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immunoprecipitation with antibodies. This procedure has been successfully used, for example, to isolate many of the synaptic vesicle associated proteins (Phizicky and Fields, 1994). Thus, either normal or mutant, free or membrane-bound PS-interacting protein components may be mixed in a solution with the candidate compound(s) under conditions which are permissive for binding, and the PS-interacting protein component may be immunoprecipitated. Proteins or other compounds which co-immunoprecipitate with the PS-interacting protein component may then be identified by standard techniques as described above. General techniques for immunoprecipitation may be found in, for example, Harlow and Lane, (1988)

Antibodies: A Laboratory Manual, Cold Spring Harbor Press, Cold Spring Harbor, NY.

The antibodies employed in this assay, as described and enabled herein, may be polyclonal or monoclonal, and include the various antibody fragments (e.g., Fab, F(ab')₂) as well as single chain antibodies, and the like.

C. The Biomolecular Interaction Assay

Another useful method for the detection and isolation of binding proteins is the Biomolecular Interaction Assay or "BIAcore" system developed by Pharmacia Biosensor and described in the manufacturer's protocol (LKB Pharmacia, Sweden). In light of the present disclosure, one of ordinary skill in the art is now enabled to employ this system, or a substantial equivalent, to identify proteins or other compounds having PS-interacting protein binding capacity. The BIAcore system uses an affinity purified anti-GST antibody to immobilize GST-fusion proteins onto a sensor chip. Obviously, other fusion proteins and corresponding antibodies may be substituted. The sensor utilizes surface plasmon resonance which is an optical phenomenon that detects changes in refractive indices. A homogenate of a tissue of interest is passed over the immobilized fusion protein and protein-protein interactions are registered as changes in the refractive index. This system can be used to determine the kinetics of binding and to assess whether any observed binding is of physiological relevance.

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D. The Yeast Two-Hybrid System

The yeast "two-hybrid" system takes advantage of transcriptional factors that are composed of two physically separable, functional domains (Phizicky and Fields, 1994). The most commonly used is the yeast GAL4 transcriptional activator consisting of a DNA binding domain and a transcriptional activation domain. Two different cloning vectors are used to generate separate fusions of the GAL4 domains to genes encoding potential binding proteins. The fusion proteins are co-expressed, targeted to the nucleus and, if interactions occur, activation of a reporter gene (e.g., lacZ) produces a detectable phenotype. For example, the Clontech Matchmaker System-2 may be used with the Clontech brain cDNA GAL4 activation domain fusion library with PS-interacting protein-GAL4 binding domain fusion clones (Clontech, Palo Alto, CA). In light of the disclosures herein, one of ordinary skill in the art is now enabled to produce a variety of PS-interacting protein fusions, including fusions including either normal or mutant functional domains of the PS-interacting proteins, and to screen such fusion libraries in order to identify PS-interacting protein binding proteins.

E. Other Methods

The nucleotide sequences and protein products, including both mutant and normal forms of these nucleic acids and their corresponding proteins, can be used with the above techniques to isolate other interacting proteins, and to identify other genes whose expression is altered by the over-expression of normal PS-interacting protein sequences, or by the under-expression of normal PS-interacting protein sequences, or by the expression of mutant PS-interacting protein sequences. Identification of these other interacting proteins, as well as the identification of other genes whose expression levels are altered in AD will identify other gene targets which have direct relevance to the pathogenesis of this disease in its clinical or pathological forms. Specifically, other genes will be identified which may themselves be the site of other mutations causing Alzheimer's Disease, or which can themselves be targeted therapeutically (e.g., to reduce their expression levels to normal, or to pharmacologically block the effects of their over-expression) as a potential treatment

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for this disease. Specifically, these techniques rely on PCR-based and/or hybridization-based methods to identify genes which are differentially expressed between two conditions (a cell line expressing normal PS-interacting proteins compared to the same cell type expressing a mutant PS-interacting protein). These techniques include differential display, serial analysis of gene expression (SAGE), and mass-spectrometry of protein 2D-gels and subtractive hybridization (reviewed in Nowak, 1995 and Kahn, 1995).

As will be obvious to one of ordinary skill in the art, there are numerous other methods of screening individual proteins or other compounds, as well as large libraries of proteins or other compounds (e.g., phage display libraries and cloning systems from Stratagene, La Jolla, CA) to identify molecules which bind to normal or mutant PS-interacting protein components. All of these methods comprise the step of mixing a normal or mutant PS-interacting protein, fusion, or fragment with test compounds, allowing for binding (if any), and assaying for bound complexes. All such methods are now enabled by the present disclosure of substantially pure PS-interacting proteins, substantially pure PS-interacting functional domain fragments, PS-interacting protein fusion proteins, PS-interacting protein antibodies, and methods of making and using the same.

20 8. Disrupting PS-Interacting Protein Interactions

The ability to disrupt specific interactions of the PS-interacting proteins with the presentions, or with other proteins, is potentially of great therapeutic value, and will be important in understanding the etiology of AD and in identifying additional targets for therapy. The methods used to identify compounds which disrupt PS-interacting protein interactions may be applied equally well to interactions involving either normal or mutant PS-interacting proteins.

Assays for compounds which can disrupt PS-interacting protein interactions may be performed by any of a variety of methods well known in the art. In essence, such assays will parallel those assays for identifying proteins and compounds with binding activity toward the PS-interacting proteins. Thus, once a

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compound with binding activity for a PS-interacting protein is identified by any method, that method or an equivalent method may be performed in the presence of candidate compounds to identify compounds which disrupt the interaction. Thus, for example, the assay may employ methods including (1) affinity chromatography; (2) immunoprecipitation; (3) the Biomolecular Interaction Assay (BIAcore); or (4) the yeast two-hybrid systems. Such assays can be developed using either normal or mutant purified PS-interacting proteins, and/or either normal or mutant purified binding proteins (e.g., normal or mutant presentlins).

For affinity methods, either the PS-interacting protein or its binding partner may be affixed to a matrix, for example in a column, and the counterpart protein (e.g., the PS-interacting protein if presentilin or another binding partner is affixed to the matrix; or a presentilin or other binding partner if the PS-interacting protein is affixed to the matrix) is then exposed to the affixed protein/compound either before or after adding the candidate compound(s). In the absence of a disruptive effect by the candidate compound(s), the interaction between the PS-interacting protein and its binding partner will cause the counterpart protein to bind to the affixed protein. Any compound which disrupts the interaction will cause release of the counterpart protein from the matrix. Release of the counterpart protein from the matrix can be measured using methods known in the art.

For PS-interacting protein interactions which are detectable by yeast two-hybrid systems, these assays may also be employed to identify compounds which disrupt the interaction. Briefly, a PS-interacting protein and its binding partner (or appropriate structural domains of each) are employed in the fusion proteins of the system, and the cells are exposed to candidate compounds to determine their effect upon the expression of the reporter gene. By appropriate choice of a reporter gene, such a system can be readily adapted for high through-put screening of large libraries of compounds by, for example, using a reporter gene which confers resistance to an antibiotic which is present in the medium, or which rescues an auxotrophic strain grown in minimal medium.

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These assays may be used to screen many different types of compounds for their disruptive effect on the interactions of the PS-interacting proteins. For example, the compounds may belong to a library of synthetic molecules, or be specifically designed to disrupt the interaction. The compounds may also be peptides corresponding to the interacting domain of either protein. This type of assay can be used to identify compounds that disrupt a specific interaction between a given PS-interacting protein variant and a given binding partner. In addition, compounds that disrupt all interactions with PS-interacting proteins may be identified. For example, a compound that specifically disrupts the folding of PS-interacting proteins would be expected to disrupt all interactions between PS-interacting proteins and other proteins. Alternatively, this type of disruption assay can be used to identify compounds which disrupt only a range of different PS-interacting protein interactions, or only a single PS-interacting protein interaction interaction.

9. Methods of Identifying Compounds Modulating PS-Interacting Protein Activity

In another series of embodiments, the present invention provides for methods of identifying compounds with the ability to modulate the activity of normal and mutant PS-interacting proteins. As used with respect to this series of embodiments, the term "activity" broadly includes gene and protein expression, PS-interacting protein post-translation processing, trafficking and localization, and any functional activity (e.g., enzymatic, receptor-effector, binding, channel), as well as downstream affects of any of these. It is known that Alzheimer's Disease is associated with increased production of the long form of Aβ peptides, the appearance of amyloid plaques and neurofibrillary tangles, decreases in cognitive function, and apoptotic cell death. Therefore, using the transformed cells and transgenic animal models of the present invention, cells obtained from subjects bearing normal or mutant PS-interacting protein genes, or animals or human subjects bearing naturally occurring normal or mutant PS-interacting proteins, it is now possible to screen candidate pharmaceuticals and treatments for their therapeutic effects by detecting changes in

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one or more of these functional characteristics or phenotypic manifestations of normal or mutant PS-interacting protein expression.

Thus, the present invention provides methods for screening or assaying for proteins, small molecules or other compounds which modulate PS-interacting protein activity by contacting a cell in vivo or in vitro with a candidate compound and assaying for a change in a marker associated with normal or mutant PS-interacting protein activity. The marker associated with PS-interacting protein activity may be any measurable biochemical, physiological, histological and/or behavioral characteristic associated with PS-interacting protein expression. In particular, useful markers will include any measurable biochemical, physiological, histological and/or behavioral characteristic which distinguishes cells, tissues, animals or individuals bearing at least one mutant presentlin or PS-interacting protein gene from their normal counterparts. In addition, the marker may be any specific or non-specific measure of presenilin or PS-interacting protein activity. PS-interacting protein specific measures include measures of PS-interacting protein expression (e.g., PS-interacting protein mRNA or protein levels) which may employ the nucleic acid probes or antibodies of the present invention. Non-specific measures include changes in cell physiology such as pH, intracellular calcium, cyclic AMP levels, GTP/GDP ratios, phosphatidylinositol activity, protein phosphorylation, etc., which can be monitored on devices such as the cytosensor microphysiometer (Molecular Devices Inc., United States). The activation or inhibition of PS-interacting protein activity in its mutant or normal form can also be monitored by examining changes in the expression of other genes (e.g., the presenilins) which are specific to the PS-interacting protein pathway leading to Alzheimer's Disease. These can be assayed by such techniques as differential display, differential hybridization, and SAGE (sequential analysis of gene expression), as well as by two dimensional gel electrophoresis of cellular lysates. In each case, the differentially-expressed genes can be ascertained by inspection of identical studies before and after application of the candidate compound. Furthermore, as noted elsewhere, the particular genes whose expression is modulated by the administration of the candidate compound can be ascertained by cloning,

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nucleotide sequencing, amino acid sequencing, or mass spectrometry (reviewed in Nowak, 1995).

In general, a cell may be contacted with a candidate compound and, after an appropriate period (e.g., 0-72 hours for most biochemical measures of cultured cells), the marker of presentilin or PS-interacting protein activity may be assayed and compared to a baseline measurement. The baseline measurement may be made prior to contacting the cell with the candidate compound or may be an external baseline established by other experiments or known in the art. The cell may be a transformed cell of the present invention or an explant from an animal or individual. In particular, the cell may be an explant from a carrier of a presentilin or PS-interacting protein mutation (e.g., a human subject with Alzheimer's Disease) or an animal model of the invention (e.g., a transgenic nematode or mouse bearing a mutant presentiin or PSinteracting protein gene). To augment the effect of presentiin or PS-interacting protein mutations on the AB pathway, transgenic cells or animals may be employed which have increased A\beta production. Preferred cells include those from neurological tissues such as neuronal, glial or mixed cell cultures; and cultured fibroblasts, liver, kidney, spleen, or bone marrow. The cells may be contacted with the candidate compounds in a culture in vitro or may be administered in vivo to a live animal or human subject. For live animals or human subjects, the test compound may be administered orally or by any parenteral route suitable to the compound. For clinical trials of human subjects, measurements may be conducted periodically (e.g., daily, weekly or monthly) for several months or years.

Because most individuals bearing a mutation in a particular gene are heterozygous at that locus (i.e., bearing one normal and one mutant allele), compounds may be tested for their ability to modulate normal as well as mutant presentilin or PS-interacting protein activity. Thus, for example, compounds which enhance the function of normal presentilins or PS-interacting proteins may have utility in treating Alzheimer's Disease or related disorders. Alternatively, because suppression of the activity of both normal and mutant copies of a gene in a heterozygous individual may have less severe clinical consequences than progression

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of the associated disease, it may be desired to identify compound which inactivate or suppress all forms of the presentilins, the PS-interacting proteins, or their interactions. Preferably, however, compounds are identified which selectively or specifically inactivate or suppress the activity of mutant presentilin or PS-interacting proteins without disrupting the function of their normal counterparts.

In light of the identification, characterization, and disclosure herein of a novel group of PS-interacting genes and proteins, the PS-interacting protein nucleic acid probes and antibodies, and the PS-interacting protein transformed cells and transgenic animals of the invention, one of ordinary skill in the art is now enabled by perform a great variety of assays which will detect the modulation of presentiin and/or PS-interacting protein activity by candidate compounds. Particularly preferred and contemplated embodiments are discussed in some detail below.

A. PS-Interacting Protein Expression

In one series of embodiments, specific measures of PS-interacting protein expression are employed to screen candidate compounds for their ability to affect presentilin activity. Thus, using the PS-interacting protein nucleic acids and antibodies disclosed and otherwise enabled herein, one may use mRNA levels or protein levels as a marker for the ability of a candidate compound to modulate PS-interacting protein activity. The use of such probes and antibodies to measure gene and protein expression is well known in the art and discussed elsewhere herein. Of particular interest may be the identification of compounds which can alter the relative levels of different variants (e.g., mutant and normal) of the PS-interacting proteins.

B. Intracellular Localization

In another series of embodiments, compounds may be screened for their ability to modulate the activity of the PS-interacting proteins based upon their effects on the trafficking and intracellular localization of the PS-interacting proteins. The presentilins and some of the PS-interacting proteins (e.g., S5a) have been seen immunocytochemically to be localized in membrane structures associated with the endoplasmic reticulum and Golgi apparatus. Differences in localization of mutant and normal presentilins or PS-interacting proteins may, therefore, contribute to the etiology

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of Alzheimer's Disease and related disorders. Compounds which can affect the localization of these proteins may, therefore, be identified as potential therapeutics. Standard techniques known in the art may be employed to detect the localization of the presentilins and PS-interacting proteins. Generally, these techniques will employ the antibodies of the present invention, and in particular antibodies which selectively bind to one or more mutant PS-interacting proteins but not to normal proteins. As is well known in the art, such antibodies may be labeled by any of a variety of techniques (e.g., fluorescent or radioactive tags, labeled secondary antibodies, avidinbiotin, etc.) to aid in visualizing the intracellular location of these proteins. The PS-interacting proteins may be co-localized to particular structures, as in known in the art, using antibodies to markers of those structures (e.g., TGN38 for the Golgi, transferrin receptor for post-Golgi transport vesicles, LAMP2 for lysosomes). Western blots of purified fractions from cell lysates enriched for different intracellular membrane bound organelles (e.g., lysosomes, synaptosomes, Golgi) may also be employed.

B. Ion Regulation/Metabolism

In another series of embodiments, compounds may be screened for their ability to modulate the activity of the presentlins or PS-interacting proteins based upon measures in intracellular Ca²⁺, Na⁺ or K⁺ levels or metabolism. As noted above, the presentlins are membrane associated proteins which may serve as, or interact with, ion receptors or ion channels. Thus, compounds may be screened for their ability to modulate presentlin and PS-interacting protein-related metabolism of calcium or other ions either in vivo or in vitro by, for example, measurements of ion channel fluxes and/or transmembrane voltage and/or current fluxes, using patch clamps, voltage clamps or fluorescent dyes sensitive to intracellular ion levels or transmembrane voltage. Ion channel or receptor function can also be assayed by measurements of activation of second messengers such as cyclic AMP, cGMP tyrosine kinases, phosphates, increases in intracellular Ca²⁺ levels, etc. Recombinantly made proteins may also be reconstructed in artificial membrane systems to study ion channel conductance and, therefore, the "cell" employed in such assays may comprise an

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artificial membrane or cell. Assays for changes in ion regulation or metabolism can be performed on cultured cells expressing endogenous normal or mutant presenilins and PS-interacting proteins. Such studies also can be performed on cells transfected with vectors capable of expressing one of the presenilins or PS-interacting proteins, or functional domains of one of the presenilins or PS-interacting proteins, in normal or mutant form. In addition, to enhance the signal measured in such assays, cells may be co-transfected with genes encoding ion channel proteins. For example, Xenopus oocytes or rat kidney (HEK293) cells may be co-transfected with sequences encoding rat brain Na⁺ β1 subunits, rabbit skeletal muscle Ca²⁺ β1 subunits, or rat heart K⁺ β1 subunits. Changes in presenilin or PS-interacting protein-mediated ion channel activity can be measured by, for example, two-microelectrode voltage-clamp recordings in oocytes, by whole-cell patch-clamp recordings in HEK293 cells. or by equivalent means.

C. Apoptosis or Cell Death

In another series of embodiments, compounds may be screened for their ability to modulate the activity of the presenilins or PS-interacting proteins based upon their effects on presenilin or PS-interacting protein-related apoptosis or cell death. Thus, for example, baseline rates of apoptosis or cell death may be established for cells in culture, or the baseline degree of neuronal loss at a particular age may be established post-mortem for animal models or human subjects, and the ability of a candidate compound to suppress or inhibit apoptosis or cell death may be measured. Cell death may be measured by standard microscopic techniques (e.g., light microscopy) or apoptosis may be measured more specifically by characteristic nuclear morphologies or DNA fragmentation patterns which create nucleosomal ladders (see, e.g., Gavrieli et al., 1992; Jacobson et al., 1993; Vito et al., 1996). TUNEL may also be employed to evaluate cell death in brain (see, e.g., Lassmann et al., 1995). In preferred embodiments, compounds are screened for their ability to suppress or inhibit neuronal loss in the transgenic animal models of the invention. Transgenic mice bearing, for example, a mutant human, mutant mouse, or humanized mutant presenilin or PS-interacting protein gene may be employed to identify or evaluate compounds

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which may delay or arrest the neurodegeneration associated with Alzheimer's Disease. A similar transgenic mouse model, bearing a mutant APP gene, has recently been reported by Games et al. (1995).

D. Aß Peptide Production

In another series of embodiments, compounds may be screened for their ability to modulate presentiin or PS-interacting protein-related changes in APP processing. The Aβ peptide is produced in several isoforms resulting from differences in APP processing. The AB peptide is a 39 to 43 amino acid derivative of BAPP which is progressively deposited in diffuse and senile plaques and in blood vessels of subjects with AD. In human brain, Aβ peptides are heterogeneous at both the N- and C-termini. Several observations, however, suggest that both the full length and Nterminal truncated forms of the long-tailed A\beta peptides ending at residue 42 or 43 (i.e., A\(\beta\)1-42/43 and A\(\beta\)x-42/43) have a more important role in AD than do peptides ending at residue 40. Thus, $A\beta 1-42/43$ and $A\beta x-42/43$ are an early and prominent feature of both senile plaques and diffuse plaques, while peptides ending at residue 40 (i.e., A\beta 1-40 and A\beta x-40) are predominantly associated with a subset of mature plaques and with amyloidotic blood vessels (see, e.g., Iwatsubo et al., 1995; Gravina et al., 1995; Tamaoka et al., 1995; Podlisny et al. 1995). Furthermore, the long-tailed isoforms have a greater propensity to fibril formation, and are thought to be more neurotoxic than Aβ1-40 peptides (Pike et al., 1993; Hilbich et al., 1991). Finally, missense mutations at codon 717 of the BAPP gene are associated with early onset FAD, and result in overproduction of long-tailed Aβ in the brain of affected mutation carriers, in peripheral cells and plasma of both affected and presymptomatic carriers, and in cell lines transfected with βAPP₇₁₇ mutant cDNAs (Tamaoka et al., 1994; Suzuki et al., 1994).

Thus, in one series of embodiments, the present invention provides methods for screening candidate compounds for their ability to block or inhibit the increased production of long isoforms of the Aß peptides in cells or transgenic animals expressing a normal or mutant presentlin gene and/or a normal or mutant PS-interacting protein gene. In particular, the present invention provides such methods in

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which cultured mammalian cells, such as brain cells or fibroblasts, have been transformed according to the methods disclosed herein, or in which transgenic animals, such as rodents or non-human primates, have been produced by the methods disclosed herein, to express relatively high levels of a normal or mutant presentil or PS-interacting protein. Optionally, such cells or transgenic animals may also be transformed so as to express a normal or mutant form of the β APP protein at relatively high levels.

In this series of embodiments, the candidate compound is administered to the cell line or transgenic animals (e.g., by addition to the media of cells in culture; or by oral or parenteral administration to an animal) and, after an appropriate period (e.g., 0-72 hours for cells in culture, days or months for animal models), a biological sample is collected (e.g., cell culture supernatant or cell lysate from cells in culture; tissue homogenate or plasma from an animal) and tested for the level of the long isoforms of the $A\beta$ peptides. The levels of the peptides may be determined in an absolute sense (e.g., nMol/ml) or in a relative sense (e.g., ratio of long to short $A\beta$ isoforms). The $A\beta$ isoforms may be detected by any means known in the art (e.g., electrophoretic separation and sequencing) but, preferably, antibodies which are specific to the long isoform are employed to determine the absolute or relative levels of the A β 1-42/43 or A β x-42/43 peptides. Candidate pharmaceuticals or therapies which reduce the absolute or relative levels of these long Aß isoforms, particularly in the transgenic animal models of the invention, are likely to have therapeutic utility in the treatment of Alzheimer's Disease, or other disorders caused by mutations in the presenilins or PS-interacting proteins, or by other aberrations in APP metabolism.

E. Phosphorylation of Microtubule Associated Proteins

In another series of embodiments, candidate compounds may be screened for their ability to modulate presentiin or PS-interacting protein activity by assessing the effect of the compound on levels of phosphorylation of microtubule associated proteins (MAPs) such as tau. The abnormal phosphorylation of tau and other MAPs in the brains of victims of Alzheimer's Disease is well known in the art. Thus, compounds which prevent or inhibit the abnormal phosphorylation of MAPs may

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have utility in treating presentilin or PS-interacting protein-associated diseases such as AD. As above, cells from normal or mutant animals or subjects, or the transformed cell lines and animal models of the invention may be employed. Preferred assays will employ cell lines or animal models transformed with a mutant human or humanized mutant presentlin or PS-interacting protein gene. The baseline phosphorylation state of MAPs in these cells may be established and then candidate compounds may be tested for their ability to prevent, inhibit or counteract the hyperphosphorylation associated with mutants. The phosphorylation state of the MAPs may be determined by any standard method known in the art but, preferably, antibodies which bind selectively to phosphorylated or unphosphorylated epitopes are employed. Such antibodies to phosphorylation epitopes of the tau protein are known in the art (e.g., ALZ50).

10. Screening and Diagnostics for Alzheimer's Disease

A. General Diagnostic Methods

The PS-interacting genes and gene products, as well as the PS-interacting protein derived probes, primers and antibodies, disclosed or otherwise enabled herein, are useful in the screening for carriers of alleles associated with Alzheimer's Disease, for diagnosis of victims of Alzheimer's Disease, and for the screening and diagnosis of related presentle and sentle dementias, psychiatric diseases such as schizophrenia and depression, and neurologic diseases such as stroke and cerebral hemorrhage, all of which are seen to a greater or lesser extent in symptomatic human subjects bearing mutations in the PS1 or PS2 genes or in the APP gene. Individuals at risk for Alzheimer's Disease, such as those with AD present in the family pedigree, or individuals not previously known to be at risk, may be routinely screened using probes to detect the presence of a mutant PS-interacting protein gene or protein by a variety of techniques. Diagnosis of inherited cases of these diseases can be accomplished by methods based upon the nucleic acids (including genomic and mRNA/cDNA sequences), proteins, and/or antibodies disclosed and enabled herein, including functional assays designed to detect failure or augmentation of the normal

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presentilin or PS-interacting protein activity and/or the presence of specific new activities conferred by mutant PS-interacting proteins. Preferably, the methods and products are based upon the human nucleic acids, proteins or antibodies, as disclosed or otherwise enabled herein. As will be obvious to one of ordinary skill in the art, however, the significant evolutionary conservation of large portions of nucleotide and amino acid sequences, even in species as diverse as humans, mice, <u>C. elegans</u>, and <u>Drosophila</u>, allow the skilled artisan to make use of non-human homologues of the PS-interacting proteins to produce useful nucleic acids, proteins and antibodies, even for applications directed toward human or other animal subjects. Thus, for brevity of exposition, but without limiting the scope of the invention, the following description will focus upon uses of the human homologues of PS-interacting proteins and genes. It will be understood, however, that homologous sequences from other species will be equivalent for many purposes.

As will be appreciated by one of ordinary skill in the art, the choice of diagnostic methods of the present invention will be influenced by the nature of the available biological samples to be tested and the nature of the information required. Alzheimer's Disease is, of course, primarily a disease of the brain, but brain biopsies are invasive and expensive procedures, particularly for routine screening. Other tissues which express the presentions or PS-interacting proteins at significant levels may, therefore, be preferred as sources for samples.

B. Protein Based Screens and Diagnostics

When a diagnostic assay is to be based upon PS-interacting proteins, a variety of approaches are possible. For example, diagnosis can be achieved by monitoring differences in the electrophoretic mobility of normal and mutant proteins. Such an approach will be particularly useful in identifying mutants in which charge substitutions are present, or in which insertions, deletions or substitutions have resulted in a significant change in the electrophoretic migration of the resultant protein. Alternatively, diagnosis may be based upon differences in the proteolytic cleavage patterns of normal and mutant proteins, differences in molar ratios of the

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various amino acid residues, or by functional assays demonstrating altered function of the gene products.

In preferred embodiments, protein-based diagnostics will employ differences in the ability of antibodies to bind to normal and mutant PS-interacting proteins. Such diagnostic tests may employ antibodies which bind to the normal proteins but not to mutant proteins, or vice versa. In particular, an assay in which a plurality of monoclonal antibodies, each capable of binding to a mutant epitope, may be employed. The levels of anti-mutant antibody binding in a sample obtained from a test subject (visualized by, for example, radiolabelling, ELISA or chemiluminescence) may be compared to the levels of binding to a control sample. Alternatively, antibodies which bind to normal but not mutant proteins may be employed, and decreases in the level of antibody binding may be used to distinguish homozygous normal individuals from mutant heterozygotes or homozygotes. Such antibody diagnostics may be used for in situ immunohistochemistry using biopsy samples of CNS tissues obtained antemortem or postmortem, including neuropathological structures associated with these diseases such as neurofibrillary tangles and amyloid plaques, or may be used with fluid samples such a cerebrospinal fluid or with peripheral tissues such as white blood cells.

C. Nucleic Acid Based Screens and Diagnostics

When the diagnostic assay is to be based upon nucleic acids from a sample, the assay may be based upon mRNA, cDNA or genomic DNA. When mRNA is used from a sample, there are considerations with respect to source tissues and the possibility of alternative splicing. That is, there may be little or no expression of transcripts unless appropriate tissue sources are chosen or available, and alternative splicing may result in the loss of some information or difficulty in interpretation. Whether mRNA, cDNA or genomic DNA is assayed, standard methods well known in the art may be used to detect the presence of a particular sequence either in situ or in vitro (see, e.g., Sambrook et al., (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Press, Cold Spring Harbor, NY). As a general matter, however, any tissue with nucleated cells may be examined.

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Genomic DNA used for the diagnosis may be obtained from body cells, such as those present in the blood, tissue biopsy, surgical specimen, or autopsy material. The DNA may be isolated and used directly for detection of a specific sequence or may be amplified by the polymerase chain reaction (PCR) prior to analysis. Similarly, RNA or cDNA may also be used, with or without PCR amplification. To detect a specific nucleic acid sequence, direct nucleotide sequencing, hybridization using specific oligonucleotides, restriction enzyme digest and mapping, PCR mapping, RNase protection, chemical mismatch cleavage, ligase-mediated detection, and various other methods may be employed. Oligonucleotides specific to particular sequences can be chemically synthesized and labeled radioactively or nonradioactively (e.g., biotin tags, ethidium bromide), and hybridized to individual samples immobilized on membranes or other solid-supports (e.g., by dot-blot or transfer from gels after electrophoresis), or in solution. The presence or absence of the target sequences may then be visualized using methods such as autoradiography, fluorometry, or colorimetry. These procedures can be automated using redundant, short oligonucleotides of known sequence fixed in high density to silicon chips.

(1) Appropriate Probes and Primers

Whether for hybridization, RNase protection, ligase-mediated detection, PCR amplification or any other standards methods described herein and well known in the art, a variety of subsequences of the PS-interacting protein sequences disclosed or otherwise enabled herein will be useful as probes and/or primers. These sequences or subsequences will include both normal sequences and deleterious mutant sequences. In general, useful sequences will include at least 8-9, more preferably 10-50, and most preferably 18-24 consecutive nucleotides from introns, exons or intron/exon boundaries. Depending upon the target sequence, the specificity required, and future technological developments, shorter sequences may also have utility. Therefore, any PS-interacting protein derived sequence which is employed to isolate, clone, amplify, identify or otherwise manipulate a PS-interacting protein sequence may be regarded as an appropriate probe or primer. Particularly contemplated as useful will be sequences

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including nucleotide positions from the PS-interacting protein genes in which diseasecausing mutations are known to be present, or sequences which flank these positions.

(2) Hybridization Screening

For in situ detection of a normal or mutant PS-interacting protein-related nucleic acid sequence, a sample of tissue may be prepared by standard techniques and then contacted with one or more of the above-described probes, preferably one which is labeled to facilitate detection, and an assay for nucleic acid hybridization is conducted under stringent conditions which permit hybridization only between the probe and highly or perfectly complementary sequences. Because many mutations consist of a single nucleotide substitution, high stringency hybridization conditions may be required to distinguish normal sequences from most mutant sequences. When the PS-interacting protein genotypes of the subject's parents are known, probes may be chosen accordingly. Alternatively, probes to a variety of mutants may be employed sequentially or in combination. Because most individuals carrying mutations in the PS-interacting proteins will be heterozygous, probes to normal sequences also may be employed and homozygous normal individuals may be distinguished from mutant heterozygotes by the amount of binding (e.g., by intensity of radioactive signal). In another variation, competitive binding assays may be employed in which both normal and mutant probes are used but only one is labeled.

(3) Restriction Mapping

Sequence alterations may also create or destroy fortuitous restriction enzyme recognition sites which are revealed by the use of appropriate enzyme digestion followed by gel-blot hybridization. DNA fragments carrying the site (normal or mutant) are detected by their increase or reduction in size, or by the increase or decrease of corresponding restriction fragment numbers. Such restriction fragment length polymorphism analysis (RFLP), or restriction mapping, may be employed with genomic DNA, mRNA or cDNA. The PS-interacting protein sequences may be amplified by PCR using the above-described primers prior to restriction, in which case the lengths of the PCR products may indicate the presence or absence of particular restriction sites, and/or may be subjected to restriction after amplification.

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The restriction fragments may be visualized by any convenient means (e.g., under UV light in the presence of ethidium bromide).

(4) PCR Mapping

In another series of embodiments, a single base substitution mutation may be detected based on differential PCR product length or production in PCR. Thus, primers which span mutant sites or which, preferably, have 3' termini at mutation sites, may be employed to amplify a sample of genomic DNA, mRNA or cDNA from a subject. A mismatch at a mutational site may be expected to alter the ability of the normal or mutant primers to promote the polymerase reaction and, thereby, result in product profiles which differ between normal subjects and heterozygous and/or homozygous mutants. The PCR products of the normal and mutant gene may be differentially separated and detected by standard techniques, such as polyacrylamide or agarose gel electrophoresis and visualization with labeled probes, ethidium bromide or the like. Because of possible non-specific priming or readthrough of mutation sites, as well as the fact that most carriers of mutant alleles will be heterozygous, the power of this technique may be low.

(5) Electrophoretic Mobility

Genetic testing based on DNA sequence differences also may be achieved by detection of alterations in electrophoretic mobility of DNA, mRNA or cDNA fragments in gels. Small sequence deletions and insertions, for example, can be visualized by high resolution gel electrophoresis of single or double stranded DNA, or as changes in the migration pattern of DNA heteroduplexes in non-denaturing gel electrophoresis. Mutations or polymorphisms in the PS-interacting protein genes may also be detected by methods which exploit mobility shifts due to single-stranded conformational polymorphisms (SSCP) associated with mRNA or single-stranded DNA secondary structures.

(6) Chemical Cleavage of Mismatches

Mutations in the PS-interacting protein genes may also be detected by employing the chemical cleavage of mismatch (CCM) method (see, e.g., Saleeba and Cotton, 1993, and references therein). In this technique, probes (up to ~ 1 kb) may be

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mixed with a sample of genomic DNA, cDNA or mRNA obtained from a subject. The sample and probes are mixed and subjected to conditions which allow for heteroduplex formation (if any). Preferably, both the probe and sample nucleic acids are double-stranded, or the probe and sample may be PCR amplified together, to ensure creation of all possible mismatch heteroduplexes. Mismatched T residues are reactive to osmium tetroxide and mismatched C residues are reactive to hydroxylamine. Because each mismatched A will be accompanied by a mismatched T, and each mismatched G will be accompanied by a mismatched C, any nucleotide differences between the probe and sample (including small insertions or deletions) will lead to the formation of at least one reactive heteroduplex. After treatment with osmium tetroxide and/or hydroxylamine to modify any mismatch sites, the mixture is subjected to chemical cleavage at any modified mismatch sites by, for example, reaction with piperidine. The mixture may then be analyzed by standard techniques such as gel electrophoresis to detect cleavage products which would indicate mismatches between the probe and sample.

(7) Other Methods

Various other methods of detecting PS-interacting protein mutations, based upon the sequences disclosed and otherwise enabled herein, will be apparent to those of ordinary skill in the art. Any of these may be employed in accordance with the present invention. These include, but are not limited to, nuclease protection assays (S1 or ligase-mediated), ligated PCR, denaturing gradient gel electrophoresis (DGGE; see, e.g., Fischer and Lerman, 1983), restriction endonuclease fingerprinting combined with SSCP (REF-SSCP; see, e.g., Liu and Sommer, 1995), and the like.

D. Other Screens and Diagnostics

In inherited cases, as the primary event, and in non-inherited cases as a secondary event due to the disease state, abnormal processing of the presentions, PS-interacting proteins, APP, or proteins reacting with the presentions, PS-interacting proteins, or APP may occur. This can be detected as abnormal phosphorylation, glycosylation, glycation amidation or proteolytic cleavage products in body tissues or fluids (e.g., CSF or blood).

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Diagnosis also can be made by observation of alterations in transcription, translation, and post-translational modification and processing, as well as alterations in the intracellular and extracellular trafficking of gene products in the brain and peripheral cells. Such changes will include alterations in the amount of messenger RNA and/or protein, alteration in phosphorylation state, abnormal intracellular location/distribution, abnormal extracellular distribution, etc. Such assays will include: Northern Blots (e.g., with PS-interacting protein-specific and non-specific nucleotide probes), Western blots and enzyme-linked immunosorbent assays (ELISA) (e.g., with antibodies raised specifically to a PS-interacting protein or PS-interacting functional domain, including various post-translational modification states including glycosylated and phosphorylated isoforms). These assays can be performed on peripheral tissues (e.g., blood cells, plasma, cultured or other fibroblast tissues, etc.) as well as on biopsies of CNS tissues obtained antemortem or postmortem, and upon cerebrospinal fluid. Such assays might also include in situ hybridization and immunohistochemistry (to localize messenger RNA and protein to specific subcellular compartments and/or within neuropathological structures associated with these diseases such as neurofibrillary tangles and amyloid plaques).

E. Screening and Diagnostic Kits

In accordance with the present invention, diagnostic kits are also provided which will include the reagents necessary for the above-described diagnostic screens. For example, kits may be provided which include antibodies or sets of antibodies which are specific to one or more mutant epitopes. These antibodies may, in particular, be labeled by any of the standard means which facilitate visualization of binding. Alternatively, kits may be provided in which oligonucleotide probes or PCR primers, as described above, are present for the detection and/or amplification of normal or mutant presenilin and/or PS-interacting protein nucleotide sequences. Again, such probes may be labeled for easier detection of specific hybridization. As appropriate to the various diagnostic embodiments described above, the oligonucleotide probes or antibodies in such kits may be immobilized to substrates and appropriate controls may be provided.

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11. Methods of Treatment

The present invention now provides a basis for therapeutic intervention in diseases which are caused, or which may be caused, by mutations in the PS-interacting proteins. As noted above, mutations in the hPS1 and hPS2 genes have been associated with the development of early onset forms of Alzheimer's Disease and, therefore, the present invention is particularly directed to the treatment of subjects diagnosed with, or at risk of developing, Alzheimer's Disease.

Without being bound to any particular theory of the invention, the effect of the Alzheimer's Disease related mutations in the presenilins appears to be a gain of a novel function, or an acceleration of a normal function, which directly or indirectly causes aberrant processing of the Amyloid Precursor Protein (APP) into Aß peptide, abnormal phosphorylation homeostasis, and/or abnormal apoptosis in the brain. Such a gain of function or acceleration of function model would be consistent with the adult onset of the symptoms and the dominant inheritance of Alzheimer's Disease. Nonetheless, the mechanism by which mutations in the presentlins may cause these effects remains unknown.

The present invention, by identifying a set of PS-interacting proteins, provides new therapeutic targets for intervening in the etiology of presenilin-related AD. In addition, as mutations in the presenilins may cause AD, it is likely that mutations in the PS-interacting proteins may also cause AD. The fact that the PS-interacting protein S5a is alternately processed in the brains of victims of sporadic AD, as well as in the brains of victims of presenilin-linked AD, suggests that, at the very least, this PS-interacting protein is involved in the etiology of AD independent of mutations in the presenilins. It is likely that the other PS-interacting proteins also may be involved in non-presenilin-linked AD.

Therapies to treat PS-interacting protein-associated diseases such as AD may be based upon (1) administration of normal PS-interacting proteins, (2) gene therapy with normal PS-interacting protein genes to compensate for or replace the mutant genes, (3) gene therapy based upon antisense sequences to mutant PS-

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interacting protein genes or which "knock-out" the mutant genes, (4) gene therapy based upon sequences which encode a protein which blocks or corrects the deleterious effects of PS-interacting protein mutants, (5) immunotherapy based upon antibodies to normal and/or mutant PS-interacting proteins, or (6) small molecules (drugs) which alter PS-interacting protein expression, alter interactions between PS-interacting proteins and other proteins or ligands, or which otherwise block the aberrant function of mutant presentilin or PS-interacting proteins by altering the structure of the mutant proteins, by enhancing their metabolic clearance, or by inhibiting their function.

A. Protein Therapy

Treatment of Alzheimer's Disease, or other disorders resulting from PS-interacting protein mutations, may be performed by replacing the mutant protein with normal protein, by modulating the function of the mutant protein, or by providing an excess of normal protein to reduce the effect of any aberrant function of the mutant proteins.

To accomplish this, it is necessary to obtain, as described and enabled herein, large amounts of substantially pure PS-interacting protein from cultured cell systems which can express the protein. Delivery of the protein to the affected brain areas or other tissues can then be accomplished using appropriate packaging or administration systems including, for example, liposome mediated protein delivery to the target cells.

B. Gene Therapy

In one series of embodiments, gene therapy may be employed in which normal copies of a PS-interacting protein gene are introduced into patients to code successfully for normal protein in one or more different affected cell types. The gene must be delivered to those cells in a form in which it can be taken up and code for sufficient protein to provide effective function. Thus, it is preferred that the recombinant gene be operably joined to a strong promoter so as to provide a high level of expression which will compensate for, or out-compete, the mutant proteins. As noted above, the recombinant construct may contain endogenous or exogenous

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regulatory elements, inducible or repressible regulatory elements, or tissue-specific regulatory elements.

In another series of embodiments, gene therapy may be employed to replace the mutant gene by homologous recombination with a recombinant construct. The recombinant construct may contain a normal copy of the targeted PS-interacting protein gene, in which case the defect is corrected in situ, or may contain a "knockout" construct which introduces a stop codon, missense mutation, or deletion which abolished function of the mutant gene. It should be noted in this respect that such a construct may knock-out both the normal and mutant copies of the targeted gene in a heterozygous individual, but the total loss of gene function may be less deleterious to the individual than continued progression of the disease state.

In another series of embodiments, antisense gene therapy may be employed. The antisense therapy is based on the fact that sequence-specific suppression of gene expression can be achieved by intracellular hybridization between mRNA or DNA and a complementary antisense species. The formation of a hybrid duplex may then interfere with the transcription of the gene and/or the processing, transport, translation and/or stability of the target mRNA. Antisense strategies may use a variety of approaches including the administration of antisense oligonucleotides or antisense oligonucleotide analogs (e.g., analogs with phosphorothioate backbones) or transfection with antisense RNA expression vectors. Again, such vectors may include exogenous or endogenous regulatory regions, inducible or repressible regulatory elements, or tissue-specific regulatory elements.

In another series of embodiments, gene therapy may be used to introduce a recombinant construct encoding a protein or peptide which blocks or otherwise corrects the aberrant function caused by a mutant presentilin or PS-interacting protein gene. In one embodiment, the recombinant gene may encode a peptide which corresponds to a domain of a PS-interacting which has been found to abnormally interact with another cell protein or other cell ligand (e.g., a mutant presentilin). Thus, for example, if a mutant PS1 TM6 \rightarrow 7 domain is found to interact with a PS-interacting protein but the corresponding normal TM6 \rightarrow 7 domain does not undergo

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this interaction, gene therapy may be employed to provide an excess of the mutant TM6→7 domain which may compete with the mutant presentiin protein and inhibit or block the aberrant interaction. Alternatively, the PS-interacting domain of a PS-interacting protein which interacts with a mutant, but not a normal, presentiin may be encoded and expressed by a recombinant construct in order to compete with, and thereby inhibit or block, the aberrant interaction.

Retroviral vectors can be used for somatic cell gene therapy especially because of their high efficiency of infection and stable integration and expression. A full length PS-interacting protein gene, subsequences encoding functional domains of these proteins, or any of the other therapeutic peptides described above, can be cloned into a retroviral vector and expression may be driven from its endogenous promoter, from the retroviral long terminal repeat, or from a promoter specific for the target cell type of interest (e.g., neurons). Other viral vectors which can be used include adenoassociated virus, vaccinia virus, bovine papilloma virus, or a herpes virus such as Epstein-Barr virus.

C. Immunotherapy

Immunotherapy is also possible for Alzheimer's Disease. Antibodies may be raised to a normal or mutant PS-interacting protein (or a portion thereof) and are administered to the patient to bind or block an aberrant interaction (e.g., with a mutant presential) and prevent its deleterious effects. Simultaneously, expression of the normal protein product could be encouraged. Alternatively, antibodies may be raised to specific complexes between mutant or wild-type PS-interacting proteins and their interaction partners.

A further approach is to stimulate endogenous antibody production to the desired antigen. Administration could be in the form of a one time immunogenic preparation or vaccine immunization. The PS-interacting protein or other antigen may be mixed with pharmaceutically acceptable carriers or excipients compatible with the protein. The immunogenic composition and vaccine may further contain auxiliary substances such as emulsifying agents or adjuvants to enhance effectiveness.

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Immunogenic compositions and vaccines may be administered parenterally by injection subcutaneously or intramuscularly.

D. Small Molecule Therapeutics

As described and enabled herein, the present invention provides for a number of methods of identifying small molecules or other compounds which may be useful in the treatment of Alzheimer's Disease or other disorders caused by mutations in the presentilins or PS-interacting proteins. Thus, for example, the present invention provides for methods of identifying proteins which bind to normal or mutant PS-interacting proteins (aside from the presentilins). The invention also provides for methods of identifying small molecules which can be used to disrupt aberrant interactions between mutant presentilins and/or PS-interacting proteins and such other binding proteins or other cell components.

Examples

15 Example 1. Isolation of PS-interacting proteins by two-hybrid yeast system.

To identify proteins interacting with the presentilin proteins, a commercially available yeast two-hybrid kit ("Matchmaker System 2" from Clontech, Palo Alto, CA) was employed to screen a brain cDNA library for clones which interact with functional domains of the presentilins. In view of the likelihood that the TM6→7 loop domains of the presentilins are important functional domains, partial cDNA sequences encoding either residues 266-409 of the normal PS1 protein or residues 272-390 of the normal PS2 protein were ligated in-frame into the EcoRI and BamHI sites of the pAS2-1 fusion-protein expression vector (Clontech). The resultant fusion proteins contain the GAL4 DNA binding domain coupled in-frame either to the TM6→7 loop of the PS1 protein or to the TM6→7 loop of the PS2 protein. These expression plasmids were co-transformed into S. cerevisiae strain Y190 together with a library of human brain cDNAs ligated into the pACT2 yeast fusion-protein expression vector (Clontech) bearing the GAL4 activation domain using modified lithium acetate protocols of the "Matchmaker System 2" yeast two-hybrid kit (Clontech, Palo Alto, CA). Yeast clones bearing human brain cDNAs which interact

with the TM6 \rightarrow 7 loop domain were selected for His- resistance by plating on SD minimal medium lacking histidine and for β gal+ activation by color selection. The His+ β gal+ clones were then purged of the pAS2-1 "bait" construct by culture in 10µg/ml cyclohexamide and the unknown "trapped" inserts of the human brain cDNAs encoding PS-interacting proteins were isolated by PCR and sequenced. Of 6 million initial transformants, 200 positive clones were obtained after His- selection, and 42 after β gal+ color selection, carried out in accordance with the manufacturer's protocol for selection of positive colonies. Of these 42 clones there were several independent clones representing the same genes.

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through the acquisition of a novel but toxic function (i.e., dominant gain of function mutation) which is mediated by a novel interaction between the mutant proteins and one or more other cellular proteins, the human brain cDNA library cloned into the pACT2 expression vector (Clontech) was re-screened using mutant TM6→7 loop domain sequences as described above and according to manufacturer's protocols. In particular, mutant presenilin sequences corresponding to residues 260-409 of PS1 TM6→7 loop domains bearing mutations L286V, L392V and Δ290-319 were ligated in-frame into the GAL4 DNA-binding domain of the pAS2-1 vector (Clontech) and used to screen the human brain cDNA:GAL4 activation domain library of pACT vectors (Clontech). Yeast were co-transformed, positive colonies were selected, and "trapped" sequences were recovered and sequenced as described above. In addition to some of the same sequences recovered with the normal TM6→7 loop domains, several new sequences were obtained which reflect aberrant interactions of the mutant presenilins with normal cellular proteins.

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The recovered and sequenced clones corresponding to these PS-interacting proteins were compared to the public sequence databases using the BL:ASTN algorithm via the NCBI e-mail server. Descriptions of several of these clones follow:

Antisecretory Factor/ Proteasome S5a Subunit. Two overlapping clones (Y2H29 and Y2H31) were identified which correspond to a C-terminal fragment of a protein alternatively identified as Antisecretory Factor ("ASF") or the Multiubiquitin

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chain binding S5a subunit of the 26S proteasome ("S5a") (Johansson et al. 1995; Ferrell et al., 1996). The complete nucleotide and amino acid sequences of the S5a subunit are available through the public databases under Accession number U51007 and are reproduced here as SEQ ID NO: 1 and SEQ ID NO: 2. The nucleotide sequences of the Y2H29 and Y2H31 clones include nucleotides 351-1330 of SEQ ID NO: 1 and amino acid residues 70-377 of SEQ ID NO: 2. Thus, residues 70-377 of the full S5a subunit include the PS-interacting domain of this protein. Residues 206-377 of S5a contain certain motifs that are important for protein-protein interactions (Ferrell et al., 1996).

The PS1-S5a subunit interaction was directly re-tested for both wild type and mutant PS1 TM6→7 loop (residues 260-409) by transforming Y187 yeast cells with the appropriate wild type or mutant (L286V, L392V or Δ290-319) cDNA ligated in-frame to the GAL4-DNA binding domain of pACT2. The Δ290-319 mutant fusion construct displayed autonomous βgal activation in the absence of any S5a "target sequence" and, therefore, could not be further analyzed. In contrast, both the L286V and L392V mutant constructs interacted specifically with the S5a construct. Quantitative assays, however, showed that these interactions were weaker than those involving the wild type PS1₂₆₀₋₄₀₉ sequence and that the degree of interaction was crudely correlated with the age of onset of FAD. The difference in βgal activation was not attributable to instability of the mutant PS1₂₆₀₋₄₀₉ construct mRNAs or fusion proteins because Western blots of lysates of transformed yeast showed equivalent quantities of mutant or wild-type fusion proteins.

Because one of the putative functions of S5a is to bind multi-ubiquitinated proteins, the PS1:S5a interaction observed in S. cerevisiae could arise either through yeast-dependent ubiquitination of the PS1₂₆₀₋₄₀₉ construct, or by direct interaction. The former would reflect a degradative pathway, a functional and perhaps reciprocal interaction between PS1 and S5a, or both. A direct interaction is favored by the fact that the PS1:S5a interaction is decreased rather than increased by the presence of the L286V and L392V mutations, and by the fact that neither of these mutations affect ubiquitin conjugation sites in the PS1₂₆₀₋₄₀₉ loop (i.e., K265, K311, K314 or K395).

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To further examine this possibility, we investigated the direct interaction of recombinant His-tagged fusion proteins corresponding to full length S5a and the PS1₂₆₀₋₄₀₉ loop. Partially purified recombinant His-tagged PS1₂₆₀₋₄₀₉ loop and His-tagged S5a proteins and appropriate controls were mixed in phosphate buffered saline. The mixture was then subjected to size exclusion chromatography, and eluates were examined by SDS-PAGE and Western blotting using anti-His-tag monoclonal antibodies (Quiagen). In the crude PS1₂₆₀₋₄₀₉ loop preparation alone, the PS1₂₆₀₋₄₀₉ loop eluted from the size exclusion column as a broad peak at 35 minutes. In the crude S5a preparation alone, S5a eluted at 25 minutes. However, when the crude PS1₂₆₀₋₄₀₉ loop and S5a preparations were mixed, there was a significant shift in the elution of PS1₂₆₀₋₄₀₉ toward a higher molecular weight complex. Co-elution of S5a and PS1₂₆₀₋₄₀₉ in the same fraction was confirmed by SDS-PAGE and Western blotting of fractions using the anti-His-tag antibody. These results are consistent with a ubiquitin-independent and, therefore, possibly functional interaction.

GT24 and related genes with homology to p120/plakoglobin family. Five over-lapping clones (Y2H6, Y2H10b, Y2H17h2, Y2H24, and Y2H25) were obtained which interact with the normal PS1 TM6→7 loop domain and which appear to represent at least one novel gene. The Y2H24 clone was also found to interact with the mutant PS1 TM6→7 loop domains. Note that it appears that more than one member of the gene family was isolated, suggesting a family of genes interacting differentially with different presenilins. The most complete available cDNA corresponding to these clones was designated GT24 and is disclosed herein as SEQ ID NO: 3 and has been deposited with GenBank as Accession number U81004. The open reading frame suggests that GT24 is a protein of at least 1040 amino acids with a unique N-terminus, and considerable homology to several armadillo (arm) repeat proteins at its C-terminus. The predicted amino acid sequence of GT24 is disclosed herein as SEQ ID NO: 4. Thus, for example, residues 440-862 of GT24 have 32-56% identity (p=1.2e⁻¹³³) to residues 440-854 of murine p120 protein (Accession number Z17804), and residues 367-815 of GT24 have 26-42% identity (p=0.0017) to residues 245-465 of the D. melanogaster armadillo segment polarity protein (Accession

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number P18824). The GT24 gene maps to chromosome 5p15 near the anonymous microsatellite marker D5S748 and the Cri-du-Chat syndrome locus. This sequence is also nearly identical to portions of two human ESTs of unknown function (i.e., nucleotides 2701-3018 of Accession number F08730 and nucleotides 2974-3348 of Accession number T18858). These clones also show lower degrees of homology with other partial cDNA and gDNA sequences (e.g., H17245, T06654, T77214, H24294, M62015, T87427 and G04019).

p0071 gene. An additional His, βgal* clone isolated in the initial screening with wild type PS1₂₆₆₋₄₀₉ "bait" had a similar nucleotide sequence to GT24 (target clone Y2H25; Accession number U81005), and would also be predicted to encode a peptide with C-terminal arm repeats. A longer cDNA sequence closely corresponding to the Y2H25 clone has been deposited in GenBank as human protein p0071 (Accession number X81889). The nucleotide and corresponding amino acid sequences of p0071 are reproduced herein as SEQ ID NOs: 5 and 6. Comparison of the predicted sequence of the p0071 ORF with that of GT24 confirms that they are related proteins with 47% overall amino acid sequence identity, and with 70% identity between residues 346-862 of GT24, and residues 509-1022 of p0071 (which includes residues encoded by the Y2H25 cDNA). The latter result strongly suggests that PS1 interacts with a novel class of arm repeat containing proteins. The broad ~ 4 kb hybridization signal obtained on Northern blots with the unique 5' end of GT24 could reflect either alternate splicing/polyadenylation of GT24, or the existence of additional members of this family with higher degrees of N-terminal homology to GT24 than p0071.

Rab11 gene. This clone (Y2H9), disclosed herein as SEQ ID NO: 7, was identified as interacting with the normal PS1 TM6 \rightarrow 7 loop domain and appears to correspond to a known gene, Rab11, available through Accession numbers X56740 and X53143. Rab11 is believed to be involved in protein/vesicle trafficking in the ER/Golgi. Note the possible relationship to processing of membrane proteins such as BAPP and Notch with resultant overproduction of toxic Aß peptides (especially neurotoxic Aß₁₋₄₂₍₄₃₎ isoforms) (Scheuner, et al, 1995).

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Retinoid X receptor- β gene. This clone (Y2H23b), disclosed herein as SEQ ID NO: 8, was identified as interacting with the normal PS1 TM6 \rightarrow 7 loop domain and appears to correspond to a known gene, known variously as the retinoid X receptor- β , nuclear receptor co-regulator or MHC Class I regulatory element, and available through Accession numbers M84820, X63522 and M81766. This gene is believed to be involved in intercellular signaling, suggesting a possible relationship to the intercellular signaling function mediated by C. elegans sel12 and Notch/lin-12 (transcription activator).

Cytoplasmic chaperonin gene. This clone (Y2H27), disclosed herein as SEQ ID NO: 9, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a known gene, a cytoplasmic chaperonin containing TCP-1, available through Accession numbers U17104 and X74801.

Unknown gene (Y2H35). This clone (Y2H35), disclosed herein as SEQ ID NO: 10, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a known gene of unknown function, available through Accession number R12984, which shows conservation down through yeast.

Unknown gene (Y2H171). This clone (Y2H171), disclosed herein as SEQ ID NO: 11, was identified as interacting with the normal PS1 TM6→7 loop domain and appears to correspond to a known expressed repeat sequence available through Accession number D55326.

Unknown gene (Y2H41). This clone (Y2H41) was identified which reacts strongly with the TM6→7 loop domains of both PS1 and PS2 as well as the mutant loop domains of PS1. The sequence, disclosed as SEQ ID NO: 12, shows strong homology to an EST of unknown function (Accession number T64843).

25 Example 2. Isolation of presentilin binding proteins by affinity chromatography.

To identify the proteins which may be involved in the biochemical function of the presentilins, PS-interacting proteins were isolated using affinity chromatography. A GST-fusion protein containing the PS1 TM6→7 loop, prepared as described in Example 3, was used to probe human brain extracts, prepared by homogenizing brain tissue by Polytron in physiological salt solution. Non-specific

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binding was eliminated by pre-clearing the brain homogenates of endogenous GST-binding components by incubation with glutathione-Sepharose beads. These GST-free homogenates were then incubated with the GST-PS fusion proteins to produce the desired complexes with functional binding proteins. These complexes were then recovered using the affinity glutathione-Sepharose beads. After extensive washing with phosphate buffered saline, the isolated collection of proteins was separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE; Tris-tricine gradient gel 4-20%). Two major bands were observed at ~14 and 20 kD in addition to several weaker bands ranging from 50 to 60 kD.

The same approach may now be used to identify proteins which have binding activity for the PS-interacting proteins and, thereby, to further elucidate the etiology of AD and to identify additional therapeutics targets for intervention in AD and related disorders.

Example 3. Eukaryotic and prokaryotic expression vector systems.

Constructs suitable for use in eukaryotic and prokaryotic expression systems have been generated using different classes of PS1 nucleotide cDNA sequence inserts. In the first class, termed full-length constructs, the entire PS1 cDNA. sequence is inserted into the expression plasmid in the correct orientation, and includes both the natural 5' UTR and 3' UTR sequences as well as the entire open reading frame. The open reading frames bear a nucleotide sequence cassette which allows either the wild type open reading frame to be included in the expression system or alternatively, single or a combination of double mutations can be inserted into the open reading frame. This was accomplished by removing a restriction fragment from the wild type open reading frame using the enzymes NarI and PflmI and replacing it with a similar fragment generated by reverse transcriptase PCR and bearing the nucleotide sequence encoding either the M146L mutation or the H163R mutation. A second restriction fragment was removed from the wild type normal nucleotide sequence for the open reading frame by cleavage with the enzymes PflmI and NcoI and replaced with a restriction fragment bearing the nucleotide sequence encoding the A246E mutation, the A260V mutation, the A285V mutation, the L286V mutation, the

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L392V mutation or the C410Y mutation. A third variant, bearing a combination of either the M146L or H163R mutation in tandem with one of the remaining mutations, was made by linking a NarI-PflmI fragment bearing one of the former mutations and a PflmI-NcoI fragment bearing one of the latter mutations.

The second class of cDNA inserts, termed truncated constructs, was constructed by removing the 5' UTR and part of the 3' UTR sequences from full length wild type or mutant cDNA sequences. The 5' UTR sequence was replaced with a synthetic oligonucleotide containing a KpnI restriction site (GGTAC/C) and a small sequence (GCCACC) to create a Kozak initiation site around the ATG at the beginning of the PS1 ORF. The 3' UTR was replaced with an oligonucleotide with an artificial EcoRI site at the 5' end. Mutant variants of this construct were then made by inserting the mutant sequences described above at the NarI-PflmI and PsImI-NcoI sites as described above.

For eukaryotic expression, these various cDNA constructs bearing wild type and mutant sequences, as described above, were cloned into the expression vector pZeoSV in which the SV60 promoter cassette had been removed by restriction digestion and replaced with the CMV promoter element of pcDNA3 (Invitrogen). For prokaryotic expression, constructs have been made using the glutathione S-transferase (GST) fusion vector pGEX-kg. The inserts which have been attached to the GST fusion nucleotide sequence are the same nucleotide sequences described above bearing either the normal open reading frame nucleotide sequence, or bearing a combination of single and double mutations as described above. These GST fusion constructs allow expression of the partial or full-length protein in prokaryotic cell systems as mutant or wild type GST fusion proteins, thus allowing purification of the full-length protein followed by removal of the GST fusion product by thrombin digestion. A further cDNA construct was made with the GST fusion vector, to allow the production of the amino acid sequence corresponding to the hydrophilic acidic loop domain between TM6 and TM7 of the full-length protein, either as a wild type nucleotide sequence or as a mutant sequence bearing either the A285V mutation, the L286V mutation or the L392V mutation. This was accomplished by recovering wild

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type or mutant sequence from appropriate sources of RNA using a 5' oligonucleotide primer with a 5' BamHI restriction site (G/GATCC), and a 3' primer with a 5' EcoRI restriction site (G/AATTC). This allowed cloning of the appropriate mutant or wild type nucleotide sequence corresponding to the hydrophilic acidic loop domain at the BamHI and the EcoRI sites within the pGEX-KG vector.

The PS-interacting protein genes may be similarly manipulated by recombinant means for expression in prokaryotic or eukaryotic hosts. In particular, GST or other fusion proteins may be produced which will be useful in assays (e.g., yeast two-hybrid studies) for therapeutics.

10 Example 4. Antibody production.

Peptide antigens corresponding to portions of the PS1 protein were synthesized by solid-phase techniques and purified by reverse phase high pressure liquid chromatography. Peptides were covalently linked to keyhole limpet hemocyanin (KLH) via disulfide linkages that were made possible by the addition of a cysteine residue at the peptide C-terminus of the presentilin fragment. This additional residue does not appear normally in the protein sequence and was included only to facilitate linkage to the KLH molecule.

A total of three New Zealand white rabbits were immunized with peptide-KLH complexes for each peptide antigen in combination with Freund's adjuvant and were subsequently given booster injections at seven day intervals. Antisera were collected for each peptide and pooled and IgG precipitated with ammonium sulfate. Antibodies were then affinity purified with Sulfo-link agarose (Pierce) coupled with the appropriate peptide. This final purification is required to remove non-specific interactions of other antibodies present in either the pre- or post-immune serum.

The specificity of each antibody was confirmed by three tests. First, each detected single predominant bands of the approximate size predicted for presenting on Western blots of brain homogenate. Second, each cross-reacted with recombinant fusion proteins bearing the appropriate sequence. Third each could be specifically blocked by pre-absorption with recombinant PS1 or the immunizing peptide.

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Antibodies to peptides derived from the PS-interacting proteins may be produced by similar means.

Example 5. Transgenic mice.

A series of wild type and mutant PS1 and PS2 genes were constructed for use in the preparation of transgenic mice. Mutant versions of PS1 and PS2 were generated by site-directed mutagenesis of the cloned cDNAs using standard techniques.

The cDNAs and their mutant versions were used to prepare two classes of mutant and wild type PS1 and PS2 cDNAs, as described in Example 3. The first class, referred to as "full-length" cDNAs, were prepared by removing approximately 200 bp of the 3' untranslated region immediately before the polyA site by digestion with EcoRI (PS1) or PvuII (PS2). The second class, referred to as "truncated" cDNAs, were prepared by replacing the 5' untranslated region with a ribosome binding site (Kozak consensus sequence) placed immediately 5' of the ATG start codon.

Various full length and truncated wild type and mutant PS1 and PS2 cDNAs, prepared as described above, were introduced into one or more of the following vectors and the resulting constructs were used as a source of gene for the production of transgenic mice.

The cos.TET expression vector: This vector was derived from a cosmid clone containing the Syrian hamster PrP gene. It has been described in detail by Scott et al. (1992) and Hsiao et al. (1995). PS1 and PS2 cDNAs (full length or truncated) were inserted into this vector at its Sall site. The final constructs contain 20 kb of 5' sequence flanking the inserted cDNA. This 5' flanking sequence includes the PrP gene promoter, 50 bp of a PrP gene 5' untranslated region exon, a splice donor site, a 1 kb intron, and a splice acceptor site located immediately adjacent to the Sall site into which the PS1 or PS2 cDNA was inserted. The 3' sequence flanking the inserted cDNA includes an approximately 8 kb segment of PrP 3' untranslated region including a polyadenylation signal. Digestion of this construct with NotI (PS1) or FseI (PS2) released a fragment containing a mutant or wild type PS gene under the

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control of the PrP promoter. The released fragment was gel purified and injected into the pronuclei of fertilized mouse eggs using the method of Hsiao et al. (1995).

Platelet-derived growth factor receptor β-subunit constructs: PS cDNAs were also introduced between the SalI (full length PS1 cDNAs) or HindIII (truncated PS1 cDNAs, full length PS2 cDNAs, and truncated PS2 cDNAs) at the 3' end of the human platelet derived growth factor receptor β-subunit promoter and the EcoRI site at the 5' end of the SV40 polyA sequence and the entire cassette was cloned into the pZeoSV vector (Invitrogen, San Diego, CA.). Fragments released by ScaI/BamHI digestion were gel purified and injected into the pronuclei of fertilized mouse eggs using the method of Hsiao et al. (1995).

Human β -actin constructs: PS1 and PS2 cDNAs were inserted into the Sall site of pBAcGH. The construct produced by this insertion includes 3.4 kb of the human β actin 5' flanking sequence (the human β actin promoter, a spliced 78 bp human β actin 5' untranslated exon and intron) and the PS1 or PS2 insert followed by 2.2 kb of human growth hormone genomic sequence containing several introns and exons as well as a polyadenylation signal. Sfil was used to release a PS-containing fragment which was gel purified and injected into the pronuclei of fertilized mouse eggs using the method of Hsiao et al. (1995).

Phosphoglycerate kinase constructs: PS1 and PS2 cDNAs were introduced into the pkJ90 vector. The cDNAs were inserted between the KpnI site downstream of the human phosphoglycerate kinase promoter and the XbaI site upstream of the 3' untranslated region of the human phosphoglycerate kinase gene. PvuII/HindIII (PS1 cDNAs) or PvuII (PS2 cDNAs) digestion was used to release a PS-containing fragment which was then gel purified and injected into the pronuclei of fertilized mouse eggs as described above.

Analysis of Aβ in transgenic murine hippocampus: To analyze the effect of a mutant human PS1 transgene in mice, a PS1 mutation observed in conjunction with a particularly severe form of early-onset PS1-linked Alzheimer's disease was used, namely the M146L missense mutation (Sherrington et al., 1995). The animals, which were heterozygous for the PS1 mutant transgene on a mixed FVB-C57BL/6

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strain background, were cross-bred with similar mice bearing the human wild-type β APP₆₉₅ cDNA under the same Syrian hamster PrP promoter similar to those animals recently described by Hsiao et al., 1995. These cross breedings were done because it is thought that human A β is more susceptible to the formation of aggregates than are murine A β peptides.

The progeny of these $PS1_{M146L}$ x βAPP_{wT} cross-breedings were then genotyped to identify animals that contained both the human wild-type βAPP_{695} transgene and also the mutant human $PS1_{M146L}$ transgene. These mice were aged until two to three months of age and then sacrificed, with the hippocampus and neocortex being dissected rapidly from the brain and frozen. Litter mates of these mice, which contained only the wild-type human βAPP_{695} transgene were also sacrificed, and their hippocampi and neocortices were dissected and rapidly frozen as well.

The concentration of both total A β peptides (A $\beta_{x\to 0}$ and A $\beta_{x\to 2(43)}$) as well as the subset of A β peptides ending on residues 42 or 43 (long-tailed A β_{42} peptides) were then measured using a two-sandwich ELISA as described previously (Tamaoka et al., 1994; Suzuki et al., 1994). These results convincingly showed a small increase in total A β peptides in the double transgenic animals bearing wild-type human β APP₆₉₅ and mutant human PS1_{M146L} transgenes compared to the wild-type human β APP₆₉₅ controls. More impressively, these measurements also showed that there was an increase in the amount of long-tailed A β peptides ending on residues 42 or 43 (A β_{42}). In contrast, litter mates bearing only the wild-type human β APP₆₉₅ transgene had A β_{42} long-tailed peptide values which were below the limit of quantitation ("BLQ").

These observations therefore confirm that the construction of transgenic animals can recapitulate some of the biochemical features of human Alzheimer's disease (namely the overproduction of A β peptide and, in particular, overproduction of long-tailed isoforms of A β peptide). These observations thus prove that the transgenic models are in fact useful in exploring therapeutic targets relevant to the treatment and prevention of Alzheimer's disease.

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Analysis of hippocampus dependent memory functions in PS1 transgenic mice: Fourteen transgenic C57BL/6 x FVB mice bearing the human PS1_{MM6V} mutant transgene under the PrP promoter (as described) above and 12 wild type litter mates aged 2.5-3 months of age (both groups were balanced for age, weight, and sex) were investigated for behavioral differences attributable to the mutant transgene. Also the qualitative observation of murine behavior in their home cages did not indicate bimodal distribution of behaviors in the sample of animals.

Experiment 1. To test for subtle differences in exploratory behavior (e.g. locomotion, scanning of the environment through rearing, and patterns of investigation of unfamiliar environment), both $PS1_{MM6V}$ and wild type litter mates were tested in the open-field (Janus, et al. 1995). The results of the test revealed no significant differences between transgenics and controls in exploration of a new environment measured by mice locomotor behaviors (walking, pausing, wall leaning, rearing, grooming), (F(1,24) = .98, NS). Thus, differences any in behavior on the Morris water maze test (see below) cannot be attributed to differences in locomotor abilities, etc.

Experiment 2. One week after the open-field test, the PS1_{MM6V} mutant transgenic mice and their litter mates were trained in the Morris water maze. In this test, a mouse has to swim in a pool in order to find a submerged escape platform. The animal solves that test through learning the location of the platform using the available extra-maze spatial cues (Morris, 1990). This test was chosen because there is strong evidence that the hippocampal formation is involved in this form of learning. The hippocampus is also a major site of AD neuropathology in humans and defects in spatial learning (geographic disorientation, losing objects, wandering, etc.) are prominent early features of human AD. As a result the test is likely to detect early changes equivalent to those seen in human AD. The Morris test is conducted in three phases. In the first phase (the learning acquisition phase), the mouse has to learn the spatial position of the platform. In the second phase (the probe trial), the platform is removed from the pool and the mouse's search for the platform is recorded. In the final phase (the learning transfer phase), the platform is replaced in

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a new position in the pool, and the mouse has to learn that new spatial position of the platform.

Transgenic and wild type mice did not differ in their latencies to find the platform during learning acquisition (F(1,24) = 0.81, NS), and both groups showed rapid learning across trials (F(10,15) = 11.57, p < 0.001). During the probe trial phase, mice from both groups searched the quadrant of the pool which originally contained the platform significantly longer than other areas of the pool which had not contained the platform (F(3,22) = 28.9, p < 0.001). However, the wild type controls showed a trend which was not quite statistically significant (t(24) = 1.21, p = 0.24) for an increased number of crossings of the exact previous position of the platform. In the learning transfer test, both groups showed the same latency of finding the new position of the platform in the initial block of trials (t(24) = 1.11, NS). Such long latency to find the new spatial position is expected because the mice spent most of their time searching for the platform in the old spatial position. However, in later trials in the learning transfer phase, the wild type mice showed shorter swim latencies to the new position of the platform compared to the $PS1_{M146V}$ mutant transgenics (F(1,24) 2.36, p = 0.14). The results indicate that $PS1_{MH6V}$ mutant transgenic mice were less flexible in transferring learned information to a new situation and tended to persevere in their search for the platform in the old location.

Thus, although no differences were found in the spontaneous exploration of a new environment and in the acquisition of new spatial information between the wild type and the PSI_{M146V} mutant transgenic mice, the PSI_{M146V} mutant transgenic mice were impaired in switching and/or adapting this knowledge in later situations.

Electrophysiological Recordings in the hippocampus of mutant transgenic mice: Five to six months old litter mate control and human PS1_{M146V} mutant transgenic mice on the same C57BL/6 x FVB strain backgrounds as above were used to study long term potentiation (LTP) as an electrophysiologic correlate of learning and memory in the hippocampus. Recordings were carried out on 400 μm thick hippocampal slices according to conventional techniques. Briefly, brains were removed and transverse sections containing hippocampi were obtained within 1 min.

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after mice were decapitated under halothane anesthesia. Slices were kept at room temperature in oxygenated artificial cerebrospinal fluid for one hour prior to recording. One slice at a time was transferred to the recording chamber, where they were maintained at 32 °C in an interface between oxygenated artificial cerebrospinal fluid and humidified air. Slices were then allowed to equilibrate in the recording chamber for another hour.

Extracellular field recordings were carried out in the CA1 subfield of the hippocampus at the Schaeffer collateral-pyramidal cell synapse. Synaptic responses were induced by the stimulation of Schaeffer collaterals at a frequency of 0.03 Hz and an intensity of 30-50 % of maximal response. Tetani to evoke long-term potentiation consisted of 5 trains of 100 Hz stimulation lasting for 200 ms at an intertrain interval of 10 seconds. Field potentials were recorded using an Axopatch 200B amplifier (Axon Instrument). Glass pipettes were fabricated from borosilicate glass with an outer diameter of 1.5 mm, and pulled with a two step Narishige puller. Data were acquired on a 486-IBM compatible computer using PCLAMP6 software (Axon Instrument).

To test for any abnormality in presynaptic function, we investigated the differences in paired-pulse facilitation, which is an example of use-dependent increase in synaptic efficacy and is considered to be presynaptic in origin. In hippocampus, when two stimuli are delivered to the Schaeffer collaterals in rapid succession, paired-pulse facilitation manifests itself as an enhanced dendritic response to the second stimulus as the interstimulus interval gets shorter. In three pairs of wild-type/transgenic mice, we did not observe any difference in the paired-pulse facilitation over an interstimulus interval range of 20 ms to 1 sec. These data suggest that in PS1_{M146V} mutant transgenic mice, the excitability of Schaeffer collateral fibers and neurotransmitter release are likely to be normal.

Tetanic stimulation induced a long-lasting increase in the synaptic strength in both control (n = 3) and $PS1_{M146V}$ mutant transgenic mice (n = 2). In slices obtained from the $PS1_{M146V}$ mutant transgenic mice, long-lasting increase in the synaptic strength was 30.% more than that obtained from control mice.

Although preferred embodiments of the invention have been described herein in detail, it will be understood by those skilled in the art that variations may be made thereto without departing from the spirit of the invention or the scope of the appended claims.

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SEQUENCE LISTING
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(1) GENERAL INFORMATION:
           (i) APPLICANT: ST. GEORGE-HYSLOP, PETER H. ROMMENS, JOHANNA M. FRASER, PAUL E.
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       (iii) NUMBER OF SEQUENCES: 12
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    (viii) ATTORNEY/AGENT INFORMATION: (A) NAME: RAE, Patricia A.
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(B) TELEFAX: (416) 595-1163
(2) INFORMATION FOR SEQ ID NO:1:
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    (C) STRANDEDNESS: single
    (D) TOPOLOGY: linear
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(D) OTHER INFORMATION: /product= "S5a"
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AA Ly	G CT s Le 7	A CA u Hi 5	T AC	T GT	C CA	A CCC n Pro 80	· Ly:	G GG	C AAC y Lys	G ATC	ACC Thr 85	Pne	TGC Cy:	C AC s Th	G GC	SC Ly	411
AT 11 9	C CG e Ar 0	C G1 g Va	G GC	C CA a Hi	T CTC s Let 9!	G GCT 1 Ala 5	Let	AAC Ly:	G CAC	CGA Arg 100	CAA Gln	GGG	AAC Lys	G AA S As	T CA n Hi 10	. S	459
A A Ly	G AT	G CG	G AT	C AT e Il 11		TTT Phe	GTC Val	GG/ Gl/	A AGO Ser 115	Pro	GTG Val	GA G	GAC Asp	AA As	n GT	G u	507
AA Ly	G GA' S As	T CT P Le	G GT u Va 12	~ ~ 7	A CTO s Lev	GCT Ala	AAA Lys	CGC Arc	, wen	AAG Lys	AAG Lys	GAG Glu	AAA Lys 135	: Va.	A AA L As	T n	555
GT: Va:	GAG L Asj	2 AT 2 Il 14		C AA e As	T TTI n Phe	GGG	GAA Glu 145	GIL	GAG Glu	GTG Val	AAC Asn	ACA Thr 150	Glu	AAC Lys	G CT	G u	603
	15	5				TTG Leu 160	No II	GIY	Lys	Asp	165	Thr	Gly	Ser	H i	5	651
170)				175	GGG Gly		561	beu	180	wsb	Ala	Leu	Ile	Se: 18:	r 5	699
				190	5	GAA Glu	GLY	GIY	195	Met	Leu	Gly	Leu	G1 y 200	Ala	a	747
AGT Ser	GAC Asp	Ph	F GAZ e Glt 201		GGA Gly	GTA Val	GAT Asp	CCC Pro 210	AGT Ser	GCT Ala	GAT Asp	CCT Pro	GAG Glu 215	CTG Leu	GC(795
		220	5,	, , ,	Jer	ATG Met	225	GIU	GIN	Arg	GIn	230	Gln	Glu	Glu		843
	235		,,			GCA Ala 240	A.L.a	361	MIA	WIG	245	Ala	Gly	Ile	Ala	L	891
250		,			255	TCA Ser	чэр	wab	ита	260	reu	Lys	Met	Thr	11e 265		939
		U		270	J.,	CGC Arg	1111	GIY	275	PIO.	Asp	Leu	Ser	Ser 280	Met		987
			285			GCT Ala	LYL	290	мес	GIR	Met	Ser	Leu 295	Gln	Gly		1035
							303					310					1083
	315					GCC Ala 320	uys	GLU	GIU	Asp A	125	гуг	Asp	Val	Met		1131
330				• •• •	335	CAG :	361	Val	rea	340	isn 1	Leu	Pro	Gly	Val 345		1179
•				350		ATT (119	ASII	355	met G	T y	ser .	Leu	Ala 360	Ser		1227
			365		,	AAG / Lys 1	Jys .	376	Lys .	газ с	10 6	Ju ,	ASD 375	Lys	Lys		1275
TGAG	ACTG	GA (GGGA	AGG	T AG	CTGA	STCT	GCT	TAGG	GAC T	GCA1	GGG	GG A	ATTC			1330

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 377 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: Met Val Leu Glu Ser Thr Met Val Cys Val Asp Asn Ser Glu Tyr Met Arg Asn Gly Asp Phe Leu Pro Thr Arg Leu Gin Ala Gin Asp Ala Val Asn The Val Cys His Ser Lys Thr Arg Ser Asn Pro Glu Asn Asn 35 Val Gly Leu Ile Thr Leu Ala Asn Asp Cys Glu Val Leu Thr Thr Leu Thr Pro Asp Thr Gly Arg Ile Leu Ser Lys Leu His Thr Val Gln Pro Lys Gly Lys Ile Thr Phe Cys Thr Gly Ile Arg Val Ala His Leu Ala Leu Lys His Arg Gln Gly Lys Asn His Lys Met Arg Ile Ile Ala Phe Val Gly Ser Pro Val Glu Asp Asp Glu Lys Asp Leu Val Lys Leu Ala Lys Arg Leu Lys Lys Glu Lys Val Asn Val Asp Ile Ile Asn Phe Gly Glu Glu Glu Val Asn Thr Glu Lys. Leu Thr Ala Phe Val Asn Thr Leu Asn Gly Lys Asp Gly Thr Gly Ser His Leu Val Thr Val Pro Pro Gly 175 Pro Ser Leu Ala Asp Ala Leu Ile Ser Ser Pro Ile Leu Ala Gly Glu 180 190 Gly Gly Ala Met Leu Gly Leu Gly Ala Ser Asp Phe Glu Phe Gly Val Asp Pro Ser Ala Asp Pro Glu Leu Ala Leu Ala Leu Arg Val Ser Met Glu Glu Gln Arg Gln Arg Gln Glu Glu Ala Arg Arg Ala Ala Ala 225 Ala Ser Ala Ala Glu Ala Gly Ile Ala Thr Thr Gly Thr Glu Asp Ser Asp Asp Ala Leu Leu Lys Met Thr Ile Ser Gln Glu Phe Gly Arg Thr Gly Leu Pro Asp Leu Ser Ser Met Thr Glu Glu Glu Gln Ile Ala Tyr Ala Met Gln Met Ser Leu Gln Gly Ala Glu Phe Gly Gln Ala Glu Ser Ala Asp Ile Asp Ala Ser Ser Ala Met Asp Thr Ser Glu Pro Ala Lys Glu Glu Asp Asp Tyr Asp Val Met Gln Asp Pro Glu Phe Leu Gln 335 Ser Val Leu Glu Asn Leu Pro Gly Val Asp Pro Asn Asn Glu Ala Ile-340 350 Arg Asn Ala Met Gly Ser Leu Ala Ser Gln Ala Thr Lys Asp Gly Lys Lys Asp Lys Lys Glu Glu Asp Lys Lys

- (2) INFORMATION FOR SEQ ID NO: 3:
 - (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3841 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 - (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 2..3121

 - (ix) FEATURE:
 (A) NAME/KEY: misc feature
 (B) LOCATION: 1..3841
 (D) OTHER INFORMATION: /note= "GT24"

	t	xi)	SE	QUE	NCE	DESC	RIPT	TION:	: SE	_ç Q ID	9- No:	3:					
	1					5	9	GI y	1111	GIN	10	Arg	хаа	Thr	GIy	15	4 6
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			50					5	5		b AI	a Pi	O P	ro A. 60	la A	CC GC La Al	a
	•	5			•	•	7	ō			i Luc	u P 1	5 A	La PI	O 51	G CG	9
8	0	-				8	5				, 61	ð	L PI	O Th	ır Ly	G CTC	1 5
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160)					165	, ,,,			e GI	170	l ly:	E AI	a Th	r Le	G TCG u Ser 175	
		•			180)			. 561	185	GIN	Ty	r Se	r Ly:	3 Hi:		574
			1	95					200)	. Aly	PIC	, 61	205	Let	G GCA Ala	622
	-	21	0	-,			.,.	215	361	. 611	nıs	GIY	220	; Leu)	Gly	CCA Pro	670
	225		-		_		230		010		nis	235	wst	PIC	11e	TAT	718
											230					AGC Ser 255	766
	•				260					265	GIY	1 n·r	ryr	Arg	270		814
			2	75		_			280	561	GTC Val	FLO	nen	285	Arg	Thr	862
		290)		•			295		~14	GCG Ala	ита	300	Pne	GIn	Arg	910
	305	_				,	310		361	N 3 II	TAC Tyr	315	Asp	Pro	Tyr	Arg	958
320			•		- , -	325			GIU	ser	CCA Pro 330	ryr	Ser	Lys	Ser	Gly 335	1006
					3 4 0		,		260	345	AGG Arg	261	PIO	Ser	11e 350	Asp	1054
			3 5	5	-		-		360	,	TGG Trp	9	лэр	365	GIU	Leu	1102
	JAA	G 1 G	Αſ	. C	AG	ATG	rrg	CAG	CAC	CAG	TTT	ccc	TCG	GTC	CAG	TCT	1150

_									-100)-							
Pro	Glu	Val 370	Ile	Gln	Met	Leu	G1n 375	His	Gln	Phe	Pro	Ser 380	Val	. Glr	Ser		
AAC Asn	GCG Ala 385	GCA Ala	GCC Ala	TAC Tyr	TTG Leu	CAA G1n 390	CAC His	CTC Leu	TGT Cys	TTT Phe	GGA Gly 395	Asp	AAC Asn	Lys	ATT		1198
AAA Lys 400	GCC Ala	GAG Glu	ATA Ile	AGG Arg	AGA Arg 405	CAA Gln	GGA Gly	GGC Gly	ATC Ile	CAG Gln 410	Leu	CTG Leu	GTG Val	GAC Asp	CTG Leu 415		1246
Leu	Asp	HIS	Arg	Met 420	Thr	GAA Glu	Val	His	Arg 425	Ser	Ala	Суs	Gly	Ala 430	Leu		1294
AGA Arg	AAC Asn	CTG Leu	GTG Val 435	TAT Tyr	GGG Gly	AAG Lys	GCC Ala	AAC Asn 440	GAT Asp	GAT Asp	AAC Asn	AAA Lys	ATT Ile 445	GCC Ala	CTG Leu		1342
AAA Lys	AAC Asn	TGT Cys 450	GGT Gly	GGC GIy	ATC Ile	CCA Pro	GCA Ala 455	CTG Leu	GTG Val	AGG Arg	TTA Leu	CTC Leu 460	CGC Arg	AAG Lys	ACG Thr		1390
ACT	GAC Asp 465	CTG Leu	GAG Glu	ATC Ile	CGG Arg	GAG Glu 470	CTG Leu	GTC Val	ACA Thr	GGA Gly	GTC Val 475	CTT Leu	TGG Trp	AAC Asn	CTC Leu		1438
480	Ser	cys	Asp	Ala	485	AAA Lys	Met	Pro	Ile	11e 490	Gln	Asp	Ala	Leu	Ala 495		1486
GTA Val	CTG Leu	ACC	AAC Asn	GCG Ala 500	GTG Val	ATT Ile	ATC Ile	Pro	CAC His 505	TCA Ser	GGC Gly	TGG Trp	GAA Glu	AAT Asn 510	TCG Ser		1534
CCT Pro	CTT Leu	CAG Gln	GAT Asp 515	GAT Asp	CGG Arg	AAA Lys	ATA Ile	CAG Gln 520	CTG Leu	CAT	TCA Ser	TCA Ser	CAG Gln 525	GTG Val	CTG Leu		1582
CGT Arg	AAC Asn	GCC Ala 530	ACC Thr	GGG Gly	TGC Cys	CTA Leu	AGG Arg 535	AAT Asn	GTT Val	AGT Ser	TCG Ser	GCC Ala 540	GGA Gly	GAG Glu	GAG Glu		1630
GCC Ala	CGC Arg 545	AGA Arg	AGG Arg	ATG Met	AGA Arg	GAG Glu 550	TGT Cys	GAT Asp	GGG Gly	CTT Leu	ACG Thr 555	GAT Asp	GCC Ala	TTG Leu	CTG Leu		1678
TAC Tyr 560	GTG Val	ATC Ile	CAG Gln	TCT Ser	GCG Ala 565	CTG Leu	GGG Gly	AGC Ser	AGT Ser	GAG Glu 570	ATC Ile	GAT Asp	AGC Ser	AAG Lys	ACC Thr 575	:	1726
GTT Val	GAA Glu	AAC Asn	TGT Cys	GTG Val 580	TGC Cys	ATT Ile	TTA Leu	AGG Arg	AAC Asn 585	CTC Leu	TCG Ser	TAC Tyr	CGG Arg	CTG Leu 590	GCG Ala	:	1774
GCA Ala	GAA Glu	ACG Thr	TCT Ser 595	CAG Gln	GGA Gly	CAG Gln	CAC His	ATG Met 600	GGC Gly	ACG Thr	GAC Asp	GAG Glu	CTG Leu 605	GAC Asp	GGG G1 y	:	1822
CTA Leu	CTC Leu	TGT Cys 610	GGC Gly	GAG Glu	GCC Ala	AAT Asn	GGC Gly 615	AAG Lys	GAT Asp	GCT Ala	GAG Glu	AGC Ser 620	TCT Ser	GGG	TGC Cys	:	1870
TGG Trp	GGC Gly 625	AAG Lys	AAG Lys	AAG Lys	AAG Lys	AAA Lys 630	AAG Lys	AAA Lys	TCC Ser	CAA Gln	GAT Asp 635	CAG Gln	TGG Trp	GAT Asp	GGA Gly	:	1918
GTA Val 640						TGT Cys						GGG Gly			ATG Met 655	1	1966
CTG Leu	TGG Trp	CAC	CCA Pro	TCA Ser 660	ATA Ile	GTC Val	AAA Lys	CCC Pro	TAC Tyr 665	CTC Leu	ACA Thr	CTG Leu	CTC Leu	TCT Ser 670	GAG Glu	2	2014
TGC Cys	TCA Ser	AAT Asn	CCA Pro 675	GAC Asp	ACG Thr	CTG Leu	GAA Glu	GGG Gly 680	GCG Ala	GCA Ala	GGC Gly	GCC Ala	CTG Leu 685	CAG Gln	AAC Asn	2	2062
TTG Leu	GCT Ala	GCA Ala 690	GGG Gly	AGC Ser	TGG Trp	AAG Lys	TGG Trp 695	TCA Ser	GTA Val	TAT Tyr	ATC Ile	CGA Arg 700	GCC Ala	GCT Ala	GTC Val	2	2110
CGA Arg	AAA Lys 705	GAG Glu	AAA Lys	GGC Gly	CTG Leu	CCC Pro 710	ATC Ile	CTC Leu	GTG Val	GAG Glu	CTG Leu 715	CTC Leu	CGA Arg	ATA Ile	GAC Asp	2	2158
AAT Asn 720	GAC Asp	CGT Arg	GTG Val	GTG Val	TGC Cys 725	GCG Ala	GTG Val	GCC Ala	ACT Thr	GCG Ala 730	CTG Leu	CGG Arg	AAC Asn	ATG Met	GCC Ala 735	2	2206
TTG Leu	GAC Asp	GTC Val	AGA Arg	AAT Asn 740	AAG Lys	GAG Glu	CTC Leu	ATC Ile	GGC Gly 745	AAA Lys	TAC Tyr	GCC Ala	ATG Met	CGA Arg 750	GAC Asp	7	2254

CT; Let	A GTO	C CA l Hi	C AG S Ar 75	G CT g Le	T CC.	A GG	A GG y Gl	G AAG y Asi 766	-10 C AAC n Asi	AGC Ser	AAC Asn	AAC Asn	ACT Thr	GC;	AGC Ser	2302
AAC Lys	G GCG S Ala	AT A Me 77	G TC t Se 0	G GA r As	T GAO P Asi	AC.	A GT F Va. 77	G ACI	A GC1	GTC Val	TGC Cys	TGC Cys 780	ACA	CT C	CAC	2350
Glu	785	l II	e Th	C AA	G AAC S Asi	790	G GAG	S AAC J Asr	GCC	Lys	GCC Ala 795	TTA Leu	CGG Arg	GAT Asp	GCC Ala	2398
300					805	1	•	_		AAA Lys 810	261	Lys	GIY	Asp	Lys	2446
				820)				825	CAG Gln	* 4 1	reu	Asn	Ser	ATG Met	2494
			833	,				840	- , -	AAA Lys	4,3	ush	GIA	Trp	Ser	2542
		000					855			ATC Ile	014	ลิลัก	Asp	Arg	Gln	2590
	003					870				ATC Ile	875		Val	Arg	Val	2638
Ser 880	Pro	ASD	AAC	Arg	TCA Ser 885	GCA Ala	AGT Ser	GCC Ala	CCA Pro	GCT Ala 890	TCA (CCT (CGG Arg	GAA Glu	ATG Met 895	2686
				900				•	905	GAG Glu	-,3		TA	Ser	Asn	2734
			915				-	920		ACT T	, u	.ry 1	25	Asp /	Ala	2782
		930				•	935			TTG 1 Leu 1	yr A	40 A	sn s	er ?	yr	2830
	943					950	-				55	er A	ra G	TN E	, LO	2878
GTC (Val 1960	י ידים	T			965		•	•	, ,	70		yr G	ın P	ro P	'he '75	2926
CAG A	י אמי	~cc /		980				9	85			LU A	9. 9.	90 V	al	2974
CAC C His H			773				1	LÕOO	•••		-3 D	- u G	X - 4-	eu L	уs	3022
TCC A Ser T	TG A	AC 1	7 A T 4													3070
GAA C Glu L 1	023				1	030		•		10	֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓֓	OAS	p Se	er T	G Cp	3118
GTG T Val 1040																.3.171
CACAA	ATAC	· · ·			ITT	GGTT	TTT	TTCT	CTG	CA AA	TTTA	GTTT	GTT	AAAG	CCT	3231
91100	MI MG	G AA	GGCT	GTGA	TAA	CCAG	TAA	GGGA	ATA	44 T	GACC	ጥ አ ጥ ጥ	~			3291
	MAIC	G CA	AGTT	AACT	TGG.	AAAT	CAG	TAGA	AGC1	4 A A A	GTCN	Tr c c m				3351
010000	MUC	A CC	TTTC	CTAG	CGT	STTN	TGT '	TAGG	GTA	C CA	GAAC	***				3411
001000	, , I G	M TT	GGTA	GGGT	GGA	SNCG	AGG (CATTO	GGGC	c cc	TCCC					3471
	CAC	. 40	MCAC	AGAA	TAG	CTTA	CAC	ACTGT	GTGG	e ce	3000	~ = = ~				3531
TTTACT GCAAGA	TGT	יייי דכי	TEC	CTTG	TORC	TCTA	IGG (ETTCA	GGTT	G CA	TTGG	GTT	CCT	CTGT.	ACA	3591
GCAAGA ATTAAA	GAAG	NA!	AAGC	GCGT	TGTG	TART:	TTT -	TGTT	GTAA	A GT	ATTTO	SATG	TAC	ATTA	CAG	3651
ATTAAA GGTTCT	AAAT	ATI	GCT	CAA	TTTC	NAAC	· · · · A	TCS	AATN	C CGC	CGTG	TTT	CCT	ATC	FAT	3711
					•			IGAA	A G A T	G TAT	rGGAT	TTC	CAGI	TTTT	CT	3771

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TTACTTTCTC CCAGTATGTT TTAACCNMMN AAAAAAAAAA AAAAAAAAA AAAAAAAAA 3831 AAAACTCGAG 3841

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1040 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

- (ii) MOLECULE TYPE: protein
- Ser Gln Leu Pro Ala Arg Gly Thr Gln Ala Arg Xaa Thr Gly Gln Ser Phe Ser Gln Gly Thr Thr Ser Arg Ala Gly His Leu Ala Gly Pro Glu Pro Ala Pro Pro Pro Pro Pro Xaa Pro Arg Glu Pro Phe Ala Pro Ser Leu Gly Ser Ala Phe His Leu Pro Asp Ala Pro Pro Ala Ala Ala Ala Ala Ala Leu Tyr Tyr Ser Xaa Ser Thr Leu Pro Ala Pro Pro Arg Gly Gly Ser Pro Leu Ala Ala Pro Gln Gly Gly Ser Pro Thr Lys Leu Gln Arg Gly Gly Ser Ala Pro Glu Gly Ala Thr Tyr Ala Ala Pro Arg Gly Ser Ser Pro Lys Gln Ser Pro Ser Arg Leu Ala Lys Ser Tyr Ser Thr Ser Ser Pro Ile Asn Ile Val Val Ser Ser Ala Gly Leu Ser Pro Ile 130 $$140\$ Arg Val Thr Ser Pro Pro Thr Val Gln Ser Thr Ile Ser Ser Pro Ile His Gln Leu Ser Ser Thr Ile Gly Thr Tyr Ala Thr Leu Ser Pro Thr Lys Arg Leu Val His Ala Ser Glu Gln Tyr Ser Lys His Ser Gln Glu Leu Tyr Ala Thr Ala Thr Leu Gln Arg Pro Gly Ser Leu Ala Ala Gly Ser Arg Ala Ser Tyr Ser Ser Gln His Gly His Leu Gly Pro Glu 210 220 Leu Arg Ala Leu Gln Ser Pro Glu His His Ile Asp Pro Ile Tyr Glu 225 230 230 Asp Arg Val Tyr Gln Lys Pro Pro Met Arg Ser Leu Ser Gln Ser Gln 255 Gly Asp Pro Leu Pro Pro Ala His Thr Gly Thr Tyr Arg Thr Ser Thr Ala Pro Ser Ser Pro Gly Val Asp Ser Val Pro Leu Gln Arg Thr Gly 275 Ser Gln His Gly Pro Gln Asn Ala Ala Ala Ala Thr Phe Gln Arg Ala 290 Ser Tyr Ala Ala Gly Pro Ala Ser Asn Tyr Ala Asp Pro Tyr Arg Gln 305 315 320 Leu Gln Tyr Cys Pro Ser Val Glu Ser Pro Tyr Ser Lys Ser Gly Pro Ala Leu Pro Pro Glu Gly Thr Leu Ala Arg Ser Pro Ser Ile Asp Ser Ile Gin Lys Asp Pro Arg Glu Phe Gly Trp Arg Asp Pro Glu Leu Pro

Glu Val Ile Gln Met Leu Gln His Gln Phe Pro Ser Val Gln Ser Asn 370

Ala Ala Ala Tyr Leu Gln His Leu Cys Phe Gly Asp Asn Lys Ile Lys 385

Ala Glu Ile Arg Arg Gln Gly Gly Ile Gln Leu Leu Val Asp Leu Leu 415

Asp His Arg Met Thr Glu Val His Arg Ser Ala Cys Gly Ala Leu Arg $\frac{420}{420}$ Asn Leu Val Tyr Gly Lys Ala Asn Asp Asp Asn Lys Ile Ala Leu Lys As Cys Gly Gly Ile Pro Ala Leu Val Arg Leu Arg Lys Thr Thr 450 460 Asp Leu Glu Ile Arg Glu Leu Val Thr Gly Val Leu Trp Asn Leu Ser 465 475 480 Ser Cys. Asp Ala Leu Lys Met Pro Ile Ile Gln Asp Ala Leu Ala Val 495 Leu Thr Asn Ala Val Ile Ile Pro His Ser Gly Trp Glu Asn Ser Pro 500 510Leu Gln Asp Asp Arg Lys Ile Gln Leu His Ser Ser Gln Val Leu Arg Asn Ala Thr Gly Cys Leu Arg Asn Val Ser Ser Ala Gly Glu Glu Ala 530 Arg Arg Arg Met Arg Glu Cys Asp Gly Leu Thr Asp Ala Leu Leu Tyr 545 550 560 Val Ile Gln Ser Ala Leu Gly Ser Ser Glu Ile Asp Ser Lys Thr Val 565 575 Glu Asn Cys Val Cys Ile Leu Arg Asn Leu Ser Tyr Arg Leu Ala Ala 580 585 Glu Thr Ser Gln Gly Gln His Met Gly Thr Asp Glu Leu Asp Gly Leu 595 Leu Cys Gly Glu Ala Asn Gly Lys Asp Ala Glu Ser Ser Gly Cys Trp 610Gly Lys Lys Lys Lys Lys Ser Gln Asp Gln Trp Asp Gly Val 625 630 640 Gly Pro Leu Pro Asp Cys Ala Glu Pro Pro Lys Gly Ile Gln Met Leu 645 650 655 Trp His Pro Ser Ile Val Lys Pro Tyr Leu Thr Leu Leu Ser Glu Cys 660 670 Ser Asn Pro Asp Thr Leu Glu Gly Ala Ala Gly Ala Leu Gln Asn Leu 675 685 Ala Ala Gly Ser Trp Lys Trp Ser Val Tyr Ile Arg Ala Ala Val Arg Lys Glu Lys Gly Leu Pro Ile Leu Val Glu Leu Leu Arg Ile Asp Asn 705 Asp Arg Val Val Cys Ala Val Ala Thr Ala Leu Arg Asn Met Ala Leu 725 730 735 Asp Val Arg Asn Lys Glu Leu Ile Gly Lys Tyr Ala Met Arg Asp Leu 740 Val His Arg Leu Pro Gly Gly Asn Asn Ser Asn Asn Thr Ala Ser Lys 755 Ala Met Ser Asp Asp Thr Val Thr Ala Val Cys Cys Thr Leu His Glu 770 780 Val Ile Thr Lys Asn Met Glu Asn Ala Lys Ala Leu Arg Asp Ala Gly 785 795 800 Gly Ile Glu Lys Leu Val Gly Ile Ser Lys Ser Lys Gly Asp Lys His 805 810 815 Ser Pro Lys Val Val Lys Ala Ala Ser Gln Val Leu Asn Ser Met Trp 820 830 Gln Tyr Arg Asp Leu Arg Ser Leu Tyr Lys Lys Asp Gly Trp Ser Gln 835 Tyr His Phe Val Ala Ser Ser Ser Thr Ile Glu Arg Asp Arg Gln Arg Pro Tyr Ser Ser Ser Arg Thr Pro Ser Ile Ser Pro Val Arg Val Ser 865 870 880 Pro Asn Asn Arg Ser Ala Ser Ala Pro Ala Ser Pro Arg Glu Met Ile 885 890 895 Ser Leu Lys Glu Arg Lys Thr Asp Tyr Glu Cys Thr Gly Ser Asn Ala 900Thr Tyr His Gly Gly Lys Gly Glu His Thr Ser Arg Lys Asp Ala Met 915

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Thr Ala Gln Asn Thr Gly Ile Ser Thr Leu Tyr Arg Asn Ser Tyr Gly 930 940 Ala Pro Ala Glu Asp Ile Lys His Asn Gln Val Ser Ala Gln Pro Val 945 950 955 960 Pro Gln Glu Pro Ser Arg Lys Asp Tyr Glu Thr Tyr Gln Pro Phe Gln 975 Asn Ser Thr Arg Asn Tyr Asp Glu Ser Phe Phe Glu Asp Gln Val His 980His Arg Pro Pro Ala Ser Glu Tyr Thr Met His Leu Gly Leu Lys Ser 995 . 1000Thr Gly Asn Tyr Val Asp Phe Tyr Ser Ala Ala Arg Pro Tyr Ser Glu 1010 1020Leu Asn Tyr Glu Thr Ser His Tyr Pro Ala Ser Pro Asp Ser Trp Val 1025 1030 1035

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 3907 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 142..3777
 (D) OTHER INFORMATION: /note= "p0071"

(xi)	SEQUENCE	DESCRIPTION:	SEQ	ΙD	NO:5:	

CTACTGTTGT T	TTTGAGGGG C	GGCAGCCG CG	cceccece e	GCACTTTTT A	ATTTTTCG	60
GGTGCCGCAG C	AGCGACCCC TO	GGCGCCGA TG	CCCTGAT C	CCTGGAGCG A	CGACGGCCG	120
CTGCCTAAGC T	GGGAAGAGG A	ATG CCA GCT Met Pro Ala	CCT GAG C Pro Glu G	CAG GCC TCA Gln Ala Ser	TTG GTG Leu Val 10	171
GAG GAG GGG Glu Glu Gly	CAA CCA CAG Gln Pro Gln 15	ACC CGC CAG Thr Arg Gln	GAA GCT G Glu Ala A 20	GCC TCC ACT Ala Ser Thr	GGC CCA :	219
GGC ATG GAA Gly Met Glu	CCC GAG ACC Pro Glu Thr 30	ACA GCC ACC Thr Ala Thr 35	ACT ATT C	CTA GCA TCC Leu Ala Ser 40	GTG AAG Val Lys	267
GAG CAG GAG Glu Gln Glu 45	CTT CAG TTT Leu Gln Phe	CAG CGA CTC Gln Arg Leu 50	ACC CGA G Thr Arg G	GAA CTG GAA Glu Leu Glu 55	GTG GAA Val Glu	315
AGG CAG ATT Arg Gln Ile 60	GTT GCC AGT Val Ala Ser	CAG CTA GAA Gln Leu Glu 65	AGA TGT A Arg Cys A	AGG CTT GGA Arg Leu Gly 70	GCA GAA Ala Glu	363
TCA CCA AGC Ser Pro Ser 75	ATC GCC AGC Ile Ala Ser 80	ACC AGC TCA Thr Ser Ser	ACT GAG A Thr Glu L 85	AAG TCA TTT Lys Ser Phe	CCT TGG Pro Trp 90	411
AGA TCA ACA Arg Ser Thr	GAC GTG CCA Asp Val Pro 95	AAT ACT GGT Asn Thr Gly	GTA AGC A Val Ser L 100	AAA CCT AGA Lys Pro Arg	GTT TCT Val Ser 105	459
GAC GCT GTC Asp Ala Val	CAG CCC AAC Gln Pro Asn 110	AAC TAT CTC Asn Tyr Leu 115	ATC AGG A Ile Arg T	ACA GAG CCA Thr Glu Pro 120	GAA CAA Glu Gln	507
GGA ACC CTC Gly Thr Leu 125	TAT TCA CCA Tyr Ser Pro	GAA CAG ACA Glu Gln Thr 130	TCT CTC C Ser Leu H	CAT GAA AGT His Glu Ser 135	GAG GGA Glu Gly	555
TCA TTG GGT Ser Leu Gly 140	AAC TCA AGA Asn Ser Arg	AGT TCA ACA Ser Ser Thr 145	Gln Met A	AAT TCT TAT Asn Ser Tyr 150	TCC GAC Ser Asp	603
AGT GGA TAC Ser Gly Tyr 155	CAG GAA GCA Gln Glu Ala 160	GGG AGT TTC Gly Ser Phe	CAC AAC A His Asn S 165	AGC CAG AAC Ser Gln Asn	GTG AGC Val Ser 170	651
AAG GCA GAC Lys Ala Asp	AAC AGA CAG Asn Arg Gln 175	CAG CAT TCA Gln His Ser	TTC ATA G Phe Ile G 180	Gly Ser Thr	AAC AAC. Asn Asn 185	699
CAT GTG GTG His Val Val	AGG AAT TCA Arg Asn Ser	AGA GCT GAA Arg Ala Glu	GGA CAA A	ACA CTG GTT Thr Leu Val	CAG CCA Gln Pro	747

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			19					19					2 (
T C S e	A GT r Va	A GC 1 A1 20	a wa	T CG n Ar	G GC g Al	C AT(G AG	g ar	A GT g Va	T AG	T TC T Se	A G1	il Pi	CA TO	T AGA	795
GC Al	A CA a G1 22	** 26	T CC	T TC o Se	T TA	T GT: r Va. 22:		C AG e Se	C AC	A GG	C GT y Va 23	1 Se	T CC	T TO	A AGG	843
GG G1 23	yac	T CT r Le	G AG u Ar	A AC g Th	T TC r Se · 24	r rer	G GG	T AG y Se	T GG	A TT: y Pho 24!	e Gl	C TC y Se	T CC	G TO	A GTG r Val 250	891
AC Th	C GA r As	C CC p Pr	C CG o Ar	A CC g Pr 25	o ne	G AAC u Asr	Pre	AG Se	T GC: r Ala 26	а тур	T TC	C TC r Se	C AC	C AC	A TTA	939
CC Pr	T GC	T GC a Al	A CG a Ar 27	G GC g Al	A GC a Al	C TCI a Ser	CCC	5 TAC 5 Ty: 27:	C TCI	A CAC	AG Ar	A CC g Pr	C GC o A1 28	C TC	C CCA	987
AC. Th	A GC	T AT. a Il 28:		G CG g Ar	G AT	T GGG e Gly	TC; Se; 290	. va.	C ACC	TCC Ser	CG	G CA g G1 29	G AC		C AAT r Asn	1035
Pr	C AAG D Ass 30		A CC	A AC	C CC	T CAA O Gln 305	Tyt	CA; Glr	A ACC	ACC	GC A1	a Ar	A GT g Va	G GG	G TCC y Ser	1083
Pro	, ne	G AC	C CT	3 ACC	G GAT	n wra	CAG Gln	ACT	CGA Arg	GTA Val 325	Ala	T TC	C CC	A TC o Se	C CAA r Gln 330	1131
GG(C CAC	G GTC	G GGG	335	. 361	TCC Ser	Pro	Lys	CGC Arg 340	Ser	GG Gl	ATO	G AC	C GC(F A1. 34!	C GTA a Val	1179
Pro	CAC Glr	G CAT	CTC 5 Let 350	LGI	A CCT	TCA Ser	CTG Leu	CAA Gln 355	ı arg	ACT	GTT Val	CAT His	GA0 B Asi 360	P Me1	G GAG	1227
CAA Glr	TTC Phe	GG # G1 y 365		GAC Glr	CAG Gln	TAT	GAC Asp 370	ATT Ile	TAT Tyr	GAG Glu	AGG	ATO Met 375	: Val	CC/	CCC Pro	1275
,	380	, ,,,	, 261	260	1111	GGC Gly 385	rea	Arg	Ser	Ser	390	Ala	Ser	Glr	His	1323
395				GII	400		ALG	Sel	АТА	405	Ser	Pro	Asp	Let	410	1371
				415	GIU	GGG Gly	ALY	rnr	420	Tyr	ser	Pro	Val	425	Arg	1419
•••		A311	430	GIY	Int	GTG Val	GIU	435	GIN	Gly	Ser	Gln	Thr 440	Ala	Leu	1467
	y	4 4 5	Gly	Val	ser	GGT Gly	450	GIY	Asn	Leu	Gln	Arg 455	The	Ser	Ser	1515
	460			Deu		TAC Tyr 465	GIII	ALG	nea	Asn	470	Ala	Leu	Asn	Thr	1563
475			- 1 -		480	CCC	1 7 1	ALY	PIO	485	GIN	туг	Arg	Val	Gln 490	1611
	-,,		. , .	495	Ary	CTT Leu	GIN	HIS	500	Val	PIO	Ala	Asp	A 5 p 5 0 5	Gly	1659
		9	510		361	ATA Ile	жэр	515	116	GIN	Lys	Asp	Pro 520	Arg	Glu	1707
		525	nry	пор		GAG Glu	530	PIO	GIU	val	IIe	535	Met	Leu	Glu	1755
	540					CAG Gln 545			VIG	A14	550	ıyr	Leu	GIn	His	1803
555	-,-		,		560	AAA Lys	· 4 I	гуз	Met	565	vaı	Cys	Arg	Leu	G1 y 570	1851
GGA	ATC	AAG	CAT	CIG	GTT	GAC	CTT	CTG	GAC	CAC .	AGA	GTT	TTG	GAA	GTT	1899

									104							
Gly	Ile	Lys	His	Leu 575	Val	Asp	Leu	Leu	-106 Asp 580	His	Arg	, Val	Leu	Glu 585	Val	
CAG Gln	AAG Lys	AAT Asn	GCT Ala 590	C y s	GGT	GCC	CTT	CGA Arg 595	AAC Asn	CTC Leu	GTT Val	TTT Phe	GG 0	' Lys	TCT	1947
ACA Thr	GAT Asp	GAA Glu 605	w an	AAA Lys	ATA	GCA Ala	ATG Met 610	AAG Lys	AAT Asn	GTT Val	GG T Gly	GGG Gly 615	ATA Ile	CCT	GCC Ala	1995
TTG Leu	TTG Leu 620	CGA Arg	CTG Leu	TTG Leu	AGA Arg	AAA Lys 625	TCT Ser	ATT	GAT Asp	GCA Ala	GAA Glu 630	GTA Val	AGG Arg	GAG Glu	CTT Leu	2043
GTT Val 635	ACA Thr	GGA Gly	GTT Val	CTT Leu	TGG Trp 640	AAT Asn	TTA Leu	TCC Ser	TCA Ser	TGT Cys 645	GAT Asp	GCT Ala	GTA Val	AAA Lys	ATG Met 650	2091
ACA Thr	ATC Ile	ATT Ile	CGA Arg	GAT Asp 655	GCT Ala	CTC Leu	TCA Ser	ACC Thr	TTA Leu 660	ACA Thr	AAC Asn	ACT Thr	GTG Val	ATT Ile 665	GTT Val	2139
CCA Pro	CAT His	TCT Ser	GGA Gly 670	TGG Trp	AAT Asn	AAC Asn	TCT Ser	TCT Ser 675	TTT Phe	GAT Asp	GAT Asp	GAT Asp	CAT His 680	AAA Lys	ATT Ile	2187
AAA Lys	TTT Phe	CAG Gln 685	ACT Thr	TCA Ser	CTA Leu	GTT Val	CTG Leu 690	CGT Arg	AAC Asn	ACG Thr	ACA Thr	GGT Gly 695	TGC Cys	CTA Leu	AGG Arg	2235
AAC Asn	CTC Leu 700	ACG Thr	TCC Ser	GCG Ala	GGG Gly	GAA Glu 705	GAA Glu	GCT Ala	CGG Arg	AAG Lys	CAA Gln 710	ATG Met	CGG	TCC Ser	TGC Cys	2283
GAG Glu 715	GGG Gly	CTG Leu	GTA Val	GAC Asp	TCA Ser 720	CTG Leu	TTG Leu	TAT Tyr	GTG Val	ATC Ile 725	CAC	ACG Thr	TGT Cys	GTG Val	AAC Asn 730	2331
ACA Thr	TCC Ser	GAT	TAC Tyr	GAC Asp 735	AGC Ser	AAG Lys	ACG Thr	GTG Val	GAG Glu 740	AAC Asn	TGC Cys	GTG Val	TGC Cys	ACC Thr 745	CTG Leu	2379
AGG Arg	AAC Asn	CTG Leu	TCC Ser 750	TAT	CGG Arg	CTG Leu	GAG Glu	CTG Leu 755	GAG Glu	GTG Val	CCC	CAG Gln	GCC Ala 760	CGG Arg	TTA Leu	2427
CTG Leu	GGA Gly	CTG Leu 765	AAC Asn	GAA Glu	TTG Leu	GAT Asp	GAC Asp 770	TTA Leu	CTA Leu	GGA Gly	AAA Lys	GAG Glu 775	TCT Ser	CCC	AGC Ser	2,475
AAA Lys	GAC Asp 780	TCT Ser	GAG Glu	Pro	AGT	TGC Cys 785	TGG Trp	GGG Gly	AAG Lys	AAG Lys	AAG Lys 790	AAA Lys	AAG Lys	AAA Lys	AAG Lys	2523
AGG Arg 795	ACT Thr	CCG Pro	CAA Gln	GAA Glu	GAT Asp 800	CAA Gln	TGG Trp	GAT Asp	GGA Gly	GTT Val 805	GGT Gly	CCT Pro	ATC Ile	CCA Pro	GGA Gly 810	2571
CTG Leu	TCG Ser	AAG Lys	TCC Ser	CCC Pro 815	AAA Lys	GGG Gly	GTT Val	GAG Glu	ATG Met 820	CTG Leu	TGG Trp	CAC	CCA Pro	TCG Ser 825	GTG Val	2619
GTA Val	AAA Lys	Pro	TAT Tyr 830	CTG Leu	ACT Thr	CTT Leu	CTA Leu	GCA Ala 835	GAA Glu	AGT Ser	TCC	AAC Asn	CCA Pro 840	GCC Ala	ACC	2667
TTG Leu	GAA Glu	GGC Gly B45	TCT Ser	GCA Ala	GGG Gly	TCT Ser	CTC Leu 850	CAG Gln	AAC Asn	CTC Leu	TCT Ser	GCT Ala 855	AGC Ser	AAC Asn	TGG Trp	2715
гуз	860	Ala	Ala	туг	IIe	865	Gly	GGC	Arg	Pro	E 7 0	Arg	Lys	Gly	Leu	2763
Pro 875	ATC Ile	CTT Leu	GTG Val	GAG Glu	CTT Leu 880	CTG Leu	AGA Arg	ATG Met	GAT Asp	AAC Asn 885	GAT Asp	AGA Arg	GTT Val	GTT Val	TCT Ser 890	2811
TCC Ser	GGT Gly	GCA Ala	ACA Thr	GCC Ala 895	TTG Leu	AGG Arg	AAT Asn	ATG Met	GCA Ala 900	CTA Leu	GAT Asp	GTT Val	CGC	AAC Asn 905	AAG Lys	2859
GAG Glu	CTC Leu	ATA Ile	GGC Gly 910	AAA Lys	TAC Tyr	GCC Ala	ATG Met	CGA Arg 915	GAC Asp	CTG Leu	GTC Val	AAC Asn	CGG Arg 920	CTC Leu	CCC Pro	2907
GIY	GIY	925	стА	PIO	ser	vaı	930	TCT Ser	Asp	Glu	Thr	935	Ala	Ala	Ile	2955
TGC Cys	TGT Cys 940	GCT Ala	CTG Leu	CAC His	GAG Glu	GTC Val 945	ACC Thr	AGC Ser	AAA Lys	AAC Asn	ATG Met 950	GAG Glu	AAC Asn	GCA Ala	AAA Lys	3003

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900	,				960	, -			AAG Lys	Leu 965		T Wat	. 110	e In	C AAA r Lys 970	305
			•	975	i		,	. Dys	980	val	ьуз	G GCA S Ala	Ala	9 Al	C CAG a Gln 5	3099
GTC Val	Leu	AAT Asn	ACA Thr 990	Leu	TGG	CAA Gln	TAT	CGG Arg 995	GAC Asp	CTC Leu	CGC	AGC Ser	ATT Ile	TA Ty	T AAA I Lys	3147
								•				101	3		A TTG	3195
	102	0	_		•	1025	5		001	Den	103	0 111	ACC Thr	AAC Asr	CAA Gln	3243
103	5				1040	0	ser		Gly	Ser 104	Thr 5	Ser			CCA Pro 1050	3291
				105	5			•	1060)	.,.	пэр	Arg	106	CAG. Gln	3339
			1070	י כ	_			1075	, ,	nsp	VI.	ACA Thr	1080	Lys	Gly	3387
		1085	5			•	109	ס ֿ ַ			1 Y L	1095	ser	ser	Tyr	3435
	1100	,				1105		•			iiic	CAT His	GIN	GIN	reu	3483
TAT Tyr 1115	TAT	AGT Ser	CAA Gln	GAT Asp	GAC Asp 1120	TCC Ser	AAC Asn	AGA Arg	AAG Lys	AAC Asn 1125	TTT Phe	GAT Asp	GCA Ala	TAC Tyr	AGA Arg 1130	3531
												TAT Tyr				3579
			1150					1155	- ,	,		CAG Gln	1160	GIA	Leu	3627
AAA Lys		1165			•	1	150		· y	er.	HE	1175	arg	Pro	Ser	3675
TAT Tyr	AGA Arg 1180	GCA Ala	GAA (Glu (CAG '	TAC (Tyr I	CCA G Pro G 1185	GG 1 y	TCC (Ser i	CCA C	AC 1	CA Ser	TGG (Trp \	TG :	TAC Tyr	GAT Asp	3723
CAA (Gln 1 1195	GAT (GCC (CAA (Gln (CAG I	AGG A Arg A 1200	AAC T Asn S	CT	TTC 1 Phe F	TTT C	TA A eu 1 205	CC Thr	TTG T Leu F	TC I	AGA Arg	TTG Leu 1210	3771
AGG 1 Arg	rga /	AAAG:	TCCAT	rc Ti	GCTG	SATTT	CA	TGATT	GAA	ATGI	'GAA	AGT G	AAGI	rgga.	AG	3827
GAAT	SAAT	GA AC	STGTO	STTTI	TTT	TTCC	TTT	TTGA	GGAA	TT A	TCAC	iece »	A T7		TATCA	2005
AGCTI	TATC	SA TA	ACCGI	CGAC	:									CGA	ARCA	3887
																3907

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 1212 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

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Gln Leu Glu Arg Cys Arg Leu Gly Ala Glu Ser Pro Ser Ile Ala Ser Thr Ser Ser Thr Glu Lys Ser Phe Pro Trp Arg Ser Thr Asp Val Pro Asn Thr Gly Val Ser Lys Pro Arg Val Ser Asp Ala Val Gln Pro Asn Asn Tyr Leu Ile Arg Thr Glu Pro Glu Gln Gly Thr Leu Tyr Ser Pro Glu Gln Thr Ser Leu His Glu Ser Glu Gly Ser Leu Gly Asn Ser Arg Ser Ser Thr Gln Met Asn Ser Tyr Ser Asp Ser Gly Tyr Gln Glu Ala Gly Ser Phe His Asn Ser Gln Asn Val Ser Lys Ala Asp Asn Arg Gln Gln His Ser Phe Ile Gly Ser Thr Asn Asn His Val Val Arg Asn Ser Arg Ala Glu Gly Gln Thr Leu Val Gln Pro Ser Val Ala Asn Arg Ala Met Arg Arg Val Ser Ser Val Pro Ser Arg Ala Gln Ser Pro Ser Tyr Val Ile Ser Thr Gly Val Ser Pro Ser Arg Gly Ser Leu Arg Thr Ser Leu Gly Ser Gly Phe Gly Ser Pro Ser Val Thr Asp Pro Arg Pro Leu Asn Pro Ser Ala Tyr Ser Ser Thr Thr Leu Pro Ala Ala Arg Ala Ala 265 Ser Pro Tyr Ser Gln Arg Pro Ala Ser Pro Thr Ala Ile Arg Arg Ile Gly Ser Val Thr Ser Arg Gln Thr Ser Asn Pro Asn Gly Pro Thr Pro Gln Tyr Gln Thr Thr Ala Arg Val Gly Ser Pro Leu Thr Leu Thr Asp 315 Ala Gln Thr Arg Val Ala Ser Pro Ser Gln Gly Gln Val Gly Ser Ser Ser Pro Lys Arg Ser Gly Met Thr Ala Val Pro Gln His Leu Gly Pro Ser Leu Gln Arg Thr Val His Asp Met Glu Gln Phe Gly Gln Gln Gln 365 Tyr Asp Ile Tyr Glu Arg Met Val Pro Pro Arg Pro Asp Ser Leu Thr Gly Leu Arg Ser Ser Tyr Ala Ser Gln His Ser Gln Leu Gly Gln Asp Leu Arg Ser Ala Val Ser Pro Asp Leu His Ile Thr Pro Ile Tyr Glu Gly Arg Thr Tyr Tyr Ser Pro Val Tyr Arg Ser Pro Asn His Gly Thr Val Glu Leu Gln Gly Ser Gln Thr Ala Leu Tyr Arg Thr Gly Val Ser Gly Ile Gly Asn Leu Gln Arg Thr Ser Ser Gln Arg Ser Thr Leu Thr 450 460 Tyr Gln Arg Asn Asn Tyr Ala Leu Asn Thr Thr Ala Thr Tyr Ala Glu Pro Tyr Arg Pro Ile Gln Tyr Arg Val Gln Glu Cys Asn Tyr Asn Arg Leu Gln His Ala Val Pro Ala Asp Asp Gly Thr Thr Arg Ser Pro Ser Ile Asp Ser Ile Gln Lys Asp Pro Arg Glu Phe Ala Trp Arg Asp Pro 515 Glu Leu Pro Glu Val Ile His Met Leu Glu His Gln Phe Pro Ser Val Gin Ala Asn Ala Ala Ala Tyr Leu Gln His Leu Cys Phe Gly Asp Asn 545 555 560 Lys Val Lys Met Giu Val Cys Arg Leu Gly Gly Ile Lys His Leu Val

Asp Leu Leu Asp His Arg Val Leu Glu Val Gln Lys Asn Ala Cys Gly 580 Ala Leu Arg Asn Leu Val Phe Gly Lys Ser Thr Asp Glu Asn Lys Ile 595Ala Met Lys Asn Val Gly Gly Ile Pro Ala Leu Leu Arg Leu Leu Arg Lys Ser Ile Asp Ala Glu Val Arg Glu Leu Val Thr Gly Val Leu Trp 625 630 630 Asn Leu Ser Ser Cys Asp Ala Val Lys Met Thr Ile Ile Arg Asp Ala 645 655 Leu Ser Thr Leu Thr Asn Thr Val Ile Val Pro His Ser Gly Trp Asn 660Asn Ser Ser Phe Asp Asp Asp His Lys Ile Lys Phe Gln Thr Ser Leu 675 Val Leu Arg Asn Thr Thr Gly Cys Leu Arg Asn Leu Thr Ser Ala Gly Glu Glu Ala Arg Lys Gln Met Arg Ser Cys Glu Gly Leu Val Asp Ser 705 715 720 Leu Leu Tyr Val Ile His Thr Cys Val Asn Thr Ser Asp Tyr Asp Ser 735Lys Thr Val Glu Asn Cys Val Cys Thr Leu Arg Asn Leu Ser Tyr Arg 740Leu Glu Leu Glu Val Pro Gln Ala Arg Leu Leu Gly Leu Asn Glu Leu 755 765 Asp Asp Leu Leu Gly Lys Glu Ser Pro Ser Lys Asp Ser Glu Pro Ser 770 Cys Trp Gly Lys Lys Lys Lys Lys Lys Lys Arg Thr Pro Gln Glu Asp 785 Gln Trp Asp Gly Val Gly Pro Ile Pro Gly Leu Ser Lys Ser Pro Lys Gly Val Glu Met Leu Trp His Pro Ser Val Val Lys Pro Tyr Leu Thr Leu Leu Ala Glu Ser Ser Asn Pro Ala Thr Leu Glu Gly Ser Ala Gly 835 840Ser Leu Gln Asn Leu Ser Ala Ser Asn Trp Lys Phe Ala Ala Tyr Ile 850 860 Arg Gly Gly Arg Pro Lys Arg Lys Gly Leu Pro Ile Leu Val Glu Leu 865 870 880 Leu Arg Met Asp Asp Arg Val Val Ser Ser Gly Ala Thr Ala Leu 885 Arg Asn Met Ala Leu Asp Val Arg Asn Lys Glu Leu Ile Gly Lys Tyr 900 910 Ala Met Arg Asp Leu Val Asn Arg Leu Pro Gly Gly Asn Gly Pro Ser 915 Val Leu Ser Asp Glu Thr Met Ala Ala Ile Cys Cys Ala Leu His Glu 930 940 Val Thr Ser Lys Asn Met Glu Asn Ala Lys Ala Leu Ala Asp Ser Gly 945 950 955 Gly Ile Glu Lys Leu Val Asn Ile Thr Lys Gly Arg Gly Asp Arg Ser Ser Leu Lys Val Val Lys Ala Ala Ala Glm Val Leu Asm Thr Leu Trp 980 985 Gln Tyr Arg Asp Leu Arg Ser Ile Tyr Lys Lys Asp Gly Trp Asn Gln
995 Asn His Phe Ile Thr Pro Val Ser Thr Leu Glu Arg Asp Arg Phe Lys 1010Ser His Pro Ser Leu Ser Thr Thr Asn Gln Gln Met Ser Pro Ile Ile 1025 1030 1035 Gln Ser Val Gly Ser Thr Ser Ser Ser Pro Ala Leu Leu Gly Ile Arg 1045 1050 1055 Asp Pro Arg Ser Glu Tyr Asp Arg Thr Gln Pro Pro Met Gln Tyr Tyr 1060 1070

-110-Asn Ser Gln Gly Asp Ala Thr His Lys Gly Leu Tyr Pro Gly Ser Ser Lys Pro Ser Pro Ile Tyr Ile Ser Ser Tyr Ser Ser Pro Ala Arg Glu 1090 Gln Asn Arg Arg Leu Gln His Gln Gln Leu Tyr Tyr Ser Gln Asp Asp 1105 1110 Ser Asn Arg Lys Asn Phe Asp Ala Tyr Arg Leu Tyr Leu Gln Ser Pro 1125 1130 1135 His Ser Tyr Glu Asp Pro Tyr Phe Asp Asp Arg Val His Phe Pro Ala 1140 Ser Thr Asp Tyr Ser Thr Gln Tyr Gly Leu Lys Ser Thr Thr Asn Tyr 1155 1160 Val Asp Phe Tyr Ser Thr Lys Arg Pro Ser Tyr Arg Ala Glu Gln Tyr 1170 1180 Pro Gly Ser Pro Asp Ser Trp Val Tyr Asp Gln Asp Ala Gln Gln Arg 1185 Asn Ser Phe Phe Leu Thr Leu Phe Arg Leu Arg

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 970 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ix) FEATURE:
 (A) NAME/KEY: misc feature
 (B) LOCATION: 1..970
 (D) OTHER INFORMATION: /note= "Y2H9"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GAATTCCCAC AGATACCACT GCTGCTCCCG CCCTTTCGCT CCTCGGCCGC GCAATGGGCA 60 CCCGCGACGA CGAGTACGAC TACCTCTTTA AAGTTGTCCT TATTGGAGAT TCTGGTGTTG 120 GARAGAGTAA TCTCCTGTCT CGATTTACTC GARATGAGTT TAATCTGGAA AGCAAGAGCA 180 CCATTGGAGT AGAGTTTGCA ACAAGAAGCA TCCAGGTTGA TGGAAAAACA ATAAAGGCAC 240 AGATATGGGA CACAGCAGGG CAAGAGCGAT ATCGAGCTAT AACATCAGCA TATTATCGTG 300 GAGCTGTAGG TGCCTTATTG GTTTATGACA TTGCTAAACA TCTCACATAT GAAAATGTAG 360 AGCGATGGCT GAAAGAACTG AGAGATCATG CTGATAGTAA CATTGTTATC ATGCTTGTGG 420 GCAATAAGAG TGATCTACGT CATCTCAGGG CAGTTCCTAC AGATGAAGCA AGAGCTTTTG 480 CAGAAAAGAA TGGTTTGTCA TTCATTGAAA CTTCGGCCCT AGACTCTACA AATGTAGAAG 540 CTGCTTTTCA GACAATTTTA ACAGAGATTT ACCGCATTGT TTCTCAGAAG CAAATGTCAG 600 ACAGACGCGA AAATGACATG TCTCCAAGCA ACAATGTGGT TCCTATTCAT GTTCCACCAA 660 CCACTGAAAA CAAGCCAAAG GTGCAGTGCT GTCAGAACAT CTAAGGCATT TCTCTTCTCC CCTAGAAGGC TGTGTATAGT CCATTTCCCA GGTCTSASAT TTAAATATAW TTGTAATTCT 780 TGTGTCACTT TTGTGTTTTA TTACTTCATA CTTATGAATT TTTCCATGTC CTAAGTCTTT 840 TGATTTTGMT TTATAAAATC ATCCACTTGT NCCGAATGNC TGCAGCTTTT TTTCATGCTA 900 TGGCTTCACT AGCCTTAGTT TNATAAACTG AATGTTTGGA TTCCTCCCCC CAAAAAAAA 960 AAAACTCGAG 970

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 264 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ix) FEATURE:

 - (A) NAME/KEY: misc feature (B) LOCATION: 1..264 (D) OTHER INFORMATION: /note= "Y2H23b"

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
GAATTCGCGG CCGNGTCGAC CCCCCACCCC CGATGCCACC ACCCCCANTG GGNTCTCCCN	60
NCCCAGTCAT CAGTTCTTCC ATGGNGTNCC CTGGTCTGCC CCCTCCAGCT CCCCCAGGCN	120
TTCTCCGGGT CTGNCAGCAG CCNCCAGATT AACTCAACAG TGTCACTCCC TGGGGGTGGG	180
TCTGGNCCCC CTGANGATGT GAAGCCACCA GTCTNAGNGG TCCGGGGTCT GTACTGTCCA	240
CCCCCTCCAG GTGGACCTGG CGCT	264
(2) INFORMATION FOR SEQ ID NO:9:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 340 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ix) FEATURE: (A) NAME/KEY: misc feature (B) LOCATION: 1340 (D) OTHER INFORMATION: /note= "Y2H27"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
GAATTCGCGG CCGCGTCGAC CGCGGTCGCG TCGACCTGTT GCCCAGGCCC TAGAGGTCAT	60
TCCTCGTACC CTGATCCAGA ACTGTGGGGC CAGCACCATC CGTCTACTTA CCTCCCTTCG	120
GGCCAAGCAC ACCCAGGAGA ACTGTGAGAC CTGGGGTGTA AATGGTGAGA CGGGTACTTT	180
GGTGGACATG AAGGAACTGG GCATATGGGA GCCATTGGCT GTGAAGCTGC AGACTTATAA	240
GACAGCAGTG GAGACGGCAG TICTGCTACT GCGAATTGAT GACATCGTTT CAGGCCACAA	300
AAAGAAAGGC GATGACCAGA GCCGGCAAGG CGGNGCTCCT	340
(2) INFORMATION FOR SEQ ID NO:10:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 404 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ix) FEATURE: (A) NAME/KEY: misc feature (B) LOCATION: 14U4 (D) OTHER INFORMATION: /note= "Y2K35"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
GAATTCGCGG TCGCGTCGAC GGTTAGTCCC ACTGGNCGCA TCGAGGGNTT CACCAACGTC	60
ATGGAGCTGT ATGGCANGAT CGCCGAGGTC TTCCNCCTGC CAACTGCCGA GGTGATGTTC	120
TGCACCCTGA NCACCCACAA AGTGGACATN GACAAGCTCC TGGGGGGGCCA GATCGGGCTG	180
GAGGACTTCA TCTTCGCCCA CGTGAAGGGG YAGCGCAAGG AGGTGGAGGT GTTCAWGTCG	240
GAGGATGYAC TCGGKCTCAC CATCACGGAC AACGGGGCTG GCTACGCTTC CATCAAGCGC	300
ATCAAGGAGG GCAGCGTGAT CGACCACATC CACCTCATCA GCGTGGGCGA CATGATCGAG	3.6.0
GCCATTAACG GGCAGAGCTT CCTGGGCTGC CGGCATTACG AGGT (2) INFORMATION FOR SEQ ID NO:11:	404
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 350 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ix) FEATURE: (A) NAME/KEY: misc feature (B) LOCATION: 1350 (D) OTHER INFORMATION: /note= "Y2H171"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
GAATTCGCGG CCGCGTCGAC AAAAAAAGTA AAAGGAACTC GGCAAATCTT ACCCCGCCTG	60
TTTACCAAAA ACATCACCTC TAGCATCACC AGTATTAGAG GCACCGCCTG CCCAGTGACA	120

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CATGTTTAAC	GGCCGCGGTA	CCCTAACCGT	GCAAAGGTAG	CATAATCACT	TGTTCCTTAA	180
GTAGGGACCT	GTATGAATGG	CTCCACGAGG	GTTCAGCTGT	CTCTTACTTT	TAACCARTGA	240
AATTGACCTG	CCCGTGAAGA	GGCGGGCATG	ACACAGCAAG	ACGAGAAGAC	CCTATGGAGC	300
TTTAATTTAT	TAATGCAAAC	AGTACCTAAC	AAACCCACAG	GGTCCTAAAC		350
(2) INFORM	ATION FOR SI	EQ ID NO:12	:			
151 61	FOURNCE CUNI					

SEQUENCE CHARACTERISTICS:
(A) LENGTH: 350 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:
(A) NAME/KEY: misc feature
(B) LOCATION: 1..350
(D) OTHER INFORMATION: /note= "Y2H41"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GAATTCGCGG	NCGCGTCGAC	AGATAATGAA	AAAACCAGAG	GTTCCCTTCT	TTGGTCCCCT	60
NNNNGATGGT	GCTATTGTGA	ATGGAAAGGT	TCTACCCATT	ATGGTTAGAG	CAACAGCTAT	120
AAATGCAAGC	CGTGCTCTGA	AATCTCTGAT	TCCATTGTAT	CAAAACTTCT	ATGAGGAGAG	180
AGCACGATAC	CTGCAAACAA	TTGTCCAGCA	CCACTTAGAA	CCAACAACAT	TTGAAGATTT	240
TGNAGCACAG	GTTTTTTCTC	CAGCTCCCTA	CCACCATTTA	CCATCTGATG	CCGTTGGCTC	300
CTACCCAGAG	ATTCTACCCA	GTGAAAACTC	CCACAGCAAC	GCAGGTAGGA		350

CLAIMS

What is claimed is:

12) presenilin-interacting domain.

- 1. An isolated nucleic acid comprising a nucleotide sequence encoding at least a presenilin-interacting domain of a presenilin-interacting protein selected from the group consisting of a mammalian S5a (approximately residues 70-377 of SEQ ID NO: 2), GT24 (approximately residues 346-862 of SEQ ID NO: 4), p0071 (approximately residues 509-1022 of SEQ ID NO: 6), Rab11 (SEQ ID NO: 7), retinoid X receptor-β (SEQ ID NO:8), cytoplasmic chaperonin (SEQ ID NO: 9), Y2H35 (SEQ ID NO: 10), Y2H171 (SEQ ID NO: 11), and a Y2H41 (SEQ ID NO:
- An isolated nucleic acid comprising a nucleotide sequence of at least 10 consecutive nucleotides selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, GenBank Accession Numbers F08730, T18858, X81889, X56740, X53143, M84820, X63522, M81766, U17104, X74801, R12984,
- 15 D55326, and T64843, and a sequence complementary to any of these sequences.
 - 3. An isolated nucleic acid as in claim 2 comprising a nucleotide sequence of at least 15 consecutive nucleotides selected from said group.
- 20 4. An isolated nucleic acid as in claim 2 comprising a nucleotide sequence of at least 20 consecutive nucleotides selected from said group.

5. An isolated nucleic acid comprising a nucleotide sequence encoding an antigenic determinant of a presentilin-interacting protein selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.

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6. A method for identifying allelic variants or heterospecific homologues of a human presentilin-interacting protein gene comprising

choosing a nucleic acid probe or primer capable of hybridizing to a human presentilin-interacting protein gene sequence under stringent hybridization conditions:

mixing said probe or primer with a sample of nucleic acids which may contain a nucleic acid corresponding to said variant or homologue;

detecting hybridization of said probe or primer to said nucleic acid corresponding to said variant or homologue.

- 7. A method as in claim 6 wherein said sample comprises a sample of nucleic acids selected from the group consisting of human genomic DNA, human mRNA, and human cDNA.
- A method as in claim 6 wherein said sample comprises a sample of nucleic
 acids selected from the group consisting of mammalian genomic DNA, mammalian mRNA, and mammalian cDNA.
 - 9. A method as in claim 6 wherein said sample comprises a sample of nucleic acids selected from the group consisting of invertebrate genomic DNA, invertebrate mRNA, and invertebrate cDNA.

- 10. A method as in claim 6 further comprising the step of isolating said nucleic acid corresponding to said variant or homologue.
- 5 11. A method as in claim 6 wherein said nucleic acid is identified by hybridization.
 - 12. A method as in claim 6 wherein said nucleic acid is identified by PCR amplification.

- 13. A method for identifying allelic variants or heterospecific homologues of a human presentilin-interacting protein gene comprising
- choosing an antibody capable of selectively binding to a human presentilin-interacting protein;

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mixing said antibody with a sample of proteins which may contain a protein corresponding to said variant or homologue;

detecting binding of said antibody to said protein corresponding to said variant or homologue.

20 14. A method as in claim 13 wherein said sample comprises a sample of proteins selected from the group consisting of human proteins, human fusion proteins, and proteolytic fragments thereof.

- 15. A method as in claim 13 wherein said sample comprises a sample of proteins selected from the group consisting of mammalian proteins, mammalian fusion proteins, and proteolytic fragments thereof.
- 5 16. A method as in claim 13 wherein said sample comprises a sample of proteins selected from the group consisting of invertebrate proteins, invertebrate fusion proteins, and proteolytic fragments thereof.
- 17. A method as in claim 13 further comprising the step of substantially purifying said protein corresponding to said variant or homologue.
 - 18. An isolated nucleic acid comprising an allelic variant or a heterospecific homologue of a human presentilin-interacting protein gene.
- 15 19. An isolated nucleic acid encoding an allelic variant or heterospecific homologue of a human presenilin-interacting protein.
 - 20. An isolated nucleic acid comprising a recombinant vector including a nucleotide sequence of any one of claims 1-19.

21. An isolated nucleic acid as in claim 20 wherein said vector is an expression vector and said presentlin-interacting protein nucleotide sequence is operably joined to a regulatory region.

- 22. An isolated nucleic acid as in claim 21 wherein said expression vector may express said presentilin-interacting protein sequence in mammalian cells.
- An isolated nucleic acid as in claim 22 wherein said cells are selected from the group consisting of fibroblast, liver, kidney, spleen, bone marrow and neurological cells.
- An isolated nucleic acid as in claim 21 wherein said vector is selected from the group consisting of vaccinia virus, adenovirus, retrovirus, neurotropic viruses and Herpes simplex.
- An isolated nucleic acid as in claim 21 wherein said expression vector encodes at least a presenilin-interacting domain of a presenilin-interacting protein
 selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.
- 26. An isolated nucleic acid as in claim 21 wherein said vector further comprises sequences encoding an exogenous protein operably joined to said
 20 presenilin-interacting protein sequence and whereby said vector encodes a presenilin-interacting protein fusion protein.
 - 27. An isolated nucleic acid as in claim 26 wherein said exogenous protein is selected from the group consisting of lacZ, trpE, maltose-binding protein, a poly-His

tag, glutathione-S-transferase, a GAL4-DNA binding domain, and a GAL4 activation domain.

- 28. An isolated nucleic acid comprising a recombinant expression vector including nucleotide sequences corresponding to an endogenous regulatory region of a presentilin-interacting protein gene.
 - 29. An isolated nucleic acid as in claim 28 wherein said endogenous regulatory region is operably joined to a marker gene.

- 30. A host cell transformed with an expression vector of any one of claims 20-29, or a descendant thereof.
- 31. A host cell as in claim 30 wherein said host cell is selected from the group consisting of bacterial cells and yeast cells.
 - 32. A host cell as in claim 30 wherein said host cell is selected from the group consisting of fetal cells, embryonic stem cells, zygotes, gametes, and germ line cells.
- 20 33. A host cell as in claim 30 wherein said cell is selected from the group consisting of fibroblast, liver, kidney, spleen, bone marrow and neurological cells.

- 34. A host cell as in claim 30 wherein said cell is an invertebrate cell.
- 35. A non-human animal model for Alzheimer's Disease, wherein a genome of said animal, or an ancestor thereof, has been modified by at least one recombinant construct, and wherein said recombinant construct has introduced a modification selected from the group consisting of (1) insertion of nucleotide sequences encoding at least a functional domain of a heterospecific normal presenilin-interacting protein, (2) insertion of nucleotide sequences encoding at least a functional domain of a heterospecific mutant presenilin-interacting protein, (3) insertion of nucleotide sequences encoding at least a functional domain of a conspecific homologue of a heterospecific mutant presenilin-interacting protein, and (4) inactivation of an endogenous presenilin-interacting protein gene.
- 36. An animal as in claim 35 wherein said modification is insertion of a
 nucleotide sequence encoding at least a functional domain of a normal human presentilin-interacting protein selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.
- 20 37. An animal as in claim 35 wherein said modification is insertion of a nucleotide sequence encoding at least a functional domain of a mutant human presentilin-interacting protein selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.

- 38. An animal as in claim 35 wherein said animal is selected from the group consisting of rats, mice, hamsters, guinea pigs, rabbits, dogs, cats, goats, sheep, pigs, and non-human primates.
- 5 39. An animal as in claim 35 wherein said animal is an invertebrate.
 - 40. A method for producing at least a functional domain of a presentilininteracting protein comprising culturing a host cell of any of claims 30-34 under suitable conditions to produce said presentilin by expressing said nucleic acid.

- 41. A substantially pure preparation of a protein selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.
- A substantially pure preparation of a polypeptide comprising an amino acid sequence of at least 10 consecutive amino acid residues selected from the group consisting SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, and GenBank Accession Numbers F08730, T18858, X81889, X56740, X53143, M84820, X63522, M81766, U17104, X74801, R12984, D55326, and T64843.

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43. A substantially pure preparation of a polypeptide as in claim 42 comprising an amino acid sequence of at least 15 consecutive amino acid residues selected from said group.

- A substantially pure preparation of a polypeptide comprising at least a presentilin-interacting domain of a presentilin-interacting protein selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.
- 45. A substantially pure preparation of a polypeptide comprising an antigenic determinant of a presentilin-interacting protein selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.
 - 46. A method of producing antibodies which selectively bind to a presentilininteracting protein comprising the steps of
- administering an immunogenically effective amount of a presentilininteracting protein immunogen to an animal;
 - allowing said animal to produce antibodies to said immunogen; and obtaining said antibodies from said animal or from a cell culture derived therefrom.
- A substantially pure preparation of an antibody which selectively binds to an antigenic determinant of a presentilin-interacting protein selected from the group consisting of a mammalian S5a, GT24, p0071, Rab11, retinoid X receptor-β, cytoplasmic chaperonin, Y2H35, Y2H171, and Y2H41 protein.

- 48. A substantially pure preparation of an antibody as in claim 47 wherein said antibody selectively binds to an antigenic determinant of a mutant presentiininteracting protein and fails to bind to a normal presentiin-interacting protein.
- 5 49. A cell line producing an antibody of any one of claims 47-48.
 - 50. A method for identifying compounds which can modulate the expression of a presenilin-interacting protein gene comprising
- contacting a cell with a test candidate wherein said cell includes a

 regulatory region of a presentilin-interacting protein gene operably joined to a coding region; and

detecting a change in expression of said coding region.

- 51. A method as in claim 50 wherein said change comprises a change in a level of an mRNA transcript encoded by said coding region.
 - 52. A method as in claim 50 wherein said change comprises a change in a level of a protein encoded by said coding region.
- 20 53. A method as in claim 50 wherein said change is a result of an activity of a protein encoded by said coding region.

- 54. A method as in claim 50 wherein said coding region encodes a marker protein selected from the group consisting of β -galactosidase, alkaline phosphatase, green fluorescent protein, and luciferase.
- 5 55. A method for identifying compounds which can selectively bind to a presentilin-interacting protein comprising the steps of

providing a preparation including at least one presentlin-interacting protein component;

contacting said preparation with a sample including at least one candidate compound; and

detecting binding of said presenilin-interacting protein component to said candidate compound.

- 56. The method in 55 wherein said binding to said presenilin-interacting component is detected by an assay selected from the group consisting of: affinity chromatography, co-immunoprecipitation, a Biomolecular Interaction Assay, and a yeast two-hybrid system.
- 57. A method of identifying compounds which can modulate activity of a presenilin-interacting protein comprising the steps of

providing a cell expressing a normal or mutant presentiin-interacting protein gene;

contacting said cell with at least one candidate compound; and detecting a change in a marker of said activity.

58. A method as in claim 57 wherein measurement of said marker indicates a difference between cells bearing an expressed mutant presentilin-interacting protein gene and otherwise identical cells free of an expressed mutant presentilin-interacting protein gene.

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59. A method as in claim 57 wherein said change comprises a change in a non-specific marker of cell physiology selected from the group consisting of pH; intracellular Ca⁺⁺, Na⁺, or K⁺; cyclic AMP levels; GTP/GDP ratios; phosphatidylinositol activity; and protein phosphorylation.

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- 60. A method as in claim 57 wherein said change comprises a change in expression of said presentlin-interacting protein.
- 61. A method as in claim 57 wherein said change comprises a change in intracellular concentration or flux of an ion selected from the group consisting of Ca²⁺, Na⁺ and K⁺.
 - 62. A method as in claim 57 wherein said change comprises a change in occurrence or rate of apoptosis or cell death.

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63. A method as in claim 57 wherein said change comprises a change in production of $A\beta$ peptides.

- 64. A method as in claim 57 wherein said change comprises a change in phosphorylation of at least one microtubule associated protein.
- 65. A method as in claim 57 wherein said cell is a cell cultured in vitro.

- 66. A method as in claim 65 wherein said cell is a transformed host cell of any one of claims 30-34.
- 67. A method as in claim 65 wherein said cell is explanted from a host bearing at least one mutant presentilin-interacting protein gene.
 - 68. A method as in claim 65 wherein said cell is explanted from a transgenic animal of any one of claims 35-39.
- 15 69. A method as in claim 57 wherein said cell is a cell in a live animal.
 - 70. A method as in claim 69 wherein said cell is a cell of a transgenic animal of any one of claims 35-39.
- 20 71. A method as in claim 57 wherein said cell is in a human subject in a clinical trial.

72. A diagnostic method for determining if a subject bears a mutant presentilininteracting protein gene comprising the steps of

providing a biological sample of said subject;

- detecting in said sample a mutant presentilin-interacting protein nucleic acid, a mutant presentilin-interacting protein, or a mutant presentilin-interacting protein activity.
- 73. A method as in claim 72, wherein a mutant presentilin-interacting protein nucleic acid is detected by an assay selected from the group consisting of direct nucleotide sequencing, probe specific hybridization, restriction enzyme digest and mapping, PCR mapping, ligase-mediated PCR detection, RNase protection, electrophoretic mobility shift detection, and chemical mismatch cleavage.
- 74. A method as in claim 72, wherein a mutant presentilin-interacting protein is detected by an assay selected from the group consisting of an immunoassay, a protease assay, and an electrophoretic mobility assay.
- 75. A pharmaceutical preparation comprising a substantially pure presentilin-20 interacting protein and a pharmaceutically acceptable carrier.
 - 76. A pharmaceutical preparation comprising an expression vector operably encoding a presentilin-interacting protein, wherein said expression vector may express said presentilin-interacting protein in a human subject, and a pharmaceutically acceptable carrier.

- 77. A pharmaceutical preparation comprising an expression vector operably encoding a presentilin-interacting protein antisense sequence, wherein said expression vector may express said presentilin-interacting protein antisense sequence in a human subject, and a pharmaceutically acceptable carrier.
- 78. A pharmaceutical preparation comprising a substantially pure antibody, wherein said antibody selectively binds to a mutant presentilin-interacting protein, and a pharmaceutically acceptable carrier.

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- A pharmaceutical preparation as in claim 78 wherein said preparation is essentially free of an antibody which selectively binds a normal presentilin-interacting protein.
- 15 80. A pharmaceutical preparation comprising a substantially pure preparation of an antigenic determinant of a mutant presentilin-interacting protein.
 - 81. A pharmaceutical preparation as in claim 80 wherein said preparation is essentially free of an antigenic determinant of a normal presentilin-interacting protein.

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82. A method of treatment for a patient bearing a mutant presentilin-interacting protein gene comprising the step of administering to said patient a therapeutically effective amount of the pharmaceutical preparation of any one of claims 75-81.

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83. A method as in claim 82, wherein said pharmaceutical preparation is targeted to a cell type is selected from the group consisting of heart, brain, lung, liver, skeletal muscle, kidney, pancreas and neurological cells.

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C. DOCU	MENTS CONSIDERED TO BE RELEVANT	,		
Category *	Citation of document, with indication, where appropriate, of	the relevant p	sassages	Relevant to claim No.
X	THE JOURNAL OF BIOLOGICAL CHEM vol. 270, no. 35, 1 September pages 20615-20620, XP002029380 JOHANSSON, E., ET AL. ON: "M CLONING AND EXPRESSION OF A PI GLAND PROTEIN MODULATING INTES SECRETION" cited in the application see the whole document	1995, OLECULA TUITARY TINAL F		1-12, 18-20, 30,31, 34, 41-47,49
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X Furth	er documents are listed in the continuation of box C.	П	Patent family members	ire listed in annex.
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT Category Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No.						
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